

**Committee for Risk Assessment  
RAC**

Annex 1  
**Background document**  
to the Opinion proposing harmonised classification  
and labelling at EU level of

**Cumene**

**EC Number: 202-704-5**

**CAS Number: 98-82-8**

CLH-O-0000006849-56-01/F

The background document is a compilation of information considered relevant by the dossier submitter or by RAC for the proposed classification. It includes the proposal of the dossier submitter and the conclusion of RAC. It is based on the official CLH report submitted to public consultation. RAC has not changed the text of this CLH report but inserted text which is specifically marked as 'RAC evaluation'. Only the RAC text reflects the view of RAC.

**Adopted  
17 September 2020**



## **CLH report**

### **Proposal for Harmonised Classification and Labelling**

**Based on Regulation (EC) No 1272/2008 (CLP Regulation),  
Annex VI, Part 2**

#### **International Chemical Identification:**

##### **Cumene**

**EC Number: 202-704-5**  
**CAS Number: 98-82-8**  
**Index Number: 601-024-00-X**

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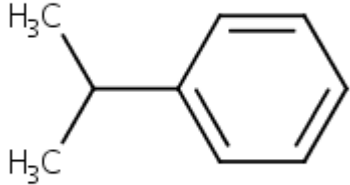
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## 1 IDENTITY OF THE SUBSTANCE

### 1.1 Name and other identifiers of the substance

**Table 1: Substance identity and information related to molecular and structural formula of the substance**

<b>Name(s) in the IUPAC nomenclature or other international chemical name(s)</b>	<b>Cumene</b>
<b>Other names (usual name, trade name, abbreviation)</b>	(1-methylethyl)benzene, isopropylbenzene, propan-2-ylbenzene
<b>ISO common name (if available and appropriate)</b>	Not applicable
<b>EC number (if available and appropriate)</b>	202-704-5
<b>EC name (if available and appropriate)</b>	Cumene
<b>CAS number (if available)</b>	98-82-8
<b>Other identity code (if available)</b>	-
<b>Molecular formula</b>	C <sub>9</sub> H <sub>12</sub>
<b>Structural formula</b>	 <p>The structural formula shows a benzene ring (a hexagon with three alternating double bonds) connected to a central carbon atom. This central carbon atom is also bonded to two other carbon atoms, each of which is bonded to three hydrogen atoms (represented as H<sub>3</sub>C), forming an isopropyl group.</p>
<b>SMILES notation (if available)</b>	CC(C)C1=CC=CC=C1
<b>Molecular weight or molecular weight range</b>	120.194 g/mol
<b>Information on optical activity and typical ratio of (stereo) isomers (if applicable and appropriate)</b>	-
<b>Description of the manufacturing process and identity of the source (for UVCB substances only)</b>	-
<b>Degree of purity (%) (if relevant for the entry in Annex VI)</b>	≥ 80 wt %

## 1.2 Composition of the substance

**Table 2: Constituents (non-confidential information)**

Constituent (Name and numerical identifier)	Concentration range (% w/w minimum and maximum in multi-constituent substances)	Current CLH in Annex VI Table 3.1 (CLP)	Current self-classification and labelling (CLP)
Cumene	≥ 80 wt % (mono-constituent)	Flam. Liq. 3 (H226), Asp. Tox. 1 (H304), STOT SE 3 (H335), Aquatic Chronic 2 (H411)	Registrants report the harmonised classification. In addition a self-classification is reported: Flam. Liq. 3 (H226), Asp. Tox. 1 (H304), STOT SE 3 (H335), Aquatic Chronic 3 (H413)  42 additional notifications (2152 notifiers, 14/3/2018) are available, see below this table.

The following C&L inventory information for self classification for **physicochemical and human health endpoints** is available for the general entry of cumene (CAS no: 98-82-8 on 14/3/2018)

Classification	Number of notifiers
Not classified	1
Flam Liq. 3 - H226	2149
Asp Tox. 1 – H304	2149
STOT SE 3 - H335, H370	2136
Acute Tox 4 – H332, 302	697
Skin Irrit. 2 – H315	4
Eye Irrit. 2 – H319	5
STOT RE 1 – H372	1

**Table 3: Impurities (non-confidential information) if relevant for the classification of the substance**

Impurity (Name and numerical identifier)	Concentration range (% w/w minimum and maximum)	Current CLH in Annex VI Table 3.1 (CLP)	Current self-classification and labelling (CLP)	The impurity contributes to the classification and labelling
No data available				

**Table 4: Additives (non-confidential information) if relevant for the classification of the substance**

Additive (Name and numerical identifier)	Function	Concentration range (% w/w minimum and maximum)	Current CLH in Annex VI Table 3.1 (CLP)	Current self-classification and labelling (CLP)	The additive contributes to the classification and labelling
No relevant additives					

**Table 5: Test substances (non-confidential information) (this table is optional)**

Identification of test substance	Purity	Impurities and additives (identity, %, classification if available)	Other information	The study(ies) in which the test substance is used
No data available				



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**2 PROPOSED HARMONISED CLASSIFICATION AND LABELLING**

**2.1 Proposed harmonised classification and labelling according to the CLP criteria**

**Table 6:**

	Index No	International Chemical Identification	EC No	CAS No	Classification		Labelling			Specific Conc. Limits, M-factors	Notes
					Hazard Class and Category Code(s)	Hazard statement Code(s)	Pictogram, Signal Word Code(s)	Hazard statement Code(s)	Suppl. Hazard statement Code(s)		
Current Annex VI entry	601-024-00-X	cumene	202-704-5	98-82-8	Flam. Liq. 3 Asp. Tox. 1 STOT SE 3  Aquatic Chronic 2	H226 H304 H335 H411	GHS02 GHS07 GHS08 GHS09 Dgr	H226 H304 H335 H411	-	-	Note C
Dossier submitters proposal	601-024-00-X	cumene	202-704-5	98-82-8	<b>Add</b> Carc. 2	<b>Add</b> H351		<b>Add</b> H351			
Resulting Annex VI entry if agreed by RAC and COM	601-024-00-X	cumene	202-704-5	98-82-8	Flam. Liq. 3 Carc. 2 Asp. Tox. 1 STOT SE 3 Aquatic Chronic 2	H226 H351 H304 H335 H411	GHS02 GHS07 GHS08 GHS09 Dgr	H226 H351 H304 H335 H411			Note C

**Table 7: Reason for not proposing harmonised classification and status under public consultation**

<b>Hazard class</b>	<b>Reason for no classification</b>	<b>Within the scope of public consultation</b>
<b>Explosives</b>	hazard class not assessed in this dossier	No
<b>Flammable gases (including chemically unstable gases)</b>	hazard class not assessed in this dossier	No
<b>Oxidising gases</b>	hazard class not assessed in this dossier	No
<b>Gases under pressure</b>	hazard class not assessed in this dossier	No
<b>Flammable liquids</b>	hazard class not assessed in this dossier	No
<b>Flammable solids</b>	hazard class not assessed in this dossier	No
<b>Self-reactive substances</b>	hazard class not assessed in this dossier	No
<b>Pyrophoric liquids</b>	hazard class not assessed in this dossier	No
<b>Pyrophoric solids</b>	hazard class not assessed in this dossier	No
<b>Self-heating substances</b>	hazard class not assessed in this dossier	No
<b>Substances which in contact with water emit flammable gases</b>	hazard class not assessed in this dossier	No
<b>Oxidising liquids</b>	hazard class not assessed in this dossier	No
<b>Oxidising solids</b>	hazard class not assessed in this dossier	No
<b>Organic peroxides</b>	hazard class not assessed in this dossier	No
<b>Corrosive to metals</b>	hazard class not assessed in this dossier	No
<b>Acute toxicity via oral route</b>	hazard class not assessed in this dossier	No
<b>Acute toxicity via dermal route</b>	hazard class not assessed in this dossier	No
<b>Acute toxicity via inhalation route</b>	hazard class not assessed in this dossier	No
<b>Skin corrosion/irritation</b>	hazard class not assessed in this dossier	No
<b>Serious eye damage/eye irritation</b>	hazard class not assessed in this dossier	No
<b>Respiratory sensitisation</b>	hazard class not assessed in this dossier	No
<b>Skin sensitisation</b>	hazard class not assessed in this dossier	No
<b>Germ cell mutagenicity</b>	data conclusive but not sufficient for classification	Yes
<b>Carcinogenicity</b>	harmonised classification proposed	Yes
<b>Reproductive toxicity</b>	data conclusive but not sufficient for classification	Yes
<b>Specific target organ toxicity-single exposure</b>	hazard class not assessed in this dossier	No
<b>Specific target organ toxicity-repeated exposure</b>	hazard class not assessed in this dossier	No
<b>Aspiration hazard</b>	hazard class not assessed in this dossier	No
<b>Hazardous to the aquatic environment</b>	hazard class not assessed in this dossier	No
<b>Hazardous to the ozone layer</b>	hazard class not assessed in this dossier	No

### 3 HISTORY OF THE PREVIOUS CLASSIFICATION AND LABELLING

Current (harmonised) classification on cumene includes Hazard statements H226, H304, H335 and H411. These Hazard statements correspond to earlier classifications under the Directive on Dangerous Substances (67/548/EEC; 25<sup>th</sup> ATP) with risk phrase R10 (flammable) corresponding to H226, R65 (Harmful: may cause lung damage if swallowed) to H304, R37 (Irritating to respiratory system) to H335 and R51/53 (Toxic to aquatic organisms/May cause long-term adverse effects in the aquatic environment) to H411.

#### RAC general comment

Cumene is an alkylbenzene mainly used as an intermediate for the production of phenol and acetone. In addition, the substance is a minor constituent of gasoline and solvents. The proposal from the dossier submitter (DS) addressed the following endpoints: mutagenicity, carcinogenicity and toxicity to the reproduction.

The substance has an existing Annex VI entry to CLP (assessed under Directive 67/548/EEC; 25<sup>th</sup> ATP). The current entry (621-024-00-X) is for cumene (isopropylbenzene) (EC 202-704-5 [1]; CAS 98-82-8 [2]) and propylbenzene (n-propylbenzene) (203-132-9 [2]; 103-65-1 [2]). The CLH dossier is only on cumene and does not include propylbenzene. A new entry (in addition to the existing one) will thus be created keeping the existing hazard classifications not under discussion in the DS proposal. Note C "*Some organic substances may be marketed either in a specific isomeric form or as a mixture of several isomers*" will be removed from the entry of cumene and propylbenzene.

Following inhalation, cumene is readily absorbed and extensively metabolised by cytochrome P450 enzymes. In both human and experimental animals, the oxidation of the side chain of cumene to 2-phenyl-2-propanol is a key step (both of these compounds are found in human and in animal urine). Other quantitatively less important metabolic pathways observed in mice or rats includes reactive metabolites in animals such as  $\alpha$ -methyl styrene (side chain oxidation of 2-phenyl-2-propanol) which was observed in expired air of mice (at trace level in rats). This metabolite may be further oxidised to  $\alpha$ -methyl styrene oxide. In addition, 2-(2-hydroxy-2-propyl)phenylsulfate and 4-(2-hydroxy-2-propyl)phenylsulfate (ring oxidation) were found in the urine of mice and rats. Some studies suggested that metabolism of cumene was more efficient in mice than in rats. These oxidized metabolites are primarily excreted as sulfate or glucuronide conjugates.

### 4 JUSTIFICATION THAT ACTION IS NEEDED AT COMMUNITY LEVEL

**There is no requirement for justification that action is needed at Community level, because this dossier only addresses hazard classes which shall normally subject to harmonized classification and labelling (Article 36 (1) of the CLP Regulation).**

### 5 IDENTIFIED USES

Cumene is mainly used as an intermediate (approximately 95 %) for the production of phenol and acetone. In addition, the substance is a minor constituent of gasolines and solvents. Cumene is also used in the synthesis

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of alpha-methylstyrene, acetophenone and detergents, the manufacture of di-isopropylbenzene and as a catalyst for acrylic polyester-type resins. It can also be found as an isomer in the general C9 aromatic hydrocarbon content of solvents, especially in those used in the printing industry (ECB, 2001).

### 6 DATA SOURCES

This assessment is based on original study reports for each of the toxicological endpoints discussed (see specific Section for reference) and on most recent reviews and assessments, i.e., ACGIH (2017), DFG (2016), IARC (2013), NTP (2013), SCOEL (2015). In addition, relevant earlier assessments were considered for comparison (ECB, 2001; US EPA, 1997; WHO, 1999). ([ECB, 2001](#); [US EPA, 1997](#); [WHO, 1999](#)). The Klimisch criteria were used in each case for the reliability assessment. The database has substantially changed since publication of the NTP technical report on the toxicological and carcinogenesis studies of cumene (NTP, 2009), as evident from some more recent key studies (e.g., Chen *et al.*, 2011; NTP, 2012)

### 7 PHYSICOCHEMICAL PROPERTIES

**Table 8: Summary of physicochemical properties**

Property	Value	Reference	Comment (e.g. measured or estimated)
<b>Physical state at 20°C and 101,3 kPa</b>	Colourless liquid with strong aromatic odour	(ECB, 2001)	
<b>Melting/freezing point</b>	-96 °C at 1013 h Pa	(ECB, 2001)	
<b>Boiling point</b>	152.7 °C at 1013 hPa	(ECB, 2001)	
<b>Relative density</b>	0.86 at 20 °C/4°C	(ECB, 2001)	
<b>Vapour pressure</b>	4.96 hPa at 20°C	(ECB, 2001)	Extrapolation; based on eight experimental data which fit well a linear regression (correlation coefficient = 0.999)
<b>Surface tension</b>	27.5 nN/m at 20 °C	(ECB, 2001)	Estimated
<b>Water solubility</b>	50 mg/L at 25 °C	(ECB, 2001)	Practically insoluble in water, soluble in ethanol and organic solvents
<b>Partition coefficient n-octanol/water</b>	3.55 at 23 °C	(ECB, 2001)	Measured, OECD 107
<b>Flash point</b>	31 °C (closed cup) 39 °C (closed cup)	(ECB, 2001)	
<b>Flammability</b>	0.9% in volume (LEL) 6.5% in volume (UEL)	(ECB, 2001)	Measured
<b>Explosive properties</b>	Explosive under influence of a flame	(ECB, 2001)	
<b>Self-ignition temperature</b>	424 °C at 1010 hPa	(ECB, 2001)	
<b>Oxidising properties</b>	None	(ECB, 2001)	
<b>Granulometry</b>	Not applicable		
<b>Stability in organic solvents and identity of relevant degradation products</b>	No data		
<b>Dissociation constant</b>	No data		
<b>Viscosity</b>	0.73 x 10 <sup>-6</sup> m <sup>2</sup> /s	(ECB, 2001)	

## 8 EVALUATION OF PHYSICAL HAZARDS

Not performed for this substance.

## 9 TOXICOKINETICS (ABSORPTION, METABOLISM, DISTRIBUTION AND ELIMINATION)

**Table 9: Summary table of toxicokinetic studies**

ADME endpoint	Species, Test conditions	Results	Reference
Absorption	<b>Humans, laboratory animals</b> (summary statement)	Cumene is readily absorbed following inhalation exposure in humans and after inhalation, oral or dermal exposure in laboratory animals	(NTP, 2013)
	<b>Human data</b> , inhalation	Mean* 50% (45% - 64%) retention in the respiratory tract, declining with exposure duration (*not specified: arithmetic or geometric)	(Senczuk und Litewka, 1976); (Brugnone <i>et al.</i> , 1989)
	<b>Human data</b> , inhalation, chemical workers (no direct occupational exposure to cumene)	Blood concentrations of cumene $\approx$ 40 times higher than in alveolar air, supporting the blood/air partition coefficient of 37 for cumene	(Brugnone <i>et al.</i> , 1989; NTP, 2013)
	<b>Animal data</b> , F344 rats; ♀, ♂, different routes of administration (oral and inhalation)	Readily absorbed from stomach and from lungs, after inhalation exposure cumene could be detected in blood within 5 min. In gavage studies in rats, maximum blood levels were reached within 4 h after a dose of 33 mg/kg or 8 to 16 h after a dose of 1350 mg/kg was applied.	(Research Triangle Institute, 1989) and (NTP, 2013)
	Rats, rabbits, dermal	Percutaneous absorption observed	(NTP, 2013)
	Animal studies, percutaneous flux with ethylbenzene	Ability of alkylbenzenes to penetrate the skin is significant	(DFG, 2016)
Distribution	<b>Animal data</b> , F344 rats; ♀, ♂, inhalation	Generally concentrations in the tissues are low since >90% were excreted Adipose tissues were observed to have slightly elevated concentrations at all doses, followed by liver and kidney. Inhalation studies in rats have reported half-lives of cumene disappearance from blood as 3.9 to 6.6 hours. There was no evidence of cumene accumulation in tissues following high or repeated oral doses in rats or mice.	(Research Triangle Institute, 1989) and (NTP, 2013)
	<b>Animal data</b> , F344 rats; ♀, ♂, gavage	Generally concentrations in the tissues are low as >90% were excreted. After exposure to a single dose of 33 mg/kg bw elevated levels of cumene were found in liver, kidney and adipose tissue. However, concentration was very low (<0.5% of total radioactivity). The findings after a multiple dosing of 33 mg/kg bw were similar to that after a single exposure. There is no indication that cumene or its	(Research Triangle Institute, 1989)

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ADME endpoint	Species, Test conditions	Results	Reference			
		radioactive metabolites accumulate in any tissue.				
	Rats, mice; gavage (radiolabelled <sup>14</sup> C)	<p>Single Exposure:</p> <p>a) Tissue retention Less than 3% tissue retention after 24 hrs. for rats; less than 1% for mice, (all tissues excluding stomach and intestines)</p> <p>b) Tissue concentration (mice) Similar for ♀ and ♂ mice, low dose (10 mg/kg); higher in ♀ mice at high dose (1000 mg/kg)</p> <p><u>c) Tissue concentrations, (relative, rats vs. mice)</u> single exposure, higher in rats than in ♀ and ♂ mice, particularly in the kidneys</p> <p>d) Relevant tissues (rats vs. mice) Highest tissue concentrations in liver, kidney, lung (mice), or in adipose tissue, liver, kidney (rat)</p> <p>Repeated Exposure (only ♂rats, ♀ mice tested) After seven consecutive daily doses in mice: highest tissue concentrations in lungs of mice and in the kidney of rats Time-dependent <sup>14</sup>C accumulation in respective tissues ; Higher tissue concentrations in rat kidney and mouse lung studies correlate with higher incidence of tumours in these studies (see Section 10.7) Repeat dosing accumulation in liver, kidney, lung, as well as in blood, brain, heart, muscle, and spleen (only in mice)</p>	(NTP, 2013), (Chen <i>et al.</i> , 2011)			
	Rats, inhalation, up to 150 days	Distribution to endocrine organs, central nervous system, bone marrow, spleen, liver	(WHO, 1999)			
Metabolism	<b>Human data</b> , volunteers exposed to cumene vapour for 8 hrs.	2-phenyl 2-propanol [M14] or conjugates (35% of absorbed dose in urine, 48 hrs. after exposure)	(Senczuk and Litewka, 1976)			
	<b>Animal data</b> , ♂- rat, ♂-, ♀- mice, gavage, <sup>14</sup> C radiolabelled (ring)	Cumene metabolites, urine, oral exposure; ranges after single exposure to 1.4-140 mg/kg (rats) or 10-1000 mg/kg (mice)	(Chen <i>et al.</i> , 2011; NTP, 2013)			
		Substance [Id-Nr.]		% of radiolabelled peaks		
				♂ rat	mouse	
					♂	♀
[M1] unknown	N.D.	N.D.-trace	1.8-3.0			
[M2] 2-(2-hydroxy-2-propyl) phenylsulfate	trace	N.D.-trace	N.D.-4.4			
[M3] 4-(2-hydroxy-2-propyl) phenylsulfate	7-11.4	N.D.	N.D.-trace			

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ADME endpoint	Species, Test conditions	Results				Reference
		[M4] unknown	5.2-5.6	N.D.	N.D.-trace	
		[M5] 2-hydroxy-2-phenylpropylsulfate	2.2-2.6	3-8.4	5.8-19.1	
		[M6] 2-phenyl-1,2-propandiol-2-glucuronide	N.D.-1.6	2.9-4-4	2.5-4.2	
		[M7] 2-phenyl-1,2-propandiol-1-glucuronide	17.8-20.1	8.6-16.9	6.1-16.5	
		[M8] 2-hydroxy-2-phenylpropionic acid	12.1-16.4	12.8-15.7	11.4-20.4	
		[M9] 2-phenyl-2-propanol glucuronide	38.1-48.4 <sup>a</sup>	33.5-42.8	29.8-36.8	
		[M10] 2-phenylpropionyl glucuronide	b	N.D.	N.D.	
		[M11] 2-phenylpropionyl glycine	N.D.	5.1-11	2.8-3.7	
		[M12] S-(2-hydroxy-2-phenylpropyl)-N-acetylcysteine	4-4.9 <sup>c</sup>	trace	trace	
		[M13] 2-phenyl-1-propanol glucuronide	4-4.9 <sup>c</sup>	1.6-5.8 <sup>c</sup>	1.5-2.3 <sup>c</sup>	
		[M14] 2-phenyl-2-propanol	Trace-1.8	N.D.-1.5	N.D.	
		[M15] 2-phenyl-1-propanol	N.D.	N.D.-1.6	N.D.	
		[M16] 2-phenylpropionic acid	Trace-2.1	N.D.-trace	N.D.-trace	
		N.D.=not detected; a= total M9 plus M10; b=M10 reported as minor metabolite that co-eluted with M9; c= total of M12 plus M13				
	<b>Rats</b> , (F344); ♀, ♂, inhalation and oral	The major metabolite of urinary excretion (50% and more) was 2-phenyl-2-propanol and its glucuronide and/or sulphate conjugates				(Research Triangle Institute, 1989)
	<b>Rats</b> (F344) single i.v.	Very similar across all routes of application; >50% 2-phenyl-2-propanol and glucuronides or sulfate conjugates; unknown metabolite: possibly “a carboxylic acid metabolite of cumene”				(US EPA, 1997)
	<b>Rats</b> (F344) single inhalation					
	<b>Rats</b> (F344) single gavage					
	<b>Rats</b> (F344) repeated gavage					
	<b>Rats, mice</b> ; oral exposure	More efficient metabolism suggested in mice compared to rats				(NTP, 2013)
	<b>Rats, mice</b> : radiolabelled Cumene, expired air analysis	In expired air: Cumene more than 95% of radiolabelled VOC, α-methylstyrene: 3-4% (mice), trace (rats)				(Chen <i>et al.</i> , 2011)
	<i>In vitro</i> , rabbits, liver and lung	Highest metabolic rate for cumene in lung and liver, compared to other aromatic (and other) solvents				(Sato und Nakajima, 1987)
	<i>In vitro</i> , rat, mice liver and lung microsomes	α-Methylstyrene, 2-phenyl-2-propanol [M14], ] 2-phenyl-1-propanol [M15]; mouse lung microsomes metabolised more cumene than microsomes from mouse liver, rat lung, or rat liver				(Chen <i>et al.</i> , 2011)

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ADME endpoint	Species, Test conditions	Results	Reference
Excretion	<b>Human data</b> , controlled exposure study, 10 individuals, inhalation exposure to 3 concentrations: 240, 480, 720 mg/m <sup>3</sup> , each for 8 hrs.)	Biphasic excretion of 2-phenyl 2-propanol [M14], initial excretion half-life 2 hrs, subsequent (post-exposure) half-life 10 hrs., approached zero after 48 hrs in urine	(Senczuk und Litewka, 1976)
	Human data, samples collected from healthy volunteers (from urban population and an U.S. Air force educational institution)	Some cumene eliminated in expired air (no details provided)	(NTP, 2013)
	<b>Rats</b> , (F344); ♀, ♂, inhalation	Urine was the major route of elimination (76.2 to 93.2%). Excretion was rapid with the majority of cumene being excreted within 24 h (78.6 to 84.6%) and after 72 h nearly complete excretion was observed (96.0 to 98.9%).	(Research Triangle Institute, 1989)
	<b>Rats</b> , (F344); ♀, ♂, oral	Urine was the predominant route of excretion. At 33 mg/kg bw after 8 h about 40% were excreted via urine. This value increased up to 90% after 72 h. A comparable course of excretion was also observed after a multiple exposure of 33 mg/kg bw. After an exposure to a high concentration of 1350 mg/kg bw excretion via urine was delayed. After 72 h excretion via urine was about 75%. Excretion via volatile breath was the predominant excretion pathway during the first hours after exposure accounting for 6 to 7% after 8 h compared to 3 to 4% excreted via urine at this time.	(Research Triangle Institute, 1989)
	<b>Animal data</b> , rats, inhalation	Half-lives, disappearance from blood, 3.9-6.6 hrs.	(NTP, 2013)
	<b>Rats</b> , gavage	Half-lives, disappearance from blood, 9-16 hrs.	(NTP, 2013)
	<b>Rats, mice</b> , oral	No evidence of accumulation	(NTP, 2013)
	Rats, mice, all routes of administration	Excreted in urine (70-90%), in feces (1-5.3%), expired air (radiolabelled VOC) (<1% - 22%); At higher doses: higher excretion via expired air ♀ mice showed higher excretion than ♂ mice via expired air	(NTP, 2013)
	Rabbits, oral	90% recovered as metabolites in urine within 24hrs.	(WHO, 1999)
	Rats, gavage	Practically no radioactivity eliminated in form of <sup>14</sup> CO <sub>2</sub>	(Chen <i>et al.</i> , 2011; DFG, 2016)
	Rats, mice; oral administration	Minor difference between single or repeated exposure pattern in excretion	(NTP, 2013)
	Rats; i.v. administration	Enterohepatic circulation of cumene glucuronide metabolites implied Elimination half-life 8.6 hrs. for ♂, 7.3 hrs. for ♀-rats	(Chen <i>et al.</i> , 2011)



### 9.1 Short summary and overall relevance of the provided toxicokinetic information on the proposed classification(s)

Cumene is readily absorbed following inhalation exposure in humans and after inhalation, oral or dermal exposure in experimental animals. From rodent studies it can be concluded that cumene is widely distributed in the body, extensively metabolised and rapidly excreted, primarily in the urine (NTP, 2013; Research Triangle Institute, 1989). Most findings on metabolism of cumene are based on studies by Chen *et al.* (2011), as shown in **Figure 9-1**, with some details on quantitative comparative data for mice vs. rats documented in Table 9.

It can be concluded that oxidation to metabolite 2-phenyl-2-propanol [M14] is a key step, both in humans and in experimental animals, and that excretion in rats and in mice primarily includes metabolites 2-phenyl-1,2-propanediol 1-glucuronide [M7], 2-hydroxy-2-phenylpropionic acid [M8] and 2-phenyl-2-propanol glucuronide [M9]. Other metabolic pathways are quantitatively less relevant, but include reactive metabolites:

- One pathway includes side-chain oxidation of [M14] to  $\alpha$ -methyl styrene (AMS). AMS induced carcinogenic effects in experimental animals (NTP, 2007). AMS probably is further oxidised to AMS-oxide, a substance, which has been demonstrated to be mutagenic in *S. typh.* (see Section 10.6).
- Another metabolic pathway is ring oxidation, with two metabolites, 2-(2-hydroxy-2-propyl) phenylsulfate [M2] and 4-(2-hydroxy-2-propyl) phenylsulfate [M3], found *in vivo* after cumene exposure in the urine of mice and rats. However, the aggregated amount of those two metabolites [M2, M3] is limited with 7-11.4% in rats and up to 4.4% in mice (% of total  $^{14}\text{C}$  recovered). Because of the postulated intermediates (arene oxides, catechol and quinonemethide) this metabolic pathway is regarded as potentially relevant for adverse health effect: For example, for another alkylated benzene substance, styrene, ring hydroxylation is associated with mouse specific metabolism in lung Clara cells, leading to cytotoxicity, Clara cell destruction and lung tumours in male mice (Cruzan *et al.*, 2012).

Chen *et al.* (2011) demonstrated accumulation of  $^{14}\text{C}$  radioactivity in the female mouse lung after repeated oral cumene exposure, in contrast to male rats, where no accumulation in the lung was observed. However, the authors did not link the recorded radioactivity to a certain metabolic pathway, did not identify specific metabolites or specific responsible enzymes (within CYP subfamily, critical for metabolism in the respiratory tract) and thus do not allow further insight into the toxicokinetics in the respiratory tract of mice or rats (further discussion in Section 10.7.1). In the study by Research Triangle Institute (1989) rats were exposed via inhalation or gavage against radiolabelled cumene. The authors did not show an accumulation of  $^{14}\text{C}$  in the lung. Adipose tissues were observed to have slightly elevated concentrations at all doses, followed by liver and kidney.

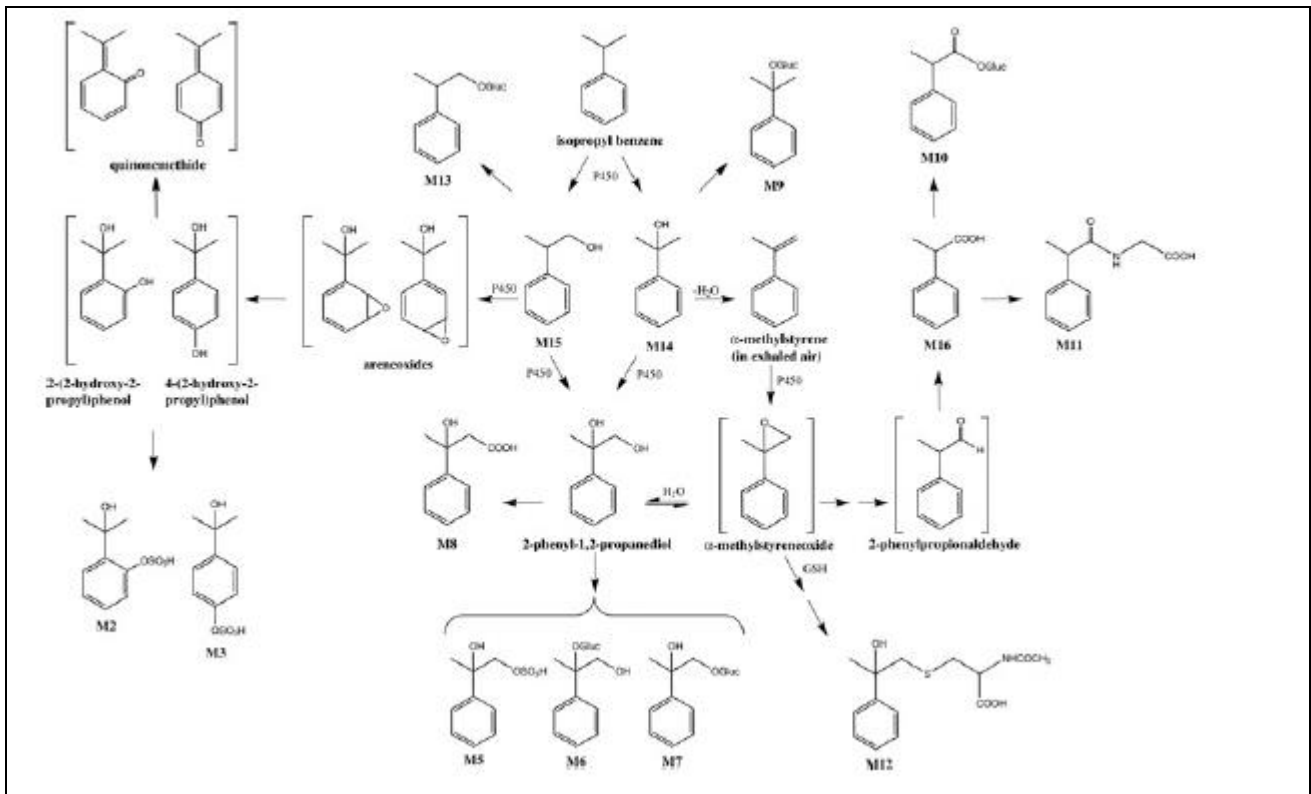


Figure 9-1: Detected and postulated metabolites of cumene adopted from DFG (2016), as modified from Chen *et al.* (2011) (mainly based on data from rodents, but assumed to be applicable to humans)

## 10 EVALUATION OF HEALTH HAZARDS

### 10.1 Acute toxicity

Evaluation not performed for this substance

### 10.2 Skin corrosion/irritation

Evaluation not performed for this substance

### 10.3 Serious eye damage/eye irritation

Evaluation not performed for this substance

### 10.4 Respiratory sensitisation

Evaluation not performed for this substance

### 10.5 Skin sensitisation

Evaluation not performed for this substance

### 10.6 Germ cell mutagenicity

**Table 10: Summary table of mutagenicity/genotoxicity tests in vitro**

Method, guideline, deviations if any	Test substance	Relevant information about the study including rationale for dose selection (as applicable)	Observations	Reference
<b>Bacterial gene mutation (Ames test)</b>				
<p><i>S. typhimurium</i> (TA 98, TA 100), ± S9-mix, preincubation (Tests 1-4),</p> <p><i>E.coli</i> WP2 ± S9-mix, preincubation (Tests 5,6)</p> <p>No explicit mentioning of OECD-TG or GLP, equivalent reliability, but only three strains tested</p> <p>Reliability according to disseminated database: study not reported</p> <p>Reliability according to authors of this evaluation: 3</p>	<p>Cumene</p> <p>Purity: &gt;99%; no impurities &gt; 0.1% observed</p> <p>Vehicle DMSO</p>	<p><u>All tests:</u></p> <p>Measures taken to avoid influence from volatility (sealed tubes), results reported by mean ± SEM, triplicate test</p> <p><u>Test 1 (TA 100)</u></p> <p>a) without activation: 0 - 250 µg/plate (≥125 µg/plate (slightly toxic or toxic) + positive control (sodium azide),</p> <p>b) with activation (10% phenobarbital/benzoflavone-induced rat liver S9-mix): 0 - 500 µg/plate (≥250 µg/plate (slightly toxic) + positive control (benzo(a)pyrene)</p> <p><u>Test 2 (TA 100)</u></p> <p>a) without activation: 0 - 250 µg/plate (≥100 µg/plate</p>	<p><u>All tests:</u></p> <p>without and with activation (S9-mix):</p> <p>→ negative</p>	(NTP, 2012)

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Method, guideline, deviations if any	Test substance	Relevant information about the study including rationale for dose selection (as applicable)	Observations	Reference
		<p>(slightly toxic) + positive control (sodium azide)</p> <p>b) 10% phenobarbital/benzoflavone-induced rat liver S9-mix 0 - 250 µg/plate(250 µg/plate (slightly toxic) + positive control (benzo(a)pyrene)</p> <p><u>Test 3 (TA 98)</u></p> <p>a) without activation: 0 - 500 µg/plate (≥250 µg/plate (slightly toxic) + positive control (2-nitrofluorene),</p> <p>b) with activation (10% phenobarbital/benzoflavone-induced rat liver S9-mix–mix): 0 - 500 µg/plate + positive control (2-aminoanthracene)</p> <p><u>Test 4 (TA 98)</u></p> <p>a) without activation: 0 - 250 µg/plate (≥100 µg/plate (slightly toxic or toxic, resp.) + positive control (2-nitrofluorene)</p> <p>b) 10% phenobarbital/benzoflavone-induced rat liver S9-mix: 0 - 500 µg/plate (≥250 µg/plate (slightly toxic) + positive control (2-aminoanthracene)</p> <p><u>Test 5 (<i>E.coli</i> WP2)</u></p> <p>a) without activation: 0 - 500 µg/plate (≥100 µg/plate (slightly toxic or toxic, resp.)) + positive control (4-nitroquinoline-N-oxide)</p> <p>b) with activation (10% phenobarbital/benzoflavone-induced rat liver S9-mix): 0 - 500 µg/plate (500 µg/plate (slightly toxic)) + positive control (2-aminoanthracene)</p> <p><u>Test 6 (<i>E.coli</i> WP2)</u></p> <p>a) without activation: 0 - 500 µg/plate (≥125 µg/plate (slightly toxic or toxic, resp.)) + positive control (4-nitroquinoline-N-oxide)</p> <p>b) with activation (10%</p>		

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Method, guideline, deviations if any	Test substance	Relevant information about the study including rationale for dose selection (as applicable)	Observations	Reference
		phenobarbital/benzoflavone-induced rat liver S9-mix): 0- 500 µg/plate (500 µg/plate (toxic)) + positive control (2-aminoanthracene)		
<p><i>S. typhimurium</i> (TA 97, TA 98, TA 100, TA 1535) ± S9-mix, preincubation</p> <p>NTP conducts its studies in compliance with FDA Good Laboratory Practice Regulations. Limited remaining uncertainties because of a) potential volatility losses, b) highest concentrations not always reaching toxicity level, but TA 102 or E. coli strains not tested</p> <p>Reliability according to disseminated database: 1</p> <p>Reliability according to authors of this evaluation: 3</p>	<p>Cumene</p> <p>Purity: &gt;99.9%; no impurities &gt; 0.1% observed</p> <p>Vehicle: no data</p>	<p><u>All tests:</u></p> <p>Each trial: triplicate plates plus concurrent positive and negative controls, 5 doses of cumene. All trials repeated. Negative trials with S9-mix repeated with higher S9-mix concentrations (10%; 30%). In some, but not in all tests, the highest concentration (i.e., 166 or 333 µg/plate) was slightly toxic. For TA97 (-S9-mix) already 100 µg/plate was slightly toxic.</p> <p>a) without activation: 0-333 µg/plate or 0-166 µg/plate</p> <p>positive controls: sodium azide (TA100; TA1535); 9-aminoacridine (TA 97), 4-nitro-o-phenylenediamine (TA98)</p> <p>b) with activation (from Aroclor 1254-induced male Sprague-Dawley rat or Syrian hamster liver S9-mix):</p> <p>positive controls (all strains): 2-aminoanthracene</p>	<p><u>All tests:</u></p> <p>without and with activation (S9-mix): → negative</p>	<p>(NTP, 2009)</p> <p>Study also reported in (ECHA Dissemination, 2018, genetic toxicity, in vitro, key #001)</p>
<p><i>S. typhimurium</i> (TA 98, TA 100, TA 1535, TA1537) ± S9-mix, preincubation</p> <p>Certified compliance with GLP, but TA 102 or E. coli strains not tested</p> <p>Reliability according to disseminated database: 1</p> <p>Reliability according to authors of this evaluation: 3</p>	<p>Cumene;</p> <p>Purity: no data</p> <p>Vehicle: Pluronic F127, prepared at 50% (w/w) in ethanol</p>	<p><u>All tests:</u></p> <p>7 dose levels of cumene (minimum 4 non-toxic dose levels) along with untreated, vehicle and positive control, ± 10% S9-mix, plus additional positive control in F127; plus single maximally water soluble dose of cumene tested on all four tester strains ± S9-mix. Prior range finding study to determine toxic potency.</p> <p>Dose range: 33, 67, 100, 333, 667, 1000, 2000 µg/plate</p> <p>All experimental results confirmed in repeat experiment.</p> <p>a) without activation:</p> <p>positive control: TA98, 5.0 µg 2-nitrofluorene</p> <p>TA100, TA1535: 2.5 µg sodium azide</p>	<p><u>All tests:</u></p> <p>without and with activation (S9-mix): → negative</p>	<p>(Lawlor und Wagner, 1987)</p> <p>Study also reported in (ECHA Dissemination, 2018, genetic toxicity, in vitro, #005,supporting)</p>

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Method, guideline, deviations if any	Test substance	Relevant information about the study including rationale for dose selection (as applicable)	Observations	Reference
		TA1537, 75 µg 9-aminoacrdine  b) with activation (Aroclor induced rat liver microsomes S9-mix):  positive control:  TA98, TA100, TA1535, TA1537: 2.0 µg aminoanthracene		
<i>S. typhimurium</i> (TA 98, TA 100, TA 1535, TA1537, TA1538) ± S9-mix  Reliability according to disseminated database: 4  Reliability according to authors of this evaluation: 4	Cumene, no details provided	Tested up to 5000 µg/plate, no details provided	<u>All tests:</u> without and with activation (S9-mix): → negative	(Huels, 1987; unpublished, cited from ECHA Dissemination, 2018, genetic toxicity, in vitro, #008, other)
<i>S. typhimurium</i> (TA 98, TA 100, TA 1535, TA1537) ± S9-mix, Reliability according to disseminated database: 4  Compliance with Monsanto Standard Operation Procedures. According to test authors: "minor GLP violations did not impact study results"  Potential Influence of volatility may not be excluded  Reliability according to authors of this evaluation: 2	Cumene  Purity: 99.5%  Vehicle: ethanol	<u>All tests:</u>  Up to 0.2 µl/plate and 20 µl/spot (spot test) ± S9-mix; 0.2 µg/plate were toxic to all four test strains ± S9-mix; triplicate testing, solvent controls, non-solvent controls, positive controls  a) without activation:  Positive controls  TA98, TA100: 4-nitroquinoline-N-oxide  TA1535: NaNO <sub>2</sub>  TA1537: 9-aminoacridine  b) with activation (S9-mix from livers of Aroclor 1254-induced male Sprague-Dawley rats and male CD-1 mice):  Positive controls  TA98: 2-acetylaminofluorene  TA100: benzo(a)pyrene  TA1535, TA1537: 2-aminoanthracene	<u>All tests:</u> without and with activation (S9-mix): → negative	(Monsanto Co, 1985)  Study also reported in:  (ECHA Dissemination, 2018, genetic toxicity, in vitro, #010, other)
<i>S. typhimurium</i> (TA 98, TA 100, TA 1535,	Cumene  Purity: no	3.6- 3606.0 µg/plate (0.03 - 30 µmol/plate) ± S9-mix (4 doses);	<u>All tests:</u> without and with	(Florin <i>et al.</i> , 1980)

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Method, guideline, deviations if any	Test substance	Relevant information about the study including rationale for dose selection (as applicable)	Observations	Reference
TA1537) ± S9, spot test and plate incorporation  TA 102 or E. coli strains not tested  Reliability according to disseminated database: 4  Reliability according to authors of this evaluation: 3	data  Vehicle: no data	toxic dose ≥ 3 µmoles/plate;  No details on substance specific test outcome provided (qualitative result documentation)  a) without activation:  positive control all testers: N-methyl-N'-nitro-N-nitrosoguanidin  b) with activation (S9-mix: from Aroclor 1254 or Methylcholanthrene induced rats)  positive control all testers: 2-aminoanthracene	activation (S9-mix): → negative	Study also reported in (ECHA Dissemination, 2018, genetic toxicity: in vitro, #011 other))
<i>S. typhimurium</i> (TA 98, TA 100, TA 1535, TA1537, TA1538) ± S9, plate incorporation  Reliability according to disseminated database: study not reported  Reliability according to authors of this evaluation: 4	Cumene, reagents of the highest available purity, but generally purity was not determined and not specified	Study included without details in report on 300 chemicals tested, identified in drinking water. Max. dose 5 mg/plate or a (lower) dose giving a toxic response (not reported for cumene). No explicit details on S9-mix metabolic activation provided. Positive and negative (solvent) controls were included.	<u>All tests:</u> without and with activation (S9-mix): → negative  Cumene was also tested negative in <i>S. typhimurium</i> desiccator testing (no strain and no details provided)	(Simmon <i>et al.</i> , 1977)
<i>S. typhimurium</i> (TA 100) ± S9-mix, Spot test  Reliability according to disseminated database: 3  Reliability according to authors of this evaluation: 4	Cumene  Purity: no data	Study included without details in report on almost 300 chemicals tested, identified in tap water. Approximately one third of those have been spot-tested in TA100 ±S9-mix. No details provided.	Compounds tested and found to be mutagenic in <i>S. typhimurium</i> TA100 include isopropylbenzene (cumene) → positive	(Tardiff <i>et al.</i> , 1978)  Study also reported in (ECHA Dissemination, 2018, genetic toxicity, in vitro, #007, other)
<b>Yeast <i>S. cerevisiae</i></b>				
Yeast <i>S. cerevisiae</i> D3 assay, suspension  Reliability according to disseminated database: study not reported  Reliability according to authors of this evaluation: 4	Cumene, reagents of the highest available purity, but generally purity was not determined and not specified	Study included without details in report on 300 chemicals tested, identified in drinking water, only "some of which" were tested in <i>Saccharomyces cerevisiae</i> D3. Positive and negative (solvent) controls were included. Cytotoxicity not reported.	negative	(Simmon <i>et al.</i> , 1977)

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Method, guideline, deviations if any	Test substance	Relevant information about the study including rationale for dose selection (as applicable)	Observations	Reference
<b>Mammalian Cells</b>				
<p>CHO/HGPRT mutation assay ± S9-mix</p> <p>Reliability according to disseminated database: 1</p> <p>Certified to be conducted in compliance with GLP, but stability of test or control substances have not been determined.</p> <p>Reliability according to authors of this evaluation: 2</p>	<p>Cumene</p> <p>Purity: 99.7%</p> <p>Vehicle: Pluronic polyol F127 (1:1 in ethanol)</p>	<p>Test for ability to induce forward mutations at the HGPRT-locus of Chinese Hamster Ovary Cells (CHO-cells).</p> <p><u>All tests:</u></p> <p>Untreated control, solvent control (F127), three doses of positive controls</p> <p>9 concentrations in dose range: 8-225 µg/ml</p> <p>a) without activation:</p> <p>positive control: ethyl methanesulfonate (0.2 µl/ml) (toxic &gt; 125 µg/ml)</p> <p>b) with activation (S9-mix from livers of Aroclor 1254-induced rats):</p> <p>positive control: benzo(a)pyrene (4 µg/ml) (toxic &gt; 125 µg/ml)</p>	<p><u>All tests:</u></p> <p>without and with activation (S9-mix):</p> <p>→ negative</p>	<p>(Yang, 1987)</p> <p>Study also reported in (ECHA Dissemination, 2018, genetic toxicity: in vitro, #004, key))</p>
<p>CHO/HGPRT mutation assay ± S9-mix</p> <p>Reliability according to disseminated database: 4</p> <p>Reliability according to the authors of this evaluation: 2</p>	<p>Cumene,</p> <p>Purity: no data</p> <p>Vehicle: Pluronic F127 (1:1 in ethanol)</p>	<p>Test for ability to induce forward mutations at the HGPRT-locus of Chinese Hamster Ovary Cells (CHO-cells).</p> <p><u>All tests:</u></p> <p>Untreated control, solvent control (F127), positive controls</p> <p><u>Test 1</u> (November, 1984)</p> <p>8, 16, 32, 64, 128, 150, 175 µg/ml ±S9-mix;</p> <p>a) without activation: cytotoxicity (colony counts) ≥ 128 µg/ml</p> <p>Positive control: Ethylmethanesulfonate</p> <p>b) with activation (S9-mix from livers of Aroclor 1254-induced rats):</p> <p>cytotoxicity (colony counts) ≥ 128 µg/ml; cell count reduction already at ≥ 16 µg/ml</p> <p>Positive control: Benzo(a)pyrene</p> <p><u>Test 2</u> (February, 1985)</p>	<p>Test 1, negative ± S9-mix, but potential positive outlier at 175 µg/ml (+S9-mix). Confirmation of effect as outlier by test 2 (negative in test 2)</p> <p>→ negative</p>	<p>(Gulf Oil Corporation, 1985a)</p> <p>Study also reported in (ECHA Dissemination, 2018, genetic toxicity, in vitro, #009, other))</p>



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Method, guideline, deviations if any	Test substance	Relevant information about the study including rationale for dose selection (as applicable)	Observations	Reference
		because of a potential positive outlier at 175 µg/ml +S9-mix Test 1 a repeat test (Test 2) was performed  150,175 µg/ml +S9-mix  Positive control: Benzo(a)pyrene		
Chromosomal aberrations (CHO cells) ± S9-mix  Reliability according to disseminated database: 1  Compliance with GLP stated by authors.  Reliability according to authors of this evaluation: 2	Cumene  Purity: 99.7%,  Vehicle: Pluronic F127 (1:1 in ethanol)	Test for chromosomal aberrations in Chinese hamster ovary cells:  negative control: untreated cells, plus vehicle control (F127)  a) without activation:  Dosing 0, 19-200 µg/ml (7 doses), Toxic at 200 µg/mL, high dose  positive control: triethylenemelamine  b) with activation (S9-mix from livers of Aroclor induced rats):  Dosing 0, 24-225 µg/ml (6doses),  Toxic at 225 µg/mL (+S9-mix), high dose	Negative –S9-mix, Increased vs. vehicle control at 156 µg/ml + S9-mix (low F127 in control), but no statistically significant increase compared to untreated control and within historical control range, regarded as negative by authors  → inconclusive	(Putman, 1987a)  Study also reported in (ECHA Dissemination, 2018, genetic toxicity: in vitro, #003, key)
UDS –S9-mix  Reliability according to disseminated database: no documented study  Authors state GLP compliance  Reliability according to authors of this evaluation: 2	Cumene  Purity: no data  Vehicle: pluronic F68 Polyol	Tested for unscheduled DNA synthesis using primary rat hepatocytes.  Negative control: vehicle (F68), untreated control, positive control: 2-acetylaminofluorene, dosing (triplicate test) : 8 – 128 µg/ml (5 doses), 128 µg/ml toxic	An increase in grain counts was obtained at 16 and 32 µg/ml. Although this increase in grain counts was not regarded to be clearly positive by authors, there was a significant increase (p<0.01) in percentage of cells in repair at those dose levels.  → positive, retested by Curren (1987)	(Gulf Oil Corporation, 1984)
UDS – S9-mix  Reliability according to disseminated database: 1  Authors state GLP compliance, but note that, e.g., purity and stability	Cumene  Purity: 99.7%  Vehicle: Pluronic Polyol F127 (1:1; ethanol)	Tested for unscheduled DNA synthesis using primary rat hepatocytes.  Primary hepatocytes, rat F344, without metabolic activation, vehicle control, negative control, positive control, test specific confounding factors not reported	No significant increase in unscheduled DNA synthesis as measured by mean number of net grain counts (i.e., an increase of at least 5 counts over control)	(Curren, 1987)  Study also reported: (ECHA Dissemination, 2018, genetic toxicity, in vitro,

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Method, guideline, deviations if any	Test substance	Relevant information about the study including rationale for dose selection (as applicable)	Observations	Reference
of test and control substance have not been determined  Reliability according to authors of this evaluation: 2		13 doses: 1-128 µg/mL; doses > 24 µg/mL toxic; fully evaluated at 6 dose levels (1-24 µg/ml).  This study was performed as a retest because of increased cell repair observed in Gulf Oil Cooperation (1984)	→ negative	key #002)
<b>(Assumed or confirmed) METABOLITES</b>				
<b>α – methyl styrene</b>				
<i>Salmonella typhimurium</i> TA97, TA98, TA100, TA1535 ± (rat or hamster) S9-mix, <i>in vitro</i>  TA 102 or E. coli strains not tested  Reliability according to disseminated database: study not reported  Reliability according to authors of this evaluation: 3	α - Methyl styrene  Purity: 99.5%  Vehicle: no data	<u>All tests:</u>  Each trial consisted of triplicate plates including concurrent positive and negative controls, plus 5 doses α - Methyl styrene. High dose limited by toxicity. All trial repeated at the same or higher S9-mix fraction  Dosing: 1-3333 µg/plate  a) without activation  positive control: TA100, TA1535: sodium azide TA97: 9-aminoacridine TA98: 4-nitro-o-phenylenediamine  slight toxicity at 333 µg/plate  b) with activation (S9-mix from Aroclor 1254-induced male Sprague-Dawley rat or Syrian hamster liver):  slight toxicity at 1000 µg/plate (S9-mix)  positive control: all strains 2-aminoanthracene	<u>All tests:</u>  without and with activation (S9-mix):  → negative	(NTP, 2007)
Chromosomal aberrations, CHO cells, no metabolic activation  Reliability according to disseminated database: 1  Reliability according to authors of this evaluation: 2	α - Methyl styrene  Purity: 99.5%  Vehicle: DMSO	<u>All tests:</u>  Negative control: vehicle (DMSO)  2 trials  <u>Trial 1:</u>  Dosing: 100-200 µg/ml (3 doses)  positive control: Mitomycin C  not toxic up to highest dose	<u>In both trials:</u>  → negative	(NTP, 2007)  Study also reported in:  (ECHA Dissemination, 2018, genetic toxicity: in vitro, #007, supporting)

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Method, guideline, deviations if any	Test substance	Relevant information about the study including rationale for dose selection (as applicable)	Observations	Reference
		<p><u>Trial 2:</u> Dosing: 37.7-251.3 µg/ml (4 doses) positive control: Mitomycin C toxic at highest dose: 251.3 µg/ml</p>		
<p>SCE, <i>in vitro</i>, tested only –S9-mix</p> <p>Reliability according to disseminated database: 2</p> <p>Reliability according to authors of this evaluation: 3</p>	<p>α - Methyl styrene</p> <p>Purity: &gt; 97%</p> <p>Vehicle: acetone</p>	<p>Test for induction of sister-chromatid exchanges by test substance in cultured human lymphocytes</p> <p>Control cultures treated with vehicle (acetone)</p> <p>Dosing: 5 doses (0.1-4 mM), cell cycle delay (measure for toxicity) increased at 4 mM.</p> <p>Limited documentation (only graphical presentation of results, no individual results for test substance and controls reported)</p>	<p>Weakly positive – S9-mix at &gt; 1 mM (less than doubling of SCEs compared to corresponding controls)</p>	<p>(Norppa und Vainio, 1983)</p> <p>Study also reported in: (ECHA Dissemination, 2018, genetic toxicity, <i>in vitro</i>, #010, supporting)</p>
<p>SCE, <i>in vitro</i>, ± S9-mix</p> <p>Reliability according to disseminated database: 2</p> <p>According to disseminated database, study design comparable to OECD guideline 479</p> <p>Reliability according to authors of this evaluation: 2</p>	<p>α - methyl styrene</p> <p>Purity: 99.5%</p> <p>Vehicle: DMSO</p>	<p><u>All tests:</u></p> <p>Negative control: vehicle (DMSO)</p> <p>a) without activation: Dosing: 5-166.7 µg/ml (4 doses) positive control: Mitomycin C 166.7 µg/ml toxic</p> <p>b) two trials with activation (S-9 mix from Aroclor 1254-induced male Sprague-Dawley rat or Syrian hamster liver) Dosing Trial 1: 5-166.7 µg/ml (4 doses) Dosing Trial 2: 50-149.9 µg/ml (3 doses)</p> <p>positive control: cyclophosphamide 166.7 µg/ml toxic</p>	<p>→ Negative without S9-mix</p> <p><u>Trial 1:</u> → at 50 µg/ml +S9 relative change in SCEs/chromosome: 28.42% , trend: <math>p \leq 0.001</math> → positive</p> <p><u>Trial 2:</u> Dose related increase of SCE at 50, 124.4, or 149.9 µg/ml +S9 (relative change in SCEs/chromosome: 39.59, 49.16, 82.77 %, respectively) trend: <math>p \leq 0.001</math> → positive</p>	<p>(NTP, 2007)</p> <p>Study also reported in: (ECHA Dissemination, 2018, genetic toxicity, <i>in vitro</i>, #003, key)</p>
<b>α - methyl styrene oxide</b>				
<p>S. Typh. TA100, preincubation</p>	<p>α - methyl styrene</p>	<p>Dosing: 7 doses (0.01-10 µmoles/ preincubation tube) plus DMSO</p>	<p>Dose related increase in number of</p>	<p>(Rosman <i>et al.</i>,</p>

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Method, guideline, deviations if any	Test substance	Relevant information about the study including rationale for dose selection (as applicable)	Observations	Reference
<p>Reliability according to disseminated database: substance not registered under REACH</p> <p>Reliability according to authors of this evaluation: 3</p>	<p>oxide</p> <p>Purity: no data</p> <p>Vehicle: DMSO</p>	<p>(negative control), highest dose (10 µmole) toxic, triplicate plates. Positive control: glycidol (no results on positive control reported), but specific potency data for other derivatives of α - methyl styrene oxide</p>	<p>revertants → positive</p>	1986)

**Table 11: Summary table of mutagenicity/genotoxicity tests in mammalian somatic or germ cells in vivo**

Method, guideline, deviations if any	Test substance,	Relevant information about the study (as applicable)	Observations	Reference
<b>Micronuclei</b>				
<p>F344 ♂rats, i.p.</p> <p>No explicit mentioning of OECD-TG. NTP conducts its studies in compliance with FDA Good Laboratory Practice Regulations.</p> <p>Reliability according to disseminated database: 1</p> <p>Reliability according to authors of this evaluation: 2</p>	<p>Cumene</p> <p>Purity: &gt;99.9%; no impurities &gt; 0.1% observed</p> <p>Vehicle: corn oil</p>	<p>Endpoint: micronucleated polychromatic erythrocytes, in bone marrow.</p> <p>2 Trials: n=5 animals/dose/trial</p> <p>Negative controls: corn oil (vehicle)</p> <p>Positive controls: injected 25 mg/kg (3x, intervals 24h) cyclophosphamide</p> <p><u>Trial 1:</u> Dosing: 6 groups: 78.13 - 2500 mg/kg three times (intervals 24h); 2500 mg/kg high mortality</p> <p><u>Trial 2:</u> 0, 312-2500 mg/kg three times; 2500 mg/kg elevated mortality</p>	<p><u>Trial 1:</u> Pairwise comparison. Highest statistically evaluated dose (1250 mg/kg) significantly elevated number of micronucleated PCE (P=0.0001), trend (P&lt;0.001; highest dose, 2500 mg/kg, excluded from statistical analysis) → positive</p> <p><u>Trial 2:</u> Pairwise comparison. Micronuclei elevated at all four tested doses (312, 625, 1250 mg/kg, 2500 mg/kg) (P=0.0052, P=0.0194, P=0.0033; P=0.0192), but not significant (criterion: P≤0.006), nonsignificant trend (P=0.085) → questionably positive → combined: positive</p>	(NTP, 2009)  Study also reported in (ECHA Dissemination, 2018, genetic toxicity, in vivo, #002, key)
<p>F344/ DuCrI, ♂rats, gavage</p> <p>No explicit mentioning of OECD-TG or GLP, but equivalent reliability ensured.</p>	<p>Cumene</p> <p>Purity: &gt;99%; no impurities &gt; 0.1% observed</p> <p>Vehicle: Corn oil</p>	<p>6 animals/dose group, vehicle control (corn oil), 1x/day, 4 consecutive days, gavage, 0, 200, 400, 800 mg/kg/d; positive control: ethyl methanesulfonate (200 mg/kg/d)</p> <p>% of circulating reticulocytes significantly reduced at top dose (30%),</p>	<p>No significant increases in micronucleated erythrocytes (NCE) or reticulocytes (PCE) were observed → negative</p>	(NTP, 2012)

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Method, guideline, deviations if any	Test substance,	Relevant information about the study (as applicable)	Observations	Reference
<p>Reliability according to disseminated database: study not reported</p> <p>Reliability according to authors of this evaluation: 1</p>		supports dose selection		
<p>B6C3F<sub>1</sub> mice, ♀,♂; gavage</p> <p>No explicit mentioning of OECD-TG or GLP, but equivalent reliability ensured.</p> <p>Reliability according to disseminated database: study not reported</p> <p>Reliability according to authors of this evaluation: 1</p>	<p>Cumene</p> <p>Purity: &gt;99%; no impurities &gt; 0.1% observed</p> <p>Vehicle: Corn oil</p>	<p>6 animals/dose group, vehicle control (corn oil), 1x/day, 4 consecutive days, gavage, 0, 312,625,1250, mg/kg/d (♂) or 0,250,500,1000 mg/kg/d (♀) ; positive control: ethyl methanesulfonate (150 mg/kg/d; n=5)</p>	<p>No significant increases in micronucleated erythrocytes (NCE) or reticulocytes (PCE) were observed</p> <p>→ negative</p>	(NTP, 2012)
<p>B6C3F<sub>1</sub> mice, ♀,♂; inhalation</p> <p>No explicit mentioning of OECD-TG. NTP conducts its studies in compliance with FDA Good Laboratory Practice Regulations.</p> <p>Reliability according to disseminated database: 1</p> <p>Reliability according to authors of this</p>	<p>Cumene</p> <p>Purity: &gt;99.9 %; no impurities &gt; 0.1% observed</p>	<p>Endpoint: micronucleated cells in normochromatic erythrocytes after 3 month inhalation exposure</p> <p>Concentrations:</p> <p>♂: 0, 62.5, 125, 250, 500, 1000 ppm (306-4900 mg/m<sup>3</sup>)</p> <p>♀: 0, 62.5, 125, 250, 500 ppm (306-2450 mg/m<sup>3</sup>)</p> <p>n=9 or 10 animals/dose group; exposure 6 hrs. per day, 5 days/week, 14 weeks</p>	<p>Peripheral Blood Erythrocytes of Mice following inhalation treatment for 3 months: no significant difference from concurrent air control group, no significant trend</p> <p>→ negative</p>	<p>(NTP, 2009)</p> <p>Study also reported in (ECHA Dissemination, 2018, genetic toxicity: in vivo, #001,key)</p>

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Method, guideline, deviations if any	Test substance,	Relevant information about the study (as applicable)	Observations	Reference
evaluation: 2				
Swiss Mice, ♀,♂; gavage No explicit mentioning of OECD-TG, GLP compliance is given (including certificate) Reliability according to disseminated database: 2 Reliability according to authors of this evaluation: 3	Cumene; purity: 2.5 g cumene in 50 mL paraffin oil (5% w/v) (according to Disseminated database)	Test for micronucleated polychromatic erythrocytes in bone marrow  Dosing: 0.25, 0.5, 1 g/kg body weight, gavage, BR Swiss mice, 2 days (highest dose some only 1 single day), at ≥ 1.25 g/kg ♀ mortality. N= 10/dose/sex.  Negative control: paraffin oil (20 ml/kg),  Positive control: cyclophosphamide (N=4)	No effect on micronucleated polychromatic erythrocytes in bone marrow under conditions of this test  → negative	(Gulf Oil Corporation, 1985b)  Study also reported in (ECHA Dissemination, 2018 genetic toxicity: in vivo, #003, supporting)
<b>Comet Assays</b>				
♂, F344/N, gavage No explicit mentioning of OECD-TG or GLP, but equivalent reliability ensured. Reliability according to disseminated database: study not reported Reliability according to authors of this evaluation: 1	Cumene  Purity: >99%; no impurities > 0.1% observed  Vehicle: corn oil	DNA-damage analysed in blood, lung, kidney, liver  6 animals/dose group, vehicle control (corn oil), 1x/day, 4 consecutive days, gavage, 0, 200, 400, 800 mg/kg/d; positive control: ethyl methanesulfonate (200 mg/kg/d)	Liver positive for % tail DNA (p=0.004 at highest dose: 800 mg/kg/d; p=0.002 for trend), all other sites negative (blood, lung, kidney)  → weakly positive for liver, male rats	(NTP, 2012)
♀,♂; B6C3F <sub>1</sub> mice, gavage No explicit mentioning of OECD-TG or GLP, but equivalent	Cumene  Purity: >99%; no impurities > 0.1% observed  Vehicle: corn	Blood, lung, kidney, liver  6 animals/dose group, vehicle control (corn oil), 1x/day, 4 consecutive days, gavage, 0, 312, 625, 1250, mg/kg/d (♂) or 0, 250, 500, 1000 mg/kg/d (♀) ; positive	♀: lung positive for % tail DNA (p=0.016 at highest dose: 1000 mg/kg/d; p=0.008 for trend), all other sites negative (blood, lung, kidney)  ♂: negative all sites	(NTP, 2012)

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Method, guideline, deviations if any	Test substance,	Relevant information about the study (as applicable)	Observations	Reference
reliability ensured.  Reliability according to disseminated database: study not documented  Reliability according to authors of this evaluation: 1	oil	control: ethyl methanesulfonate (150 mg/kg/d; n=6)	→ weakly positive for lung, female mice	
FLARE: Fragment Length Analysis with Repair Enzyme  Reliability according to disseminated database: study not reported  Reliability according to authors of this evaluation: 4	Cumene  Purity: no data	80 SD rats were assigned to 4 dose groups, exposed to cumene vapour for 90 days, 6h/d, to 0, 8.05, 80.13 or 800.85 ppm (40-3920 mg/m <sup>3</sup> ). In hepatocytes and lymphocytes olive tail moment and tail length were measured and rOGG1 mRNA expression from hepatocytes was scored for cells from different exposure durations (1d, 14d, 28d, 90d). OGG1 is a DNA damage repair gene. The other assays indicate DNA damage similar to the Comet assay.	Significant changes olive tail moment and tail length, indicating DNA damage in hepatocytes and lymphocytes from cumene exposure. OGG1 gene expression to repair oxidative DNA damage in liver was inhibited after significant increase at the first day of exposure. The results demonstrate oxidative DNA damage, but a dose-response or duration-effect relationship cannot be established from this study.	(Kim <i>et al.</i> , 2008)
<b>Mutations in Tumours</b>				
Biochemical analysis (including mutation analysis)  Reliability according to disseminated database: study not reported  Reliability according to authors of this evaluation: 1	Cumene  Purity: >99.9%; no impurities > 0.1% observed  (from NTP, 2009)	Study includes 52 cumene induced lung tumours examined for <i>K-ras</i> mutations, p53 mutations, p53 protein expression and “loss of heterozygosity” (LOH).  (52= 6 adenoma, 46 carcinoma) and compared to control (concurrent: n=7 tumours; historical: n=117 tumours). 45 tumours in exposed ♂ examined, 9 tumours in ♀.	<b><u>K-ras or p53 mutation in tumours observed:</u></b>  50% in adenoma (3/6), 52% in carcinoma (24/46)  <b><u>K-ras:</u></b>  Mutations in cumene induced lung tumours  -dose response (treatment ppm; no. of tumours with <i>K-ras</i> mutations %)  Control (historical): 0 ppm:28% Control (concurrent): 0 ppm: 14% Exposed 125 ppm: 25% Exposed 250 ppm: 77% Exposed 500 ppm: 94% Exposed 1000 ppm: 100%	(Hong <i>et al.</i> , 2008)

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Method, guideline, deviations if any	Test substance,	Relevant information about the study (as applicable)	Observations	Reference
			<p>Exposed (total): 87%</p> <p>K-<i>ras</i> mutations more prevalent in ♂ (91%; n=41/45) vs. ♀ (57%; n=4/7) (few tumours from ♀ analysed!)</p> <p><u>K-<i>ras</i> mutation spectra:</u></p> <p><u>predominant K-<i>ras</i> mut. in exposed:</u></p> <p>-codon 12, G→T transversion (36%), G→T transversion in hist. control (18%)</p> <p>-codon 61, A→G transitions (29%), A→G transitions in hist. control (6%)</p> <p><u>predominant K-<i>ras</i> mut. in control:</u></p> <p>Codon 12, G→A transitions (42%)</p> <p><b>p53:</b></p> <p>Control (historical): no data provided</p> <p>Control (concurrent): 0 ppm: 0%</p> <p>Exposed (total): 52%</p> <p>p53 mutations more prevalent in ♂ (58%; n=26/45) vs. ♀ (14%; n=1/7) (few tumours from ♀ analysed!)</p> <p>p53 mutations were correlated with increased p53 protein expression and protein expression was exposure related:</p> <p>p53 <u>protein expression</u> changed in tumours:</p> <p>control: 1/7 (14%)</p> <p>125 ppm: 1/4 (25%)</p> <p>250 ppm: 6/13 (46%)</p> <p>500 ppm: 8/18 (44%)</p> <p>1000 ppm: 14/17 (82%)</p> <p>Exposed (total): 29/52 (56%)</p> <p><b>LOH analysis:</b></p> <p>LOH on chromosome 6 near K-<i>ras</i> gene was observed:</p> <p>12% in carcinomas (cumene exposed)</p> <p>0% in adenoma (cumene exposed)</p> <p>0% in spontaneous carcinoma</p> <p>LOH of the C3H/He allele was observed on chromosome 4 near p16 gene (allele loss of p16 detected in human cancer):</p>	



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Method, guideline, deviations if any	Test substance,	Relevant information about the study (as applicable)	Observations	Reference
			13% in carcinomas (cumene exposed) 17% in adenoma (cumene exposed) 0% in spontaneous carcinoma	
<b>(Assumed or confirmed) metabolites</b>				
<b><math>\alpha</math> - methyl styrene</b>				
Micronuclei, <i>in vivo</i> , ♂, ♀ in mice  Reliability according to disseminated database: 2  Reliability according to authors of this evaluation: 2	$\alpha$ - methyl styrene  Purity: 99.5%	3-month inhalation exposure of ♂ and ♀ mice (♂, ♀: 0, 75, 150, 300, 600, 1000 ppm; i.e., 0, 360-4800 mg/m <sup>3</sup> , n=10/sex/group). Peripheral blood samples were scanned for the frequency of micronuclei in normochromatic erythrocytes (NCE) and in polychromatic erythrocytes (PCE)	<u>Peripheral Blood:</u> ♀: Trend ( $p \leq 0.001$ ) and highest concentration (1000 ppm; $p=0.0006$ ) positive for increase of micronucleated cells (NCE); no increase in micronucleated PCE seen at the 1000 ppm dose. No dose-response in the percent PCE  ♂ negative response, no dose related changes  → weakly positive, significance uncertain	(NTP, 2007)  Study also reported in (ECHA Dissemination, 2018, genetic toxicity, <i>in vivo</i> , key)
Micronuclei, <i>in vivo</i> , ♂, in mice  Reliability according to disseminated database: study not reported  Reliability according to authors of this evaluation: 2	$\alpha$ - methyl styrene  Purity: 99%	ICR-mice, (n= 6/dose, orally) dosage: 0, 500, 1000, 2000 mg/kg, single exposure. Bone marrow cells were scanned for the frequency of micronuclei in polychromatic erythrocytes (PCE). Positive control: mitomycin C. No inhibition of proliferation within the dose range of this test. No further data on cytotoxicity provided.	→ negative	(Rim <i>et al.</i> , 2012)

For  $\alpha$  - methyl styrene only the most relevant studies on genotoxicity are provided in Table 10 or Table 11, respectively. A more complete overview is found, e.g., in NTP (2007).

**Table 12: Summary table of human data relevant for germ cell mutagenicity**

Type of data/report	Test substance,	Relevant information about the study (as applicable)	Observations	Reference
No human data on cumene are available, which are relevant for germ cell mutagenicity assessment				

**10.6.1 Short summary and overall relevance of the provided information on germ cell mutagenicity**

No human data were available for the assessment of genotoxicity of cumene. Results of *in vitro*- or *in vivo*-testing are summarised in Table 10 or Table 11, respectively.

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Cumene was not mutagenic in the *Salmonella typh.* mutagenicity assay in a variety of strains<sup>1</sup>, in *E. coli* or in yeast with or without metabolic activation. A single study with a positive result in *S. typh.* TA100 (Tardiff *et al.*, 1978) is not regarded as reliable because of insufficient reporting. Cumene was tested in mammalian cells for genotoxicity (CHO/HGPRT assay; chromosomal aberrations in CHO cells and UDS test) with negative results; however, some early inadequate tests with equivocal or positive findings had to be repeated and, then, confirmed the overall negative outcome (NTP, 2013).

The metabolite,  $\alpha$ -methylstyrene, was negative in *Salmonella typh.*, and did not induce chromosomal aberrations *in vitro*. However, positive effects in sister chromatid exchange *in vitro* were observed (Norppa und Vainio, 1983; NTP, 2007). The putative metabolite,  $\alpha$ -methylstyrene oxide, yielded positive results in a reverse mutation test in *S. typh.* (Rosman *et al.*, 1986). No data were available on genotoxicity of other metabolites of cumene.

*In vivo*, intraperitoneal injection induced small, but significant increases in micronuclei in the bone marrow of male F344 rats in two trials (NTP, 2009). However, the substance was found to be negative in a gavage test for micronuclei with male F344/DuCrI rats in a more recent assessment (NTP, 2012). Further tests on micronuclei with mice (B6C3F<sub>1</sub>, Swiss) with gavage or inhalation exposure provided negative results. Comet assays in male rats and female or male mice gave largely negative results in blood, lung, kidney or liver cells. However, the response was weakly positive for male rats in the liver (trend, highest dose) and for female mice in the lung (trend, highest dose) (NTP, 2012). To clarify the DNA damage from reactive oxygen species, Kim *et al.* (2008) performed a “Fragment Length Analysis with Repair Enzyme” (FLARE) test in combination with a Comet assay after subchronic inhalation exposure in hepatocytes and lymphocytes of SD rats. The authors found some indications for oxidative DNA damage from cumene exposure; however, there was no clear duration-response relationship observed and the study is qualified as being insufficient in reporting of methods and results (NTP, 2013).

Analysis of mutations in the cumene-induced lung tumours in mice from the NTP carcinogenicity study (Table 11) found significant increases of *K-ras* and P53- mutations and different types of mutations from cumene exposed mice compared to mutations in spontaneous tumours in the control group. In addition, loss of heterozygosity (LOH) was detected in cumene induced tumours, with no such changes in spontaneous tumours. The authors discuss a (primary or secondary) genotoxic and/or an epigenetic mode of action for the observed changes (Hong *et al.*, 2008; NTP, 2013; Wakamatsu *et al.*, 2008). G→T transversions, as observed predominantly in cumene induced lung tumours, are associated with active oxygen species and are consistent with 8-OH-G adducts produced during oxidative damage to DNA. G→T transversions in *K-ras* codon 12 is the most common mutation detected in human adenocarcinoma (Hong *et al.*, 2008).

The metabolite,  $\alpha$ -methylstyrene, was positive *in vivo* in female mice in normochromic erythrocytes for micronuclei induction (trend, highest dose tested), but neither an increase of micronucleated polychromatic erythrocytes nor genotoxicity were observed in male mice (NTP, 2007). Another recent test on micronuclei formation in male mice bone marrow cells was negative (Rim *et al.*, 2012). There are no *in vivo* data available for the postulated metabolite  $\alpha$ -methylstyrene oxide or for other metabolites of cumene.

In conclusion, there are no data on germ cell mutagenicity from cumene or metabolites. For somatic cells, the vast majority of available tests gave negative results and there are only few indications for a genotoxic potential:

- Some DNA damage in male liver or female mice may not be excluded, as evidenced by recent Comet assay analysis from NTP (2012). It is speculated that this DNA damage may be a secondary genotoxic effect, e.g., due to oxidative damage in target organs,
- The postulated metabolite  $\alpha$ -methylstyrene oxide may be mutagenic; however, the quantitative relevance of this substance for cumene metabolism has not been assessed and the finding is not confirmed by direct observations with cumene,

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<sup>1</sup> Tests for reverse mutations in bacteria mostly lack strains for detecting cross-linking activity (TA102, *E. coli* WP2 strains) and in consequence were rated Reliability 3. But cross-linking activity is not assumed to be critical for cumene and negative results obtained in these tests (e.g. NTP, 2012) are considered meaningful for the assessment of mutagenic effects of cumene in bacteria.

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- Changed profiles and increases of *K-ras* and *p53* mutations in cumene induced lung tumours may either point to mutagenicity of cumene or secondary genotoxicity (e.g., from reactive oxygen species, resulting in genetic instability and/or impairment of repair mechanisms) or epigenetic changes (e.g., from altered histone deacetylase, as discussed in Section 10.7.1).

Conclusions with respect to classification for germ cell mutagenicity are shown in Section 10.6.3.

### 10.6.2 Comparison with the CLP criteria

For potential classification on germ cell mutagenicity, criteria from CLP Regulation (EC, 2017) were applied:

#### a) Comparison with Category 1 criteria

- *The classification in Category 1A is based on positive evidence from human epidemiological studies (EC, 2017)*

There are no epidemiological data to support classification of cumene in Category 1A.

- *The classification in Category 1B is based on positive result(s) from in vivo heritable germ cell mutagenicity tests in mammals (EC, 2017)*

There exist no *in vivo* heritable germ cell mutagenicity tests in mammals for cumene.

- *Classification in Category 1B can also be based on “positive result(s) from in vivo somatic cell mutagenicity tests in mammals, in combination with some evidence that the substance has potential to cause mutations to germ cells. It is possible to derive this supporting evidence from mutagenicity/genotoxicity tests in germ cells in vivo, or by demonstrating the ability of the substance or its metabolite(s) to interact with the genetic material of germ cells” (EC, 2017).*

This criterion is rejected as, for cumene, no primary *in vivo* somatic cell mutagenicity has been demonstrated and, by a weight of evidence approach, there only exists a concern for secondary genotoxicity and epigenetic interactions with DNA. There is not sufficient evidence that cumene interacts with the genetic material of germ cells. For metabolites, there are no studies to indicate that  $\alpha$ -methyl styrene, a confirmed metabolite of cumene, interacts with the genetic material of germ cells. There is insufficient evidence to qualify  $\alpha$ -methyl styrene as an *in vivo* mutagen in somatic cells. The postulated metabolite  $\alpha$ -methyl styrene oxide, which is assumed to be genotoxic in somatic cells (Rosman *et al.*, 1986), has not been shown to interact with the genetic material of germ cells.

Therefore, there is no evidence that the substance has the potential to cause germ cell mutations. Classification in Category 1 is not justified.

#### b) Comparison with Category 2 criteria

- *Classification in category 2 is based on somatic cell mutagenicity tests in vivo, in mammals (ECHA, 2017)*

For cumene primary somatic cell mutagenicity has not been demonstrated *in vivo* or *in vitro*. However, there is some evidence for potential DNA damages from cumene exposure shown by the Comet assay only in specific target tissues in high concentrations from inhalation exposure (NTP, 2012) and from intraperitoneal application in male rat (NTP, 2009). These genotoxic events are not regarded as a mutagenic effect. Mutations have been observed in cumene-induced tumours, but those are regarded as induced via epigenetic or secondary genotoxic in mode of action (Wakamatsu *et al.*, 2008). One of the discussed “mode of action” proposes that epigenetic events from cumene exposure lead to amplifications of pre-existing spontaneous mutations. Potential genotoxic “modes of action” refer to genotoxicity secondary to oxidative stress (Kim *et al.*, 2008; NTP, 2009; Wakamatsu *et al.*, 2008). This interaction is not believed to lead to germ cell mutagenicity without an effect threshold.

- *Classification in category 2 is also based on other in vivo somatic cell genotoxicity, which are supported by positive results from in vitro mutagenicity assays (ECHA, 2017)*

The application of this criterion is rejected, as there are no sufficient indications of *in vitro* mutagenicity for cumene. Mutagenicity, as has been shown for the postulated metabolite  $\alpha$ -methyl styrene oxide (Rosman *et al.*, 1986), has not been evidenced to be relevant for exposures to cumene.

- *This hazard class is primarily concerned with substances that may cause mutations in the germ cells of humans that can be transmitted to the progeny. However, the results from mutagenicity or genotoxicity tests in vitro and in mammalian somatic and germ cells in vivo are also considered in classifying substances and mixtures within this hazard class” (EC, 2017). ECHA (2017) further comments:*

*Thus, classification as a germ cell mutagen (Category 1A, 1B, and 2) classifies for the hazard heritable genetic damage as well as providing an indication that the substance could be carcinogenic.*

### 10.6.3 Conclusion on classification and labelling for germ cell mutagenicity

There is no evidence that cumene is a germ cell mutagen.

Accordingly, a “weight of evidence” approach is taken, as Guidance on the application of CLP criteria specifically requests: “If there is also negative or equivocal data, a weight of evidence approach using expert judgement has to be applied” (ECHA, 2017).

Basically,

- Cumene might have caused some DNA damage in male liver of rats or lungs of female mice, as evidenced by recent Comet assay analysis from NTP (2012). It is speculated that this DNA damage may be a secondary genotoxic effect due to oxidative damage or genomic instability in the target organ (Hong *et al.*, 2008),
- There are some indications of genotoxicity for the metabolite  $\alpha$ -methylstyrene due to increased sister chromatid exchanges in vitro, but those data are not sufficient to classify  $\alpha$ -methylstyrene as a genotoxic substance (EC, 2017),
- the postulated metabolite  $\alpha$ -methylstyrene oxide may be mutagenic,
- some further metabolites from ring-oxidation of cumene are assumed to be reactive,
- changed profiles and increases of K-*ras*, p53 mutations in cumene induced lung tumours may be either point to mutagenicity of cumene or secondary genotoxicity (e.g., from reactive oxygen species, resulting in genetic instability and/or impairment of repair mechanisms).

In conclusion, the evidence for a primarily genotoxic mode of action for cumene carcinogenicity is unlikely.

**Therefore no classification as a germ cell mutagen is warranted for cumene.**

<b>RAC evaluation of germ cell mutagenicity</b>
<p><b>Summary of the Dossier Submitter’s proposal</b></p> <p><i>In vitro</i>, six negative Ames tests, one positive and one negative spot test in bacteria and one negative mutation assay in yeast on cumene were available. Cumene was also considered negative by the DS in two studies for <i>in vitro</i> gene mutation in mammalian cells. The available <i>in vitro</i> cytogenicity test (Putman, 1987a) was negative without metabolic activation, but inconclusive in presence of metabolic activation. A positive result observed in a first unscheduled DNA synthesis (UDS) study (Gulf oil corporation, 1984) was not confirmed in a retest (Curren, 1987). An overall negative outcome was concluded by the DS <i>in vitro</i>.</p>

Four *in vivo* micronucleus tests were available, two were performed in mice (inhalation or gavage administration) and two in rats (intraperitoneal or gavage administration). These studies were considered reliable with limitations and similar to OECD TG except the gavage study in mice which was rated as unreliable (Gulf corporation, 1985b). The justification for the Klimish score 3 was not provided. All tests were negative except the study performed by intraperitoneal route in rats (NTP, 2009).

Three *in vivo* comet assays were reported with cumene (NTP, 2012 and Kim *et al.*, 2008). In the NTP gavage studies in male rats and male and female mice, the results were negative in blood and kidney. Weakly positive results were reported in male rats in the liver and in female mice in the lung. A fragment length analysis with repair enzyme (FLARE) combined with a comet assay in lymphocytes and hepatocytes was performed by Kim *et al.*, 2008 after subchronic inhalation exposure in rats. There were some indications of oxidative DNA damage from cumene exposure, but no dose-response was observed in the study.

Analysis of mutations in the cumene-induced lung tumours in mice of the NTP, 2009 carcinogenicity study found significant increases of *K-ras* and *p53* mutations and different types of mutations in cumene exposed mice compared to mutations in spontaneous tumours of the control group (Hong *et al.*, 2008). In addition, loss of heterozygosity (LOH) was detected in cumene induced tumours, not observed in spontaneous tumours.

Data on metabolites were also retrieved by the DS. The metabolite  $\alpha$ -methylstyrene was negative in an Ames assay and in an *in vitro* chromosomal aberration assay (NTP, 2007). *In vivo*,  $\alpha$ -methylstyrene was weakly positive in female mice but not in male mice in an *in vivo* micronucleus assay following 3-month inhalation exposure (NTP, 2007). Negative results were observed in an *in vivo* single gavage micronucleus study in male mice (Rim *et al.*, 2012).  $\alpha$ -methylstyrene is postulated to be further oxidised to  $\alpha$ -methylstyrene oxide. This metabolite was reported to be positive in an Ames assay (Rosman *et al.*, 1986).

Overall, the DS concluded that most of the data available on cumene are negative and that there are only few indications of a genotoxic potential:

- DNA damage in male liver or lung of female mice. It is speculated by the DS that the DNA damage may be due to oxidative damage in target organs.
- The postulated  $\alpha$ -methylstyrene oxide metabolite may be mutagenic, but the quantitative relevance of this metabolite is unknown and not confirmed by direct observation with cumene.
- Changed profiles and increases of *K-ras* and *p53* mutations in cumene induced lung tumours may either point to mutagenicity of cumene or secondary genotoxicity (e.g. from reactive oxygen species, resulting in genetic instability and/or impairment of repair mechanisms) or epigenetic changes.

Overall, the DS concluded that the CLP criteria for germ cell mutagenicity were not fulfilled as no evidence is available that cumene is a germ cell mutagen and as the evidence for a primarily genotoxic mode of action (MoA) for cumene carcinogenicity is unlikely.

### Comments received during consultation

One MS requested details on the reliability of some studies and highlighted that in some cases, positive intraperitoneal studies may already lead to classification as Muta. 2; H341.

Two industry representatives and two individuals agreed with no classification for germ cell

mutagenicity for cumene. They commented that cumene does not pose a mutagenic hazard. They provided some remarks on the DS's proposal:

- *K-ras* and *p53* reported mutation may be more a resulting effect from rapidly dividing tissues than a cause. Moreover, it may be a consequence of irritation combined with inflammation leading to reactive oxygen species (ROS) generation rather than any direct activity from cumene itself.
- Although a positive *in vivo* micronucleus assay was observed in rats *via* intraperitoneal injection, negative results for clastogenicity/aneugenicity were provided by other studies with more relevant route of exposure.
- The borderline increase in the percentage tail DNA in the *in vivo* comet assay may have been related to random background variations and did not correlate with cumene tumorigenic profile.

## **Assessment and comparison with the classification criteria**

### ***In vitro* results**

Six negative studies for gene mutation in the Ames test were provided on cumene in various vehicles (ethanol, DMSO, pluronic F127 in 50% ethanol). Only one study was rated reliable with limitation (Monsanto Co, 1985). Klimish score in the other studies was either not reliable or not assignable (due to missing strains, inadequate exposure due to volatility or limited data information on the study). Considering the overall database, all strains recommended in OECD TG 471 were tested up to cytotoxic concentration, including strain *E. Coli* WP2. Both the preincubation methods or direct plate incorporation were used. Sealed tubes were used in the NTP, 2012 study. One negative *in vitro* gene mutation study was also available in yeast. A positive and a negative spot test were also reported but the results of these studies are considered of negligible weight as compared to the six negative Ames assays. Overall, RAC agrees with the DS that cumene did not induce gene mutation in bacteria in presence or absence of metabolic activation.

Two studies for gene mutation in mammalian cells were also available (Gulf oil corporation, 1985a; Yang, 1987). The studies were reported to be similar to OECD TG 476. According to NTP, 2013 evaluation, Gulf oil corporation study had to be retested due to variable background and colony forming efficiency. The same limitations were noted in the retest study from Yang, 1987 (see supplemental information – in depth analysis by RAC). Therefore, reliability of these studies is questionable. In addition, due to the limitations of these studies, RAC considers the increase in mutation frequency in presence of metabolic activation observed in both studies inconclusive rather than positive.

One *in vitro* mammalian chromosome aberration test was available with cumene (Putman, 1987a). The study was performed according to OECD TG 473 but some limitations were noted by RAC: limited information of methods and results provided in the CLH dossier and in the ECHA disseminated database, lower number of metaphases analysed compared to current test guideline, only short-term exposure duration, no repeated experiments and cell growth for main experiment was not reported. The study was negative in absence of rat metabolic activation. A statistically significant increase in cells with structural aberrations were reported at the highest dose tested (156 µg/ml) in presence of metabolic activation compared to the vehicle control. The increase was not statistically significant compared to untreated control and was within historical control range of the laboratory. The increase in chromosomal aberration may thus not be toxicologically relevant.

Cumene did not induce unscheduled DNA synthesis (UDS test) *in vitro*. Although positive results were observed in one study, negative results were obtained following retest. Nevertheless, due to its low sensitivity, the UDS test is considered of low weight.

Overall, cumene was not mutagenic in bacteria in presence or absence of metabolic activation. Inconclusive results were obtained for gene mutation in mammalian cells in presence of metabolic activation due to study limitations (see supplemental information – in depth analysis by RAC). Negative results were observed for cytogenicity in mammalian cells in presence or absence of metabolic activation. Nevertheless, some limitations were noted in the study.

### ***In vivo results***

In mice, negative results were obtained in a 3-month inhalation exposure bone marrow micronucleus study up to 500 ppm in females and 1000 ppm in males. In male rats, positive results were obtained following intraperitoneal route (NTP, 2009). To clarify the positive result, micronucleus studies in peripheral blood were performed by gavage in male rats up to 800 mg/kg and male and female mice up to 1000 and 1250 mg/kg, respectively during 4 consecutive days (NTP, 2012). The studies were negative. Proof of exposure was observed in rats. The positive results obtained following intraperitoneal route of exposure is of low weight compared to the three negative studies using relevant routes of human exposure (oral, inhalation). Nevertheless, as this is the only micronucleus assay via the intraperitoneal route, the finding cannot be completely neglected. The studies were equivalent to OECD TG 474. Overall, cumene did not induce damage at chromosomal levels in rats and mice *in vivo*.

An *in vivo* rodent comet assay was also performed on the same male rats and male and female mice that were evaluated for the micronucleus endpoint (NTP, 2012). The study was equivalent to OECD TG 489. The substance was tested via gavage application in corn oil. There were two main limitations in this study:

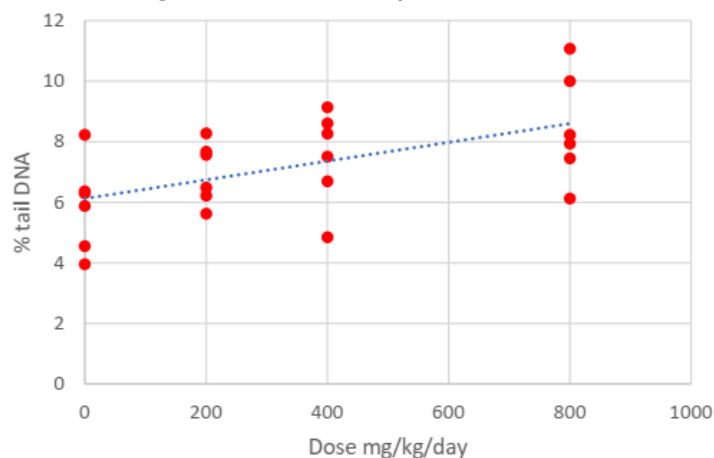
- Historical negative and positive controls were not provided. High variability could be an issue in comet assays.
- No information on cytotoxicity in tested tissues was provided. Histopathology at the top dose in the organs were not available. Nevertheless, Hedgehog cells were excluded from analysis. Moreover, in the 2-week and 13-week inhalation studies in rats and mice (NTP, 2009), only liver weight changes were noted without concomitant necropsy findings up to maximum tolerated dose. Therefore, liver toxicity is not expected to be an issue in the interpretation of the comet assay. No effects on lungs were noted in the 13-week study.

The assay was performed in blood leukocytes, liver, lung and kidney. Negative results were observed in male mice (all tissues), male rat leukocytes, lung and kidney and in female mice leukocytes, liver and kidney. An increase in the % Tail DNA was observed in male rat liver (statistically significant only at the top dose) and female mice lung. The table below reports the results of the comet assay in lung and liver (From NTP, 2012 report).

Dose (mg/kg)	% Tail DNA (mean $\pm$ SD)		
	Female mice, lung	Male mice, lung	Male rat, liver
0	6.8 $\pm$ 0.3	11.9 $\pm$ 1.2	5.9 $\pm$ 0.6
250	7.3 $\pm$ 0.6	12.2 $\pm$ 0.8	7.0 $\pm$ 0.4
500	7.8 $\pm$ 0.7	13.7 $\pm$ 1.3	7.5 $\pm$ 0.6
1000	8.7 $\pm$ 0.7*	13.0 $\pm$ 1.3	8.5 $\pm$ 0.7**
EMS	25 $\pm$ 1.2***	16.7 $\pm$ 1.0**	37 $\pm$ 0.6***

\* $p \leq 0.05$ ; \*\* $p \leq 0.01$ ; \*\*\* $p \leq 0.001$ ; EMS: Ethylmethane sulfonate

The figure below described individual data for % tail DNA in male rat liver with trend line (as provided by one individual during the consultation).



There are three main criteria to conclude on a positive result in a comet assay: dose-relation, statistical significance and the biological relevance (increases above concurrent historical negative control range). In male rat liver and female mice lung, the increase in % tail DNA at the top dose was statistically significant compared to concurrent negative control. Moreover, the increases were also dose-related (trend test). The distribution of the historical negative control data was not provided in the NTP report. Therefore, there are some uncertainties on the toxicological significance of the increase in the % tail DNA in mouse female lung and male liver. According to the DS in the response to comment, an additional comet assay for styrene-acrylonitrile trimer was performed in 2012 by NTP in juvenile F344 rats (3-4 week of age). A higher DNA damage background in liver was found ( $10.605 \pm 2.951$  %). Nevertheless, RAC notes that DNA background may differ between F344 young adult rats used in the cumene study (8-weeks of age). Moreover, the high variability observed in the styrene-acrylonitrile trimer study (individual values between 5.3 and 21.9%) was not observed in the case of cumene. Indeed, the variability in the controls and treated groups was similar based on standard variation. Therefore, there are no data suggesting that the current control results for liver would not be reliable. Regarding the lung tissue, higher background values were obtained in negative control and lower background values in positive controls male mice. Variability in background values were thus observed in this tissue.

### **Genotoxicity of cumene metabolites**

$\alpha$ -methyl-styrene (AMS) did not induce gene mutation in Ames test (TA 102 and *E. coli* not tested) and chromosomal aberration *in vitro* in mammalian cells. No *in vitro* test for gene mutation in mammalian cells was available. Positive effects in sister chromatid exchange (SCE) *in vitro* were observed in two studies (NTP, 2007; Norppa and Vainio, 1983). Nevertheless, SCE assay is considered to have a lower weight than the negative *in vitro* cytogenicity test. The putative metabolite  $\alpha$ -methylstyrene oxide was reported to be mutagenic in an Ames test (TA 100) in the CLH dossier.

*In vivo*,  $\alpha$ -methylstyrene was positive in female mice for micronuclei induction (trend, highest dose tested) following 3-month inhalation exposure, but was negative in male mice (NTP, 2007). Another recent test on micronuclei formation in male mice bone marrow cells was negative following single administration by gavage (Rim *et al.*, 2012). There are no *in vivo* data available for the postulated metabolite  $\alpha$ -methylstyrene oxide. No data were available on other metabolites of cumene.



RAC notes that the pathways leading to reactive metabolism were only minor pathways of the substance (not fully quantified).

### **Mechanism of genotoxicity**

Some studies investigated the genotoxicity MoA of cumene.

In Kim *et al.*, 2008 DNA damage from reactive oxygen species was measured in rats that were treated with cumene by inhalation at doses up to 800 ppm for up to 13 weeks using fragment length analysed with repair enzyme (FLARE) formamidopyrimidine (Fpg)/endonuclease III (Endo III) in conjunction with comet assay. Based on the limitations reported by the DS, RAC considered the study not adequate for the evaluation of cumene (inadequate reporting, unacceptable controls, and inappropriate statistical analysis).

In the published study from Hong *et al.*, 2008, point mutations were evaluated in the *K-ras* (exon 1 and 2) and *p53* genes (exons 5 to 8) in a subset of lung neoplasms observed in the carcinogenicity studies (NTP, 2009). LOH was also analysed at the p16 locus in chromosome 16 and near the *K-ras* gene on chromosome 6. A significant dose-dependent increase of *K-ras* and *p53*- mutations from cumene exposed mice compared to mutations in spontaneous tumours in the control group were observed. *K-ras* codon 12 G to T transversion and *K-ras* codon 61 A to G transitions clearly differs from untreated mice (0.008% vs 2%). The table below presents the *K-ras* and *p53* mutations in lung neoplasms of mice in the two-year study of cumene (Hong *et al.*, 2008).

Treatment (ppm)	Activate <i>K-ras</i> (%)	Activate <i>p53</i> (%)
0 (Concurrent control)	1/7 (14)	0/7 (0)
0 (Historical control)	33/117 (28)	Not provided
Cumene (total of neoplasm)	45/52 (87)	27/52 (52)
125	1/4 (25)	0/4 (0)
250	10/13 (77)	5/13 (38)
500	17/18 (94)	11/18 (38)
1000	17/17 (100)	11/17 (65)

According to the authors, the mutation observed in the *p53* genes (Exon 5 and 7 only, no mutation detected at exon 6 and 8) was clearly induced by cumene as this mutation was not detected in spontaneous tumours. No differences in mutation spectrum was observed between adenomas and carcinomas. Mutations were higher in males than in females. In addition, cumene-induced lung carcinomas showed LOH on chromosome 4 near the *p53* gene (13%) and on chromosome 6 near the *K-ras* gene (12%). No LOH was observed in spontaneous carcinomas or in normal lung tissues examined. The authors concluded that direct and indirect DNA damage may have contributed to the mutations. Direct DNA adducts and subsequent point mutation may have been caused by reactive metabolites of cumene. Indirect damage from oxidative stress may also have contributed to the mutations. G to T transversion are consistent with 8-OH-G adducts produced during oxidative damage. The authors concluded that the patterns of *K-ras* and *p53* mutations identified in the cumene-induced lung tumours suggest that DNA damage and genomic instability may be the contributing factors to the mutation profile and development of lung cancer in mice.

### **Comparison with criteria**

**In conclusion**, there are no human data in the literature, and based on the animal data available, there is no concrete evidence that cumene is mutagenic to germ cells or that it

distributes to the reproductive tissues. Therefore, the criteria to classify a substance as a germ cell mutagen in Category 1B according to the CLP criteria are not met.

The classification in category 2 is based on positive evidence obtained from somatic cell mutagenicity tests *in vivo* in mammals or other *in vivo* somatic cell genotoxicity tests which are supported by positive results from *in vitro* mutagenicity assays.

Cumene did not induce damage at chromosomal levels *in vitro* in mammalian cells. The positive results observed in the intraperitoneal micronucleus test *in vivo* may indicate an intrinsic genotoxic potential of the substance. It is further noted that a weakly positive result in females was also obtained in a micronucleus test after 3-month inhalation exposure to  $\alpha$ -methyl styrene in mice (but not in males). Nevertheless, the negative results observed in the *in vivo* micronucleus assays with cumene using relevant route of exposure (inhalation, gavage) in mice and rats decrease the concern.

As regards gene mutations, gene mutation assays in bacteria were negative. *In vitro* gene mutation assays in mammalian cells were inconclusive due to study deficiencies. *In vivo*, the positive comet assays in liver of male rats and lung of female mice performed according to a relevant route of exposure (inhalation) indicate potential of cumene to induce gene mutation in somatic cells. As discussed above, the absence of historical control range raises some uncertainties on the biological relevance of the results of the comet assay. Hong *et al.*, 2008 study showed that in mice lung tumours induced by cumene increase in point mutation in *K-ras*, and *p53* mutations in specific investigated exons were observed. Although the results may be explained by direct mutagenicity through reactive metabolites, the substance may also act through an indirect MoA *via* oxidative damages. Epigenetic changes may also be involved (Wakamatsu *et al.*, 2008). They are discussed in more detail in the carcinogenicity section under "Assessment and comparison with the classification criteria". Nevertheless, the analysis was only performed on a subset of lung tumour tissues leading to some uncertainties on the results. RAC also notes that except for mice lungs, the positive results of the comet assay were not fully consistent with the tumorigenic profile of cumene in rats (kidney and respiratory epithelium tumours) and mice (lung and liver tumours). Nevertheless, the difference in the route of exposure between the studies (oral vs inhalation) make the comparison difficult.

Overall, although a weak genotoxic potential of cumene cannot be excluded, RAC agrees with the DS that the criteria for Germ cell mutagen in category 2 (H341) are not fulfilled. **No classification for germ cell mutagenicity is warranted for cumene.**

### **Supplemental information - In depth analyses by RAC**

Two studies for gene mutation in mammalian cells were available. The studies were reported to be similar to OECD TG 476 and GLP-compliant. In view of the positive comet assays obtained *in vivo*, the results of these studies may be important in the overall weight of evidence analysis. Therefore, an in-depth analysis by RAC of these two studies is provided below.

In the first study (Gulf oil corporation, 1985a), cumene was tested in pluronic polyol in 50% ethanol up to 175  $\mu\text{g}/\text{ml}$  with and without metabolic activation. Positive results observed at 175  $\mu\text{g}/\text{ml}$  were not repeated in a second experiment. RAC has not enough details on methods and results for an independent assessment of the reliability of the study. Nevertheless, according to NTP, 2013 evaluation, this study had to be retested due to variable background and colony forming efficiency. Therefore, the acceptability of this study

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is questionable.

In the retest study (Yang, 1987), cumene was tested up to 225 µg/ml in CHO cells with and without S9 mix. RAC notes the following limitations in the test method (based on study summary provided in the CLH and ECHA disseminated website):

- the solvent used was not well-established (pluronic polyol in 50% ethanol). Nevertheless, mutation frequency obtained with the solvent were in the range of the untreated controls of the study and within the historical control range of the laboratory for well-established solvent. Therefore, the negative control could be considered acceptable;
- No historical control data for untreated control was provided.
- Only two dose levels were analysed instead of the four recommended in the OECD TG due to cytotoxicity at  $\geq 125$  µg/ml (relative cloning efficiency <10%) in the absence of S9 mix. Also, only three dose levels were analysed in the presence of metabolic activation (100, 125, 225 µg/ml).
- Cytotoxicity was not dose-related as the substance was too toxic to clone at 150, 175 and 200 µg/ml but cytotoxicity was 40% of control at 225 µg/ml. Therefore, in this study, relative cloning efficiency was variable.
- No statistical analysis was performed;
- Difficulties in dosing the substance (due to its viscosity) was noted by the authors of the study leading to difficulties in reproducibility between replicates.

No increase in mutation frequency was noted in absence of S9. In presence of S9 an increase in mutation frequency was noted at 225 µg/ml. In experiment B, the increase was above the range of spontaneous mutant frequency of  $5 - 20 \times 10^{-6}$  reported in the OECD TG and might therefore be of biological relevance (see detailed results in the table below as provided in the ECHA disseminated website). No dose-relation was noted but the low number of analysed concentration and the uncertainties in dosing may explain the absence of dose-relation. Overall, due to the above limitations and uncertainties in dosing and considering the increase in mutation frequency in presence of S9, the results of the study are inconclusive.

Treatment (Cumene µg/ml)	Experiment A		Experiment B	
	Relative cloning efficiency	Mutants/10 <sup>6</sup> clonable cells	Relative cloning efficiency	Mutants/10 <sup>6</sup> clonable cells
Untreated control	95%	4.8	106%	15.5
Solvent	100%	6.8	100%	1.7
225 µg/ml	40%	10.1	40%	27.6
125 µg/ml	60%	2.3	51%	12.9
100 µg/ml	89%	3.5	103%	19.6
BaP – solvent	25%	326	31%	348
BaP + solvent	31%	324	32%	330

## 10.7 Carcinogenicity

**Table 13: Summary table of animal studies on carcinogenicity (overall rates according to NTP, additional information on historical control data is available in Annex I)**

Method, guideline, deviations if any, species, strain, sex, no/group	Test substance, dose levels, duration of exposure	Results	Reference
<p>2-year carcinogenicity study according to OECD 451 in Mice, B6C3F<sub>1</sub> (♂)</p> <p>50 males per concentration group</p> <p>GLP: according to FDA Good Laboratory Practice Regulations (21 CFR, Part 58)</p> <p>Reliability according to disseminated database: 2</p> <p>Reliability according to authors of this evaluation: 1</p>	<p>Cumene</p> <p>Purity: 99.9 %</p> <p>Inhalation exposure:</p> <p>0, 250, 500, and 1000 ppm</p> <p>(0, 1225, 2450, 4900 mg/m<sup>3</sup>)</p> <p>6 h/d plus T<sub>90</sub> (12 min), 5 d/w, 105 w</p>	<p><b><u>Lung</u></b></p> <p>Alveolar epithelium, bronchiole, metaplasia<sup>a</sup>: 5/50, 43/50**, 42/50**, 39/50**</p> <p>Bronchiole, hyperplasia: 0/50, 11/50**, 17/50**, 18/50**</p> <p>Alveolar/bronchiolar adenoma, multiple: 1/50, 12/50**, 15/50**, 20/50**</p> <p>Alveolar/bronchiolar adenoma (includes multiple)<sup>b</sup>: 13/50 P&lt;0.001<sup>c</sup>, 31/50***, 31/50***, 29/50***</p> <p>Alveolar/bronchiolar carcinoma, multiple: 0/50, 8/50**, 20/50**, 17/50**</p> <p>Alveolar/bronchiolar carcinoma (includes multiple<sup>b</sup>): 9/50 P&lt;0.001<sup>c</sup>, 19/50*, 32/50***, 33/50***</p> <p>Alveolar/bronchiolar adenoma or carcinoma<sup>b,d</sup>: 19/50 P&lt;0.001<sup>c</sup>, 38/50***, 42/50***, 43/50***</p> <p><b><u>Liver</u></b></p> <p>Eosinophilic foci<sup>a</sup>: 6/50, 5/50, 16/50**, 14/50*</p> <p>Hepatocellular adenoma, multiple: 17/50, 20/50, 22/50, 26/50</p> <p>Hepatocellular adenoma (includes multiple): 34/50, 33/50, 37/50, 35/50</p> <p>Hepatocellular carcinoma, multiple: 3/50, 1/50, 4/50, 7/50</p> <p>Hepatocellular carcinoma (includes multiple): 13/50, 18/50, 21/50, 17/50</p> <p>Hepatocellular adenoma or carcinoma<sup>b,e</sup>: 40/50 P=0.250<sup>c</sup>, 42/50, 43/50, 41/50</p> <p><b><u>Hemangiosarcoma</u></b></p> <p>Hemangiosarcoma, spleen<sup>b,f</sup>: 0/50 P=0.002<sup>c</sup>, 0/50, 0/49, 4/50*</p> <p>Hemangiosarcoma, all organs<sup>g,h</sup>: 0/50 P=0.015<sup>c</sup>, 1/50, 2/50, 4/50*</p> <p><b><u>Thyroid gland</u></b></p> <p>Follicular cell, hyperplasia<sup>a</sup>: 7/50, 7/50, 7/49, 11/50</p> <p>Follicular cell, adenoma<sup>b,i</sup>: 0/50 P=0.010<sup>c</sup>, 0/50, 0/49, 3/50<sup>j</sup></p>	<p>(NTP, 2009)</p> <p>Study also reported in (ECHA Dissemination, 2018, carcinogenicity, #002, key)</p>
2-year	Cumene	<b><u>Lung</u></b>	(NTP, 2009)

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Method, guideline, deviations if any, species, strain, sex, no/group	Test substance, dose levels duration of exposure	Results	Reference
<p>carcinogenicity study according to OECD 451 in Mice, B6C3F<sub>1</sub> (♀)</p> <p>50 females per concentration group</p> <p>GLP: according to FDA Good Laboratory Practice Regulations (21 CFR, Part 58)</p> <p>Reliability according to disseminated database: 2</p> <p>Reliability according to authors of this evaluation: 1</p>	<p>Purity: 99.9 %</p> <p>Inhalation exposure: 0, 125, 250, and 500 ppm</p> <p>(0, 1225, 2450, 4900 mg/m<sup>3</sup>)</p> <p>6 h/d plus T<sub>90</sub> (12 min), 5 d/w, 105 w</p>	<p>Alveolar epithelium, bronchiole, metaplasia<sup>a</sup>: 0/50, 42/50**, 49/50**, 47/50**</p> <p>Bronchiole, hyperplasia: 0/50, 17/50**, 10/50**, 14/50**</p> <p>Alveolar/bronchiolar adenoma, multiple: 0/50, 13/50**, 20/50**, 30/50**</p> <p>Alveolar/bronchiolar adenoma (includes multiple)<sup>b</sup>: 1/50 P&lt;0.001<sup>c</sup>, 26/50***, 36/50***, 38/50***</p> <p>Alveolar/bronchiolar carcinoma, multiple: 0/50, 6/50*, 7/50**, 19/50**</p> <p>Alveolar/bronchiolar carcinoma (includes multiple)<sup>b</sup>: 3/50 P&lt;0.001<sup>c</sup>, 16/50***, 20/50***, 34/50***</p> <p>Alveolar/bronchiolar adenoma or carcinoma<sup>b,k</sup>: 4/50 P&lt;0.001<sup>c</sup>, 31/50***, 42/50***, 46/50***</p> <p><b>Liver</b></p> <p>Eosinophilic focus<sup>a</sup>: 8/50, 11/50, 7/50, 14/50</p> <p>Hepatocellular adenoma, multiple: 9/50, 13/50, 9/50, 10/50</p> <p>Hepatocellular adenoma (includes multiple)<sup>b</sup>: 18/50 P=0.040<sup>c</sup>, 23/50, 27/50<sup>l</sup>, 29/50*</p> <p>Hepatocellular carcinoma, multiple: 2/50, 1/50, 2/50, 0/50</p> <p>Hepatocellular carcinoma (includes multiple): 10/50, 7/50, 6/50, 12/50</p> <p>Hepatocellular adenoma or carcinoma<sup>b,m</sup>: 25/50 P=0.024<sup>c</sup>, 26/50, 29/50<sup>l</sup>, 36/50*</p>	<p>Study also reported in (ECHA Dissemination, 2018, carcinogenicity, #002, key)</p>
<p>2-year carcinogenicity study according to OECD 451 in Rat, F344/N (♂)</p> <p>50 males per concentration group</p> <p>GLP: according to FDA Good Laboratory Practice Regulations (21 CFR, Part 58)</p> <p>Reliability</p>	<p>Cumene</p> <p>Purity: 99.9 %</p> <p>Inhalation exposure: 0, 250, 500, and 1000 ppm</p> <p>(0, 1225, 2450, 4900 mg/m<sup>3</sup>)</p> <p>6 h/d plus T<sub>90</sub></p>	<p><b>Nose</b></p> <p>Olfactory epithelium, hyperplasia, basal cell<sup>a</sup>: 0/50, 19/50**, 27/49**, 26/50**</p> <p>Respiratory epithelium, hyperplasia: 0/50, 15/50**, 16/49**, 23/50**</p> <p>Goblet cell, hyperplasia: 3/50, 11/50*, 7/49, 5/50</p> <p>Glands, respiratory epithelium, adenoma: 0/50, 0/50, 1/49, 0/50</p> <p>Respiratory epithelium, adenoma, multiple: 0/50, 1/50, 2/49, 6/50*</p> <p>Respiratory epithelium, adenoma (includes multiple and all sites)<sup>b,n</sup>: 0/50 P=0.004<sup>c</sup>, 7/50**, 18/49***, 10/50***</p> <p><b>Kidney</b></p> <p>Renal tubule, hyperplasia<sup>a</sup>: 0/50, 3/50, 8/50**, 6/50*</p> <p>Papilla, mineralisation: 5/50, 35/50**, 44/50**, 41/50**</p> <p>Pelvis, transitional epithelium, hyperplasia: 3/50, 5/50, 14/50**,</p>	<p>(NTP, 2009)</p> <p>Study also reported in (ECHA Dissemination, 2018, carcinogenicity, #001, key)</p>

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Method, guideline, deviations if any, species, strain, sex, no/group	Test substance, dose levels duration of exposure	Results	Reference
<p>according to disseminated database: 2</p> <p>Reliability according to authors of this evaluation: 1</p>	<p>(12 min), 5 d/w, 105 w</p>	<p>15/50**</p> <p>Nephropathy: 47/50, 47/50, 47/50, 50/50</p> <p>Renal tubule, adenoma<sup>b</sup>: 1/50 P=0.219<sup>c</sup>, 4/50, 5/50, 4/50</p> <p>Renal tubule, carcinoma, bilateral: 0/50, 0/50, 1/50, 0/50</p> <p>Renal tubule, carcinoma (includes bilateral)<sup>b</sup>: 1/50 P=0.180<sup>c</sup>, 1/50, 3/50, 3/50</p> <p>Renal tubule, adenoma or carcinoma<sup>b,o</sup>: 2/50 P=0.087<sup>c</sup>, 5/50, 8/50*, 7/50</p> <p>Renal tubule, lipoma: 1/50, 0/50, 0/50, 1/50</p> <p><b>Testis</b></p> <p>Interstitial cell, hyperplasia<sup>a</sup>: 12/50, 18/50, 19/50, 9/50</p> <p>Bilateral interstitial cell, hyperplasia: 0/50, 0/50, 0/50, 1/50</p> <p>Interstitial cell, adenoma: 18/50, 14/50, 13/50, 9/50</p> <p>Bilateral interstitial cell, adenoma: 18/50, 24/50, 27/50, 37/50</p> <p>Interstitial cell, adenoma (includes bilateral)<sup>b,p</sup>: 36/50 P=0.006<sup>c</sup>, 38/50, 40/50, 46/50**</p>	
<p>2-year carcinogenicity study according to OECD 451 in Rat, F344/N (♀)</p> <p>50 females per concentration group</p> <p>GLP: according to FDA Good Laboratory Practice Regulations (21 CFR, Part 58)</p> <p>Reliability according to disseminated database: 2</p> <p>Reliability according to authors of this</p>	<p>Cumene</p> <p>Purity: 99.9 %</p> <p>Inhalation exposure: 0, 250, 500, and 1000 ppm</p> <p>(0, 1225, 2450, 4900 mg/m<sup>3</sup>)</p> <p>6 h/d plus T<sub>90</sub> (12 min), 5 d/w, 105 w</p>	<p><b>Nose</b></p> <p>Olfactory epithelium, hyperplasia, basal cell<sup>a</sup>: 0/50, 14/48**, 25/50**, 31/50**</p> <p>Respiratory epithelium, hyperplasia: 0/50, 0/48, 4/50, 6/50*</p> <p>Respiratory epithelium, adenoma<sup>d</sup>: 0/50 P=0.320<sup>c</sup>, 5/48*, 4/50, 3/50</p> <p><b>Kidney</b></p> <p>Nephropathy<sup>a</sup>: 38/50, 37/50, 41/50, 44/50</p>	<p>(NTP, 2009)</p> <p>Study also reported in (ECHA Dissemination, 2018, carcinogenicity, #001, key)</p>

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Method, guideline, deviations if any, species, strain, sex, no/group	Test substance, dose levels duration of exposure	Results	Reference
evaluation: 1			
Footnotes:			
Significant difference from chamber control group determined by Poly-3 test: * $P \leq 0.05$ , ** $P \leq 0.01$ , *** $P \leq 0.001$ .			
<p><sup>a</sup> Overall rate, number of animals with lesion per number of animals examined microscopically</p> <p><sup>b</sup> Overall rate, number of animals with neoplasms per number of animals with lung/liver/tissue/thyroid gland/nose/kidney/testis examined microscopically (only with regard to the organ under investigation)</p> <p><sup>c</sup> For chamber control incidence, P value is given that is associated with the trend test determined by Poly-3 test (accounts for differential mortality in animals that do not reach terminal sacrifice).</p> <p><sup>d</sup> Historical incidence for inhalation studies: 146/449 (32.5% <math>\pm</math> 5.9%), range 26%-44%</p> <p><sup>e</sup> Historical incidence for inhalation studies: 264/449 (58.8% <math>\pm</math> 9.6%), range 50%-80%</p> <p><sup>f</sup> Historical incidence for 2-year inhalation studies with chamber control groups (mean <math>\pm</math> standard deviation): 6/444 (1.4% <math>\pm</math> 1.5%), range 0%-4%</p> <p><sup>g</sup> Overall rate, number of animals with neoplasm per number of animals necropsied</p> <p><sup>h</sup> Historical incidence for inhalation studies: 21/450 (4.7% <math>\pm</math> 3.7%), range 0%-12%</p> <p><sup>i</sup> Historical incidence for 2-year inhalation studies with chamber control groups (mean <math>\pm</math> standard deviation): 5/441 (1.1% <math>\pm</math> 2.0%), range 0%-6%</p> <p><sup>k</sup> Historical incidence for inhalation studies: 34/449 (7.6% <math>\pm</math> 4.0%), range 2%-14%</p> <p><sup>l</sup> One animal with adenoma also had hepatoblastoma</p> <p><sup>m</sup> Historical incidence for inhalation studies: 145/447 (32.4% <math>\pm</math> 8.8%), range 22%-50%</p> <p><sup>n</sup> Historical incidence for 2-year inhalation studies with chamber control groups (mean <math>\pm</math> standard deviation): 1/447 (0.2% <math>\pm</math> 0.7%), range 0%-2%</p> <p><sup>o</sup> Historical incidence for inhalation studies: 6/449 (1.3% <math>\pm</math> 1.4%), range 0%-4%</p> <p><sup>p</sup> Historical incidence for 2-year inhalation studies with chamber control groups (mean <math>\pm</math> standard deviation): 345/449 (76.8% <math>\pm</math> 5.9%), range 66%-84%</p> <p><sup>q</sup> Historical incidence for inhalation studies: 0/496</p>			

**Table 14: Summary table of human data on carcinogenicity**

Type of data/report	Test substance,	Relevant information about the study (as applicable)	Observations	Reference
No human data on cumene are available, which are relevant for carcinogenicity classification assessment				

**Table 15: Summary table of other studies relevant for carcinogenicity**

Type of study/data	Test substance,	Relevant information about the study (as applicable)	Observations	Reference
Transformation assay; BALB/3T3 cells; without activation  Certified GLP compliance, but stability of test and control substances	Cumene  Purity: 99.7%  Vehicle: F127	3 day exposure of BALB/3T3 cells to 0, 50-200 $\mu$ g/ml (4 doses) with survival of 102%, 87%, 19% and 4%, respectively. Positive control: 3-methylcholanthrene  Retest based on Gulf Oil	No increase of Type III (or Type II) foci in cumene treated cells compared to vehicle treated cells.  → negative	(Putman, 1987b)  Study is also reported in (ECHA Dissemination, 2018, genetic toxicity, in vitro, #006,

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Type of study/data	Test substance,	Relevant information about the study (as applicable)	Observations	Reference
<p>have not been tested</p> <p>Reliability according to disseminated database: 1</p> <p>Reliability according to authors of this evaluation:3</p>		<p>Corporation (1984)</p>		<p>supporting)</p>
<p>Transformation assay; BALB/3T3 cells; without activation</p> <p>Compliance with GLP stated by authors</p> <p>Reliability according to disseminated database: study not reported</p> <p>Reliability according to authors of this evaluation:3</p>	<p>Cumene</p> <p>Purity: not disclosed, but may be requested from the sponsor of the study</p> <p>Vehicle: Pluronic F68 Polyol</p>	<p>Mouse embryo cells (BALB/3T3), dosing: (untreated, vehicle, 5-90 µg/ml, 4 doses, positive control), highest dose (90 µg/ml) extremely toxic (eliminated from study). Colony forming efficiency reduced at 60 µg/ml</p> <p>Positive control: 3-methylcholanthrene</p>	<p>Transformation (type III foci) observed at 60 µg/ml.</p> <p>→ positive</p> <p>Re-tested by Putman (1987b)</p>	<p>(Gulf Oil Corporation, 1984)</p>
<p>Mechanistic study:</p> <p>Gene expression analysis</p> <p>Reliability according to disseminated database: study not reported</p> <p>Reliability according to authors of this evaluation:1</p>	<p>Cumene</p> <p>Purity: &gt;99.9%; no impurities &gt; 0.1% observed</p> <p>(from NTP, 2009)</p>	<p>8/23 tissue from cumene induced lung tumours in mice (→NTP, 2009, Table 13) plus 4 normal lung tissues (untreated mice) were selected for gene expression analysis. Lung tumour tissues were chosen based on the absence of necrosis and inflammatory cell infiltration.</p> <p>In a microarray analysis gene expression changes were separated into 3 groups: control lung tissue, tumours with <i>K-ras</i> mutations, tumours without <i>K-ras</i> mutations.</p> <p>Specific analysis focused on gene expression linked to</p>	<p>281 Genes different between normal lung and tumours without <i>K-ras</i>; 627 genes differed between normal lung and tumours with <i>K-ras</i> mutation. <i>K-ras</i></p> <p>66 genes were differently expressed between tumours with <i>K-ras</i> and tumours without <i>K-ras</i> or normal lung tissue.</p> <p><u>Gene expression profile of cumene-induced lung tumours linked the MAPK signalling pathway:</u></p> <p>Many of the significantly altered genes in cumene-induced lung tumours were associated with the MAPK signalling pathway. The majority of genes associated with MAPK pathway were significantly altered only in tumours with <i>K-ras</i> mutations (genes known to promote MAPK activation, genes activated by MAPK signalling, genes involved in the inactivation of MAPK</p>	<p>(Wakamatsu <i>et al.</i>, 2008)</p>



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Type of study/data	Test substance,	Relevant information about the study (as applicable)	Observations	Reference
		MAP kinase (MAPK) signalling pathway and to potential histone modification due to histone deacetylases (HDACs) activity changes, which may influence DNA unfolding and transcription.	<p>pathway were downregulated)</p> <p><u>Some genes linked to tumour suppression, invasion and metastasis were only significantly altered in cumene induced tumours with K-ras mutations.</u></p> <p><u>Gene expression profile of cumene-induced lung tumours linked to histone modification:</u></p> <p>Genes associated with the HDAC complex were significantly altered in tumours; K-ras mutation status of the tumours appeared to correlate with upregulated genes. The HDAC complex has been shown to play a role in human cancer.</p> <p>Both, genetic and epigenetic factors may contribute to cumene-induced lung cancer. Epigenetic alterations in gene expression are likely to be involved in cumene-induced lung neoplasms.</p>	

### 10.7.1 Short summary and overall relevance of the provided information on carcinogenicity

There are no human data available for assessment of the carcinogenic hazard of cumene.

From the data presented in Table 13 cumene is carcinogenic in experimental animals. The effect is more pronounced in mice after chronic inhalation exposure, with

- significantly increased lung alveolar/bronchial adenoma or carcinoma (combined) in male and female mice,
- significantly increased liver adenoma or carcinoma (combined) in female mice,
- slight increase of renal tubular cell adenoma or carcinoma (combined) in male rats, significantly increased nose adenoma in the respiratory epithelium in male rats
- various further tumour sites with increased tumour incidence at the highest exposure concentration, which, however, are not in focus for a more detailed analysis (rat: adenoma in testes; male mice: haemangiosarcoma in spleen and in all organs (combined), follicular-cell adenoma of the thyroid gland).

Each observed tumour type will be discussed separately, taking into account available information on the Mode of Action (MoA). However, even if each tumour site is discussed *per se*, the final conclusion (see Section 10.7.3) also needs to consider that cumene appears to be a carcinogen in experimental mice as a clear evidence for a carcinogenic effect is observed for lung tumours in mice.

#### 10.7.1.1 Lung tumours in B6C3F<sub>1</sub> mice

In the study by NTP (2009) lung tumours were statistically increased in mice, as evidenced by alveolar/bronchiolar adenoma or carcinoma in males (19/50, 38/50\*\*\*, 42/50\*\*\*, 43/50\*\*\* (\*\*\*) p≤0.001; P for trend: P<0.001) and in females (4/50, 31/50\*\*\*, 42/50\*\*\*, 46/50\*\*\*, P for trend: P<0.001), for further

explanation on the data, see Table 13). Even if some of the background dose response data are not strictly monotonously increasing with dose (e.g., male bronchiole metaplasia (0/50; 42/50; 49/50; 47/50)), this does not invalidate this overall clear evidence for an exposure related causal effect in both sexes. The question of human relevance is separately discussed below.

### a) Genotoxicity as MoA

As discussed in Section 10.6.3, a direct interaction of cumene or metabolites with DNA (primary genotoxicity) is not a likely MoA for lung tumours. However, in a Comet assay significant increases in DNA damage in lungs of female mice were observed *in vivo* (only weak, but significant increase at the highest dose tested and significant trend) (NTP, 2012).

In addition, *K-ras* mutations and *p53* mutations were evaluated in spontaneously occurring and cumene-induced tumours in mice (see Table 11 for details). The data show differences in the incidence of *K-ras* mutations between cumene-induced (87%) and spontaneous lung tumours (14%) and historical controls (28%). The type of *K-ras* mutations differed between tumours from exposed and unexposed animals (e.g., predominant *K-ras* mutations in lung tumours from cumene-exposed mice were codon 12 G→T transversions (36% vs. 18% in historical controls); in contrast, codon 12 G→A transitions (42%) was the most common mutation in spontaneous lung tumours). Mutations in the *p53* tumour suppressor gene were not observed in spontaneous lung tumours of the concurrent controls, but were evident in 52% of the cumene-induced lung tumours (no historical control data provided). Furthermore, a loss of heterozygosity (LOH) occurred in cumene-induced mouse lung tumours, but not in spontaneous tumours in control mice (Hong *et al.*, 2008). The *K-ras* and *p53* mutations showed a dose-dependent increase (total of all exposed groups) and similar mutation rates were reported for adenomas and carcinomas. However, as such mutations were more prevalent in exposed males than females, this observation would not be in accordance with the elevated female sensitivity compared to males for cumene-induced lung tumours.

As a further possible MoA, leading to secondary genotoxicity, induction of reactive oxygen species (ROS) by cumene is discussed. ROS-dependent changes including oxidative damage could explain the positive results in the Comet assay (Hong *et al.*, 2008). However, no specific studies with cumene providing evidence for this potential MoA (e.g., analyses of oxo-deoxyguanosine adducts) were identified.

The observed increase in *K-ras* and *p53* mutations may also be caused by an epigenetic MoA: cumene is discussed to cause growth advantage for preneoplastic or neoplastic cells carrying these (possibly spontaneous) mutations and “these molecular changes may be an effect rather than a cause” of the multistage carcinogenic process (NTP, 2013).

In conclusion, genotoxicity can currently not be excluded as contributing to MoA for lung cancer in mice from cumene exposure, but the relevance within this process is currently unknown.

### b) Increased *K-ras* mutations or *p53* mutations and their relevance to humans

The observed increase of *K-ras* mutations in tumours from cumene-exposed mice may or may not be due to a genotoxic event, but this increased incidence is possibly involved in the MoA of cumene carcinogenesis in mice.

An analysis compared the gene expression patterns in cumene-induced tumours with *K-ras* mutations with those in spontaneous occurring tumours without *K-ras* mutations (Wakamatsu *et al.*, 2008). The former were associated with increased expression of genes

- involved in the mitogen activated protein kinase (MAPK) signalling pathway,
- linked to invasion and metastasis,
- linked to inhibition of apoptosis,
- linked to increased angiogenesis,
- linked to increased metastatic potential.

According to the authors of this analysis, the difference in gene expression suggests that cumene-induced carcinomas with *K-ras* mutations have a higher degree of malignancy than tumours without *K-ras* mutations.

There is no indication that these *K-ras* mutations are a species-specific factor in tumorigenesis in mice. In contrast, *K-ras* and p53 mutations have also been found in human lung cancer (Hoenerhoff *et al.*, 2009). Activation of the *K-ras* proto-oncogene and inactivation of the p53 tumour suppressor gene were also frequently observed in human pulmonary adenocarcinoma (NTP, 2013). From that, NTP concludes that “many of the genes with altered expression in the mouse tumor model represent major genes that may play a role in lung and other cancers in humans”.

### c) Lung tumours in mice from exposure to alkylbenzenes

A publication by Cruzan *et al.* (2009) compared tumour incidences in rodents for several alkylbenzenes and other aromatic compounds (such as styrene, ethylbenzene, cumene,  $\alpha$ -methylstyrene, coumarin, naphthalene) and found an obvious discrepancy in tumour incidence in the different rodent species: generally, the incidence of bronchiolo-alveolar adenomas or carcinomas in lungs of mice was significantly increased for most of those substances, whereas no such lung tumours were observed in rats. With focus on more specific studies on styrene, the authors hypothesised that the observed carcinogenicity is linked to a hydroxylation of the aromatic ring (not the side-chain epoxide), for which a specific CYP enzyme (CYP 2F2) is responsible, leading to a reactive metabolite. Cytotoxicity mediated by reactive metabolites formed from CYP2F2 metabolism precedes hyperplasia and finally (at a late stage) tumours. CYP2F2 is expressed in the Clara cells (club-cells) of mice, and is expressed to a much lesser extent in rats and humans. In addition, Clara cells in the lower respiratory tract of mice differ significantly in quantity, function and distribution from humans. Based on those observations, a workshop by “Toxicology Excellence for Risk Assessment” (TERA) was organised in 2013 to discuss the relevance of the respective mouse lung tumours to humans (TERA, 2013). The workshop participants largely confirmed the view of Cruzan *et al.* and concluded that this issue was similar in relevance as the one recently discussed for the peroxisome proliferator-activated receptor alpha (PPAR $\alpha$ ) - MOA and the interspecies extrapolation of the respective mouse liver cancer. PPAR $\alpha$  is – in general – not regarded as quantitatively relevant for humans. The workshop participants stated:

*“Therefore, while this mode of action is theoretically possible in humans if sufficient concentrations of active metabolites are produced, this is highly unlikely to occur given the cross-compound evidence of the central role of mouse-specific CYP2F2 in mediating cytotoxicity. Thus, the hypothesized MOA developed from this cross-compound analysis suggests these chemicals are not expected to cause lung tumors in humans”*(TERA, 2013).

Based on the observations by Cruzan *et al.* (2009) mainly on styrene and further evidence provided by later studies (Cruzan *et al.*, 2012), some recent assessors assume such an analogy of cumene to styrene (e.g., AGS, 2014; DFG, 2016; SCOEL, 2015) and conclude, e.g.: *“For the induction of observed lung ... tumours species-specific mechanisms appear to be decisive”* (SCOEL, 2015).

However, the panel of the TERA workshop also proposed that the following evaluations and criteria would be necessary to demonstrate this MOA (as validated for styrene) for other compounds:

- Evaluate the ring oxidation potential of the chemical’s structure, looking for demonstration of ring-oxidized metabolites, including *in vitro* CYP2F2 metabolism studies
- Look at the genetic activity profiles (GAPs), to determine if mutation is an early and influential key event in the mode of action
- Look for evidence of acute cytotoxicity in mice and rats (*in vivo*)
- If the cytotoxicity response is specific to mice (and not rats), then use CYP2F2 knockout mouse to demonstrate that the response is dependent upon CYP2F2 metabolism
- Lastly, test in the humanized TG mouse to confirm humans will not metabolize sufficient compound via CYP2F1 to produce lung tumors in a “susceptible” system (TERA, 2013).

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Apparently none of those criteria fits to the current toxicological state of knowledge on cumene: neither the respective validations have been performed (TG mouse test; CYP2F2 knockout mouse test; GAP analysis, *in vitro* CYP2F2 studies) nor the data generated for cumene support the suggested MOA. *Inter alia*, the postulated significant participation of CYP2F2 in cumene metabolism in the mice lung has not been demonstrated.

Therefore, current conclusions that the mouse model would not be appropriate for cumene from analogy to other alkylbenzenes are, at least, premature. In fact, several indications suggest a MoA for cumene different from styrene:

- Metabolites for cumene from ring-hydroxylation were found only in small quantities in <sup>14</sup>C analysis by Chen *et al.* (2011) (see metabolites [M2], [M3] in Table 9) and there is no obvious quantitative difference indicating that mice were more prone to this metabolic pathway compared to the rat.
- Cruzan *et al.* (2009) postulated that the CYP2F2 pathway leads to cytotoxicity as an essential step for subsequent hyperplasia and tumours. However, in the long-term NTP studies on cumene there was no observed cytotoxicity in the lower respiratory tract in mice preceding hyperplasia (NTP, 2009; 2013).
- There has been Clara cell loss in bronchioles with styrene exposure. However, this loss has not been observed for (ethylbenzene and) cumene (US EPA, 2014).
- The observed *K-ras* mutations in tumours from cumene exposure may be part of an alternative MoA (see above), which has not been observed or discussed for styrene.

Due to these findings, the postulated analogy to styrene is questionable. In an even more recent workshop by U.S. EPA on the relevance of mice tumours for humans, the similarity of tumours from exposure to alkylbenzenes in mice was further discussed and it was concluded: “*Although structurally related chemicals may cause lung tumors in the B6C3F1 mouse, the mechanism may not be similar*” (Pandiri, 2015; US EPA, 2014).

In conclusion, there are indications from other alkylbenzenes that lung tumours observed in the mouse model would not be relevant for humans because of a (largely) species specific MoA. However, there is insufficient evidence that this MoA seen with other alkylbenzenes is applicable to cumene. Therefore, significant concerns remain that the observed adenoma and adenocarcinoma in the lung in B6C3F<sub>1</sub> mice from exposure to cumene may in fact be meaningful for humans.

### 10.7.1.2 Liver tumours in female B6C3F<sub>1</sub> mice

In the study by NTP (2009) liver tumours were statistically increased in female mice, as evidenced by hepatocellular adenoma or carcinoma in exposed animals (25/50, 26/50, 29/50, 36/50\* (\* p<0.05; P for trend =0.024), further explanation on the data, see Table 13). Even if some of the background dose response data are not increasing with dose (e.g., hepatocellular carcinoma only (10/50; 7/50; 6/50; 12/50)), this does not invalidate this overall clear evidence for an exposure related causal effect in female mice. The question of human relevance is separately discussed below.

#### a) Genotoxicity as MoA

As discussed in Section 10.6.3, a direct interaction of cumene or metabolites with DNA (primary genotoxicity) is probably not a critical MoA for liver tumours of B6C3F<sub>1</sub> mice. Also the Comet assay did not show significant DNA damage in the liver of mice, but only in rat (NTP, 2012). There is only limited evidence that the metabolite  $\alpha$ -methylstyrene (AMS; not marked as one of the [M1]-[M16]-metabolites by Chen *et al.* (2011)), may be genotoxic (Norppa und Vainio, 1983; NTP, 2007; Rim *et al.*, 2012). AMS oxide, a postulated metabolite from cumene and AMS, is genotoxic *in vitro* (see Section 10.6.1 for details). With small amounts of AMS detected in exhaled air from cumene exposure (see Section 9.1 for details), a (primary or secondary) genotoxic MoA for hepatocellular adenoma and carcinoma of cumene in mice cannot be fully excluded; however, indications of a genotoxic MoA are limited.

**b) Relevance of liver tumours in B6C3F<sub>1</sub> mice for humans**

Hepatocellular adenoma and carcinoma are frequently observed in B6C3F<sub>1</sub> mice and were observed in female mice after long term inhalation exposure to cumene. This strain of mice is associated with a high background incidence of liver tumours (NTP, 2009, Table 24). With low evidence of a genotoxic MoA, the relevance of increased liver tumours in B6C3F<sub>1</sub> mice for human exposure has been questioned. Felter *et al.* (2018) reported the results of a workshop on “human relevance of rodent liver tumors”. Workshop discussions focused on two nuclear receptor-mediated MoAs (“Constitutive Androstane Receptor” (CAR) and “Peroxisome Proliferator Activated Receptor-alpha” (PPAR $\alpha$ )) and on cytotoxicity. Most, but not all, participants considered the CAR and the PPAR $\alpha$  MoAs as not relevant to humans based on quantitative and qualitative differences. In contrast, cytotoxicity was considered as clearly relevant to humans, but associated with a threshold MoA.

There are no data to link cumene to either a CAR- or PPAR $\alpha$ -like MoA, with a general deficit to analyse CYP-specifications and critical metabolites for cumene in the species of interest. However, many heterogeneous substances activate the CAR and lead to tumour promotion in mice, as does the model substance phenobarbital (Elcombe *et al.*, 2014). Examples are pyrene (Zhang *et al.*, 2015), ethyl isobutyl ketone (Hughes *et al.*, 2016), or tetrahydrofuran (Choi *et al.*, 2017). Sweeney *et al.* (2015) assume this MoA also for ethylbenzene, which is similar to cumene in structure, but details are not available.

There are no obvious indications of cytotoxicity preceding neoplastic effects in the chronic inhalation study with cumene (NTP, 2009). In the 14-weeks studies there was some minimal chronic focal liver inflammation in female mice exposed to the lowest concentration (62.5 ppm) of the test regimen, but without a relation between dose and response, and some increase in relative liver weights at elevated exposures (125 ppm) (DFG, 2016). However, DFG (2016) suggests “chronic organ damage” as a non-genotoxic MoA of cumene: “*In analogy to ethylbenzene, the isopropyl benzene-induced neoplasms in the liver of female mice and the eosinophilic foci of male animals could therefore be the result of increased cell proliferation following chronic organ damage*”. If this consideration was confirmed by better data, it would support categorising cumene as a threshold carcinogen.

In summary, relevance for humans of the observed liver tumours in mice from cumene exposure may be low or not existing. However, the MoA is still largely unknown and has been insufficiently examined. Therefore, these data support the conclusion that cumene’s induction of liver tumours in female mice is uncertain with respect to the relevance for humans.

**10.7.1.3 Renal tumours in F344/N male rats**

In the study by NTP (2009) a suggestive increase of renal tubular cell adenoma or carcinoma (combined) in male rats due to exposure to cumene was observed.

- (i) The incidence of renal tubular hyperplasia was increased significantly in all exposed groups (hyperplasia is possibly linked to the MoA of cancer),
- (ii) The incidence of renal tubular adenoma was (insignificantly) higher than control in all exposure groups,
- (iii) Renal tubular carcinoma were (insignificantly) increased above control in the two higher exposure groups,
- (iv) Renal tubular cell adenoma or carcinoma (combined) were elevated in all exposure groups, close to significance (but not significant for trend (P=0,087)), and significantly increased in the mid exposure group (8/50\* vs. 2/50 in control),
- (v) For one of the metabolites of cumene (alpha-methylstyrene) there was also some evidence of carcinogenic activity in male F344/N rats based on increased incidences of renal tubule adenomas and carcinomas (combined) (NTP, 2007).

The question of human relevance is separately discussed below.

**a) Genotoxicity as MoA**

As discussed in Section 10.6.3, a direct interaction of cumene or metabolites with DNA (primary genotoxicity) is not a plausible MoA for renal tumours. The results of a Comet assay did not show a significant increase in DNA damage in the kidneys of male rats *in vivo* after exposure to cumene (NTP, 2012). There is only limited evidence that the metabolite  $\alpha$ -methylstyrene (AMS; not marked as one of the [M1]-[M16]-metabolites by Chen *et al.* (2011)) may be genotoxic (Norppa und Vainio, 1983; NTP, 2007; Rim *et al.*, 2012). AMS oxide, a postulated metabolite of cumene and AMS, is genotoxic *in vitro* (see Section 10.6.1 for details). With small amounts of AMS detected in exhaled air after cumene exposure (see Section 9.1 for details), a (primary or secondary) genotoxic MoA for renal tumours of cumene in the rat is unlikely, however cannot be fully excluded.

## b) Relevance of renal tumours in male rats for humans

Cumene leads to renal tumours in male rats after inhalation exposure. One of the metabolites, confirmed in rodents, also is associated with renal tubular adenoma and carcinoma (combined) in male but not in female rats (NTP, 2007). However,  $\alpha$ -methylstyrene apparently is only a minor metabolite of cumene (see Section 9.1).

A major MoA of renal tumours in male rats is  $\alpha_{2u}$ -globulin accumulation, observed as hyaline droplets, in proximal tubule. This may lead to epithelial degeneration and necrosis, granular casts, cell proliferation, chronic progressive nephropathy (more often in older rats), atypical hyperplasia within the proximal tubules, and progression to tumours (Capen *et al.*, 1999; Swenberg und Lehman-McKeeman, 1999).

The International Agency for Research on Cancer (IARC) developed a list of criteria, which must be met, for identifying agents where this is the sole MoA of renal tumours. Those criteria are:

- Lack of genotoxic activity,
- Male rat specificity for nephropathy and renal tumorigenicity,
- Indication of the characteristic sequence of histopathological changes, of which protein droplet accumulation is obligatory,
- Identification of the protein accumulating in the tubule cells as  $\alpha_{2u}$ - globulin,
- Reversible binding of the chemical or metabolite to  $\alpha_{2u}$ - globulin,
- Induction of sustained increased cell proliferation in the renal cortex,
- Similarities in dose-response relationship of the tumour outcome with the histopathological endpoints (protein droplets,  $\alpha_{2u}$ - globulin accumulation, cell proliferation) (Capen *et al.*, 1999).

The NTP (2009) concluded that the lesions observed in male rats were characteristic of  $\alpha_{2u}$ -globulin accumulation. However, not all of the IARC-criteria were met: male specificity of nephropathy (some nephrotoxicity was also observed in females), and evidence of sustained cell proliferation were not provided; reversible binding to  $\alpha_{2u}$ - globulin was not assessed and genotoxicity as a possible MoA is not completely ruled out. IARC (2013) points out the “*one of the mutagenic metabolites of cumene,  $\alpha$ -methylstyrene oxide, could play a role in the initiation of such tumours*” and concludes “*the data do not support a mechanism that involves  $\alpha_{2u}$ -globulin- associated nephropathy in the development of these kidney tumours.*” Note that IARC regards  $\alpha$ -methylstyrene oxide as a confirmed metabolite of cumene, which is, however, only a proposed metabolite according to Chen *et al.* (2011). Therefore, at least the quantitative relevance of this metabolic pathway is uncertain. NTP (2016) concludes: “*Overall, the data provide evidence that cumene causes kidney tumors largely via  $\alpha_{2u}$ -globulin nephropathy; however, it cannot be ruled out that other mechanisms, such as genotoxicity, also contribute to kidney tumor formation. Although it is likely that genotoxicity plays a role in cumene-induced carcinogenicity at some tissue sites, the strongest evidence for genotoxicity was found for lung and liver tumors, and the extent to which genotoxicity contributes to the formation of kidney tumors is unknown. Thus, the relevance of the kidney tumors in male rats to human cancer is uncertain.*” (NTP, 2016).

Based on relevant indications of a species specific effect, we agree with this NTP conclusion (“human relevance is uncertain”).

#### 10.7.1.4 Nasal tumours in male F344/N rats

In the experimental study by NTP (2009) nasal tumours were statistically increased in male rats, as evidenced by adenoma of the respiratory epithelium (including multiple and all sites (0/50, 7/50\*\*, 18/49\*\*\*, 10/50\*\*\* (\*\*\*)  $p \leq 0.001$ ; P for trend:  $P < 0.001$ )) and in females ((4/50, 31/50\*\*\*, 42/50\*\*\*, 46/50\*\*\*, P for trend:  $P = 0.004$ ), for further explanation of the data, see Table 13). Even if response data are not monotonously increasing with dose, this does not invalidate this overall clear evidence for an exposure related causal effect in male rats. The relevance of these findings is further supported by an (insignificant) increase of respiratory epithelium adenoma in female rats (0/50; 5/48\*; 4/50; 3/50; P at low dose exposure  $< 0.05$ ; for details see Table 13). A potential progression to malignant tumours is separately discussed below.

Increases in the incidence of benign nasal tumours (adenoma of the respiratory epithelium) were observed in rats of both sexes (NTP, 2009). NTP (2016) assumes that this kind of tumours cannot progress to malignancy. They cited a publication by Brown (1991) as evidence. However, from analysis of the original study by Brown (1991), no such definite statement was found. In NTP (2009) a different conclusion was reported: "Progression of nasal respiratory epithelial adenomas to malignancy has been described in the literature".

DFG (2016) believes that the CYP enzymes probably responsible for transforming cumene to reactive metabolites in the nasal cavity are much less expressed in humans compared to other mammalian species. Therefore no relevance for human cancer risk is assumed, "*however, according to current data [the relevance of nasal tumours for humans] cannot be excluded*". AGS (2014) uses the dose-response data for nasal tumours in rats to calculate cancer risks, to support the occupational exposure limit (OEL) derived from non-cancer effects quantitatively and derived a very low associated excess risk at air concentrations of 10 ppm (occupational exposure scenario) implicitly acknowledging a possible relevance of this cancer endpoint for human exposure.

Overall, the observed effect in male rats is regarded relevant for human exposure, but progression to malignancy is uncertain. A significant contribution of cytotoxicity (secondary to cytotoxicity) to hyperplasia and subsequent occurrence of tumours (i.e., threshold-type mode of action) is possible.

#### 10.7.1.5 Other tumour sites

In male mice, haemangiosarcoma of the spleen and follicular-cell adenoma in the thyroid gland may have been treatment related based on marginal increases over historical control values. However, haemangiosarcoma occur in multiple tissue types and are not specific to or rare in the spleen (NTP, 2013). In addition, the incidence in all organs was within the historical control ranges for inhalation studies and for all routes. Follicular-cell adenoma were only insignificantly increased at the highest exposure group; only the trend was significant ( $P = 0.01$ ). The unadjusted overall tumour rate for thyroid adenoma (6%) at the high exposure concentration was within the historical control range (0%–6%) for inhalation studies and for all routes.

Further, the incidence of interstitial-cell adenoma of the testes of rats was significantly increased at the highest exposure level with a positive trend and exceeded the historical control from inhalation studies. However, this type of adenoma does not progress to malignancy (NTP, 2013).

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**Table 16: Compilation of factors to be taken into consideration in the hazard assessment**

Species and strain	Tumour type and background incidence	Multi-site responses	Progression of lesions to malignancy	Reduced tumour latency	Responses in single or both sexes	Confounding effect by excessive toxicity?	Route of exposure	MoA and relevance to humans
mice, B6C3F <sub>1</sub>	a) lung alveolar/bronchiolar adenoma or carcinoma (background incidence: 19/50, males; 4/50 females)	yes, see Table 13	yes, see Table 13	Yes (first incidence high dose group; males: 420 days, compared to 628 days in control; females: 513 days, compared to 533 days)	both sexes	not assumed to be due to excessive toxicity	inhalation	possibly relevant; discussed in detail in Section 10.7.1.1
	b) liver high background incidence of adenoma or carcinoma in this strain (40/50 males, 25/50 females )		yes, see Table 13	Yes (males), no (females) (first incidence high dose group; males: 391 days, compared to 551 days in control; females: 662 days, compared to 607 days)	significant only in females			relevance uncertain; discussed in detail in Section 10.7.1.2
	c) hemangiosarcoma in males (background incidence: 0/50; incidence of treated animal within historical control ranges)		tumour is malignant	No tumours in control group	only observed in male mice			relevance uncertain; see Section 10.7.1.5
	d) thyroid gland follicular cell adenoma in males (background incidence: 0/50 (only hyperplasia in control); unadjusted overall tumour rate within historical control)		only (non-malignant) adenoma observed	No tumours in control group	only observed in male mice			inadequate evidence; see Section 10.7.1.5
rat, F344/N	a) nose adenoma respiratory epithelium (no elevated background incidence: 0/50 males, 0/50 females)	yes, see Table 13	no, see Table 13, but cannot be excluded (see Section 10.7.1.4)	No tumours in control group	both sexes (but not clearly significant in females)	not assumed to be due to excessive toxicity	inhalation	uncertain, but low evidence; discussed in detail in Section 10.7.1.4



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Species and strain	Tumour type and background incidence	Multi-site responses	Progression of lesions to malignancy	Reduced tumour latency	Responses in single or both sexes	Confounding effect by excessive toxicity?	Route of exposure	MoA and relevance to humans
	b) kidney (renal tubule adenoma or carcinoma in control males only: 2/50)		yes, see Table 13	Yes (first incidence high dose group; males: 618 days, compared to 729 days in control)	males only			uncertain, but low evidence; discussed in detail in Section 10.7.1.3
	c) testes (adenoma; high background incidence, males: 36/50)		only (non-malignant) adenoma observed, does not progress to malignancy	Inconclusive (first incidence high dose group; males: 541 days, compared to 558 days in control)	males only			inadequate evidence; discussed in detail in Section 10.7.1.5

### 10.7.2 Comparison with the CLP criteria

IARC (2013) classified cumene “possibly carcinogenic to humans (Group 2B)”, based on “sufficient evidence in animals” and “no data available in humans”, according to the IARC criteria.

For potential classification on carcinogenicity, criteria of the CLP Regulation (EC, 2017) were applied.

- *Category 1A, known to have carcinogenic potential for humans, classification is largely based on human evidence (EC, 2017)*

As indicated, there are no relevant data on exposure to cumene for classification into Category 1A.

- *Evidence for Category 1B is derived from animal experiments for which there is sufficient evidence to demonstrate animal carcinogenicity (presumed human carcinogen) (EC, 2017)*

Based on the results provided by NTP (2009) with studies in mice and rats, overall there is sufficient evidence in animals for carcinogenicity. This conclusion is in agreement with IARC (2013). However, the criterion above is closely linked to the relevance for humans (see remark in brackets: “presumed human carcinogen”), as further discussed by EC:

- *Substances which have induced benign and malignant tumours in well performed experimental studies on animals are considered also to be presumed or suspected human carcinogens unless there is strong evidence that the mechanism of tumour formation is not relevant for humans. (EC, 2017)*

As evidenced in detail in Section 10.7.1, the relevance for humans of observed tumours in animal studies has been seriously questioned in the case of cumene. In further specifications, EC requests:

- *Additional considerations (as part of the weight of evidence approach) ...a number of other factors need to be considered that influence the overall likelihood that the substance poses a carcinogenic hazard to humans....Some important factors which may be taken into consideration, when assessing the overall level of concern are...mode of action and its relevance for humans, such as cytotoxicity with growth stimulation, mitogenesis, immunosuppression, mutagenicity. (EC, 2017)*

These considerations are described in detail in Section 10.7.1 for each tumour site and animal species discussed as potentially relevant for carcinogenicity classification. It is concluded that there are serious doubts that the respective modes of action for carcinogenic effects in experimental animals are relevant for humans in case of cumene, but that relevant concerns remain.

- *Category 2: suspected human carcinogen. The placing of a substance in Category 2 is done on the basis of evidence obtained from human and/or animal studies, but which is not sufficiently convincing to place the substance in Category 1A or 1B, based on strength of evidence together with additional considerations.” (EC, 2017)*

When assessing the overall level of concern for classification of carcinogenicity for cumene, according to criteria in Section 3.6.2.2.6 (EC, 2017), we conclude that

(a) tumour type and background incidence;

Observed tumor types (lung alveolar/bronchiolar adenoma or carcinoma as evidenced from mice; hepatocellular adenoma or carcinoma as evidenced from female mice; renal tubular adenoma or carcinoma found (insignificantly) increased in male rats, respiratory adenoma as evidenced from male rats) are generally assumed to be relevant for classification. However, for some of those observed tumours in rodent studies the mode of action may possibly not be relevant for humans (see criterion k, below) or progression to malignancy has been questioned (see criterion c, below). Background incidence for hepatocellular tumours in B6C3F<sub>1</sub>-mice is high, adding to uncertainties on the relevance for classification based on respective observations in this mouse strain, which is very sensitive to respective hepatocellular tumours. For hepatocellular tumours in B6C3F<sub>1</sub>-mice quantitative species extrapolation is not possible, if the mode of action is not clearly genotoxic. The qualitative relevance is uncertain.

(b) multi-site responses;

Based on overall evidence, cumene is regarded to show multi-site tumour responses in the rodent studies (lung in mice, liver in female mice, renal tumours in male rats, nasal epithelium in rats) and, furthermore, but

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with less evidence: testes in rats, thyroid gland and hemangiosarcoma in spleen of mice. However, this does not imply that those tumours are all induced by one overall relevant mode of action. Furthermore, it does not imply that all of those tumours in rodents are relevant to humans.

(c) progression of lesions to malignancy;

Some of the observed tumours in rodent studies may potentially progress to malignancy as assumed by default (lung tumours in mice, hepatocellular tumours in mice, kidney tumours in rats, follicular cell adenoma in the thyroid gland), for other tumours this progression to malignancy is uncertain (this type of nasal adenoma in rats) or not expected (interstitial cell adenoma in testes of rats).

(d) reduced tumour latency;

This criterion was not addressed and is not regarded influencing classification for the tumours observed in the rodent experimental studies on cumene.

(e) whether responses are in single or both sexes;

Observed tumours were not significantly increased in both sexes for all tumour sites apart from the lung, which was affected similarly and significantly in male and female mice. Specifically, hepatocellular tumours have only been increased significantly in female mice (and only insignificantly in male mice); kidney tumours have only insignificantly been increased in male rats, with only insignificant increase of nonmalignant nephropathic effects in female rats; and nasal adenoma were significantly increased only in male rats with significant (but not neoplastic) hyperplasia respiratory epithelium and only insignificant increase of adenoma in female rats. From these observations, cumene is not regarded to be tumourigenic to only one sex. For some tumour locations a sex- and species-specific mode of action is discussed (e.g., kidney tumours in male rats), however, without firm conclusions.

(f) whether responses are in a single species or several species;

Tumourigenic responses have been observed in more than one species (i.e., rats and mice). However, the tumour sites usually differed between the two species and some of the observed tumours may be species-specific.

(g) structural similarity to a substance(s) for which there is good evidence of carcinogenicity;

Some other aromatic hydrocarbons are associated with similar tumours, but the evidence for their carcinogenicity does not help to eliminate uncertainties on the mode of action and human relevance of the tumours observed in the NTP studies on cumene. Specifically, for relevance of tumours in mice to humans comparisons with other aromatic hydrocarbons were discussed intensively (TERA, 2013; US EPA, 2014), as also addressed in Section 10.7.1.1 of this report.

(h) routes of exposure;

This criterion was not addressed and is not regarded influencing classification for the tumours observed in the rodent experimental studies on cumene. Only inhalation studies on carcinogenicity were available. However, *in vivo* genotoxicity studies also included data from oral (gavage) exposure.

(i) comparison of absorption, distribution, metabolism and excretion between test animals and humans;

Even though absorption, distribution, metabolism and excretion appear to be similar between animal species and humans, some differences in quantitative metabolism have been reported (see Section 9, this report). Specifically, higher tissue concentrations in rat kidney and mouse lung studies correlate with higher incidence of tumours in these studies. There are relevant uncertainties on the metabolism of cumene in the mouse lung and whether this metabolism is species specific.

(j) the possibility of a confounding effect of excessive toxicity at test doses;

There is no reason to assume relevant influences of confounders in the outcome of the critical studies on cumene.

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(k) mode of action and its relevance for humans, such as cytotoxicity with growth stimulation, mitogenesis, immunosuppression, mutagenicity.

These aspects are discussed in detail in Section 10.7.1, of this report.

From the definition of Category 2 in Regulation 1272/2008 (EC, 2017), and from the overall summary discussion based on Regulation criteria a-k, Section 3.6.2.2.6 (above) , and from the discussion described in detail in Section 10.7.1, for each tumour site and animal species for cumene, the substance is regarded a “suspected human carcinogen”. The relevance of the observed tumours in experimental animals is uncertain (less than sufficient evidence), which would be needed for classification in Category 1B.

### 10.7.3 Conclusion on classification and labelling for carcinogenicity

Cumene is a multi-site carcinogen in rodent inhalation studies. No data exist for other exposure pathways. There are, however, several indications of limited or no relevance of the observed tumours for human exposure. Various MoAs have to be considered and a definite conclusion on a species-specific MoA, not relevant for human exposure, is not possible. Specifically,

- for lung tumours in mice, transformation of cumene in the lower respiratory tract by CYP2F enzymes with ring oxidation in mice Clara cells would be a possible species specific mode of action, but some observations (like the increase incidence of K-ras mutations in tumours of exposed mice and the lack of cytotoxic effects in the animals) point to alternative MoAs or are not in compliance with the postulated mode of action. Moreover, involvement of specific enzymes in the metabolism of cumene in the lungs have not been shown and no knock out-model experiments are available to validate the hypothesised species specific MoA;
- for liver tumours in mice, there are indications for species specific MoA via nuclear receptors (CAR, PPAR $\alpha$ ), but no evidence is available for the involvement of these receptors. Furthermore, a cytotoxic MoA is also discussed. This would also be relevant for human exposure, but would implicate an effect threshold for tumorigenicity. However, as no clear cytotoxic effects were observed in mice, this MoA also cannot be confirmed;
- for renal tumours in male rats, a MoA has been suggested which involves  $\alpha_2\text{u}$ -globulin and hyaline droplets accumulation to finally result in species and sex specific tumours. Even though this MoA is supported by the observed effects, full compliance with the IARC criteria for this kind of mechanism is not provided. Thus, human relevance cannot be excluded;
- also, for further tumour sites in experimental animals (i.e., hemangiosarcoma of the spleen, follicular cell adenoma in the thyroid gland and testis adenoma) there also remain some uncertainties with regard to causality by cumene exposure and/or relevance to humans;
- for nasal tumours in rats, relevance to humans can be assumed, however progress to malignancy has been questioned and is regarded uncertain;
- the majority of *in vitro* and *in vivo* studies on genotoxicity do not indicate a genotoxic MoA for any of the cancer sites. However, a) increase of specific K-*ras* and p53-mutations in mouse lung tumours induced by cumene exposure, b) minor DNA damage, as indicated by Comet assays *in vivo*, c) potential genotoxicity of the presumed metabolite  $\alpha$ -methylstyrene oxide, as evidenced from *in vitro* testing, and d) some further reactive metabolites formed by ring-hydroxylation of cumene (i.e, quinone methide or catechol via isopropylphenol formation) may be involved in the mode of action. The relevance of primary genotoxicity mediated by  $\alpha$ -methylstyrene oxide may be questioned, because of the low concentrations of this presumed metabolite. However, available data are insufficient for further conclusions. Some of the observed genetic alterations could result from secondary genotoxicity or from epigenetic mechanisms.

Therefore, classification to

**Carc. 2, ‘H351: Suspected of causing cancer’,**

is warranted.

## RAC evaluation of carcinogenicity

### Summary of the Dossier Submitter's proposal

For the assessment of carcinogenicity, the DS included two high-quality carcinogenicity studies in mice and rats via the inhalation route (NTP, 2009). In addition, two transformation assays in mice (Putman, 1987b; Gulf Oil corporation, 1984) and a mechanistic study in mice lung tumours (Wakamatsu *et al.*, 2008) were available.

Based on the available data the DS concluded that a classification of cumene in Category 2, H351 is warranted. Based on the results of the NTP, 2009 study in mice and rats, there is sufficient evidence in animals to demonstrate carcinogenicity.

The DS concluded that the evidence on MoA for the tumours was insufficient to completely dismiss their relevance to humans. The following tumours were taken into account by the DS:

- Kidney tubular cell tumours in male rats. The DS considered it plausible that the  $\alpha_2$  - globulin nephropathy MoA, specific to male rats, could be the underlying cause of the observed kidney tumours. Nevertheless, the fact that progressive chronic nephropathy was also seen in female rats and that there is no specific cumene data on the MoA, leads to uncertainties on the proposed MoA.
- Nasal respiratory epithelium adenoma in male rats. These tumours are assumed to be relevant to human but progression to malignancy was considered uncertain.
- Lung alveolar/bronchiolar adenoma or carcinoma in male and female mice. Transformation of cumene by CYP2F2 in the lower respiratory tract was considered as a possible species specific MoA. Nevertheless, some specific data on cumene suggest that other MoA may be involved.
- Liver adenoma or carcinoma in male mice. There is some indication of a specific MoA *via* nuclear receptors (CAR, PPAR $\alpha$ ). Nevertheless, no evidence is available on the involvement of these receptors.

### Comments received during consultation

In contrast to the DS, two MSs were in favour of a more stringent classification as Carc 1B. Their conclusion was based on the observation of tumours in both sexes (lung tumours in mice) and two species (tumours in the lung and liver in mice, tumours in the nose and kidney in rat). Although these MSs acknowledged that the kidney tumours in male rat and the liver tumours in female mice could be associated with a MoA non-relevant to humans, they considered the evidence provided on the different modes of action was not sufficient to exclude human relevance. One of the MS placed special emphasis on the significantly increased lung alveolar/bronchiolar adenomas/carcinomas in male and female mice. This MS also pointed out that the mutations observed in these tumours, in contrast to spontaneous tumours, were also relevant in human lung cancer and considered these mutations as evidence for genotoxicity.

One MS was unsure whether the data supported a classification as Cat 1B or 2. This MS recommended a tabular comparison of the arguments in favour and against each proposed

MoA and the characteristics of the different tumours observed (significance, malignancy, multi-site, dose-response,...) and the discussed modes of actions. This MS also pointed out that the CLP regulation assumes human relevance of findings in animals "unless there is strong evidence that the mechanism of tumour formation is not relevant for humans" (CLP, Annex I: 3.6.1.1).

The DS prepared the tabular comparison as suggested by the MS, including the results of the pilot study on CAR/PXR MoA provided by industry during PC. This tabular overview can be found attached to this Background document (see page 90).

Three industry representatives and one individual were not in favour of classification.

The individual argued that overall the tumour responses were weak and that the data presented on the different modes of action would either disprove human relevance (considered the evidence presented for CAR MoA, CYP2F2/Clara cell MoA and  $\alpha$ 2u-globulin MoA as sufficient to rule out human relevance) or that progress to malignancy was not expected (nasal tumours in male and female rats). As for the remaining tumours observed in mice and rat the commenter considered them to fall within the background incidence.

Furthermore, the other industry commenters considered the evidence insufficient to support a classification as carcinogen. With regard to the NTP (2009) studies in rat and mice they pointed out that the CLH report would not address saturation of certain metabolic pathways for cumene resulting in the formation of critical metabolites, however, without providing evidence for this shift towards critical metabolic pathways at these elevated doses.

They further criticised that despite no classification for germ cell mutagenicity is proposed, a lengthy discussion of the genotoxic potential of cumene is included in the dossier. The DS clarified that even though they proposed no classification for germ cell mutagenicity there are remaining uncertainties regarding the genotoxic potential of cumene in somatic cells.

In a similar manner to the individual commenter they were of the view that the provided evidence for the MoA with non-human relevance for lung, liver and kidney tumours was sufficient.

For the remaining tumours they concluded that those were within the spontaneous background ranges and/or did not meet the statistical threshold relevant to common tumours.

Overall Industry was of the view that only limited criteria for the classification as carcinogen were met.

### **Assessment and comparison with the classification criteria**

Two carcinogenicity assays were included in the CLH report, one in B6C3F1 mice and one in F344 rats (NTP, 2009). Additionally, mechanistic studies were available in the dossier.

#### **Mouse**

Mice were exposed to cumene vapour concentrations of 0, 125 (female mice only), 250, 500, or 1000 (male mice only) ppm. An exposure concentration-related decrease in survival occurred in male mice, and the survival of 1000 ppm males was significantly less than that of the chamber controls. Mean body weights for the 1000 ppm males were generally less than those of the chamber controls after week 8 of the study, and those of the 500 ppm females were less from week 28 until week 76 of the study (NTP, 2009), but the decrease in body weight did not exceed 10% at any time point. Dose selection was based on the results of a 3-month study in which 8/10 females died at 1000 ppm. The observed mortality occurred in the first week of dosing and was considered an acute effect. Liver weights of mice exposed to

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500 or 1000 ppm were significantly increased and mean body weights of males exposed to 500 or 1000 ppm were significantly less than those of the chamber controls (NTP, 2009). Thinness and some difficulties in breathing was seen in some of the top dose males and females, but no other clinical signs were observed.

Lung tumours

Lung tumours were statistically significantly increased in mice as evidenced by alveolar/bronchiolar adenoma or carcinoma in males (19/50, 38/50\*\*\*, 42/50\*\*\*, 43/50\*\*\* at 0, 250, 500 and 1000 ppm, respectively (\*\*\*)  $p \leq 0.001$ ; P for trend:  $P < 0.001$ ) and in females (4/50, 31/50\*\*\*, 42/50\*\*\*, 46/50\*\*\* at 0, 125, 250 and 500 ppm, respectively P for trend:  $P < 0.001$ ), for further information on lung lesions see the table below. Historical incidence for NTP 2-year inhalation studies in male mice were: 32.5% (Range: 26-44%) and in female mice: 7.6% (range 2-14%) (NTP, 2009).

As noted by the DS not all of the related lesions (e.g. bronchiolar metaplasia in males) followed a clear dose-response, but a dose-related increase in severity was seen for these effects. The data clearly indicate an exposure related neoplastic effect in both sexes, with a slightly higher susceptibility in females.

Dose (ppm)	Incidence of lung lesions							
	Males				Females			
	0	250	500	1000	0	125	250	500
Alveolar epithelium, bronchiale, metaplasia	5/50	43/50 **	42/50 **	39/50 **	0/50	42/50 **	49/50 **	47/50 **
Bronchiale, hyperplasia	0/50	11/50 **	17/50 **	18/50 **	0/50	17/50 **	10/50 **	14/50 **
Alveolar/bronchiolar adenoma, multiple	1/50	12/50 **	15/50 **	20/50 **	0/50	13/50 **	20/50 **	30/50 **
Alveolar/bronchiolar adenoma (includes multiple)	13/50	31/50 ***	31/50 ***	29/50 ***	1/50	26/50 ***	36/50 ***	38/50 ***
Alveolar/bronchiolar carcinoma, multiple	0/50	8/50 **	20/50 **	17/50 **	0/50	6/50 *	7/50 **	19/50 **
Alveolar/bronchiolar carcinoma (includes multiple)	9/50	19/50 *	32/50 ***	33/50 ***	3/50	16/50 ***	20/50 ***	34/50 ***
Alveolar/bronchiolar adenoma or carcinoma	19/50 <sup>a</sup>	38/50 <sup>a</sup> ***	42/50 <sup>a</sup> ***	43/50 <sup>a</sup> ***	4/50 <sup>b</sup>	31/50 <sup>b</sup> ***	42/50 <sup>b</sup> ***	46/50 <sup>b</sup> ***

<sup>a</sup> Historical control data: 146/449 (32,5% ± 5,9%), range 26% - 44%

<sup>b</sup> Historical control data: 34/449 (7,6% ± 4,0%), range 2% - 14%

\* ≤ 0.05, \*\* ≤ 0.01, \*\*\* ≤ 0.001 (Poly-3 test)

The DS discussed primary genotoxicity, secondary genotoxicity (formation of ROSs), epigenetic factors and cytotoxicity via Cyp2f2 expression in Clara cells (Club cells) as a possible underlying cause of the observed lung tumours.

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### a) Primary genotoxicity:

The DS considered direct interaction of cumene or metabolites of cumene with DNA (Primary genotoxicity) an unlikely cause of the observed tumours. However, RAC considers the significant increase in DNA damage in lungs of female mice observed in an *in vivo* Comet assay as supportive for a genotoxic MoA. It has to be noted that the Comet assay only gave positive results in female lung tissues, but tumours were seen in both sexes, though female mice were more susceptible than males.

The remaining genotoxicity/mutagenicity tests with cumene were largely negative, but also results for the metabolite AMS were considered by the DS. For this metabolite most of the assays were also negative, but it gave weakly positive results in two SCE assays (Norppa & Vainio, 1983, NTP, 2007). Inconclusive results were observed *in vivo* in the micronucleus assay (positive in female mice and negative in male mice following repeated exposure, negative in male mice following single exposure).

Although not confirmed *in vivo* it is likely that also AMS oxide is formed as a metabolite. For AMS oxide positive results were obtained in a gene mutation test in bacteria.

There was a clear increase in *K-ras* and *p53* mutations in cumene induced tumours compared to spontaneously occurring tumours from concurrent and historical controls. The type of mutations seen in the cumene induced tumours was also different. In addition, an increased loss in heterozygosity was seen in cumene-induced mouse lung tumours, which was not seen in spontaneous tumours in control mice (Hong *et al.*, 2008).

Wakamatsu *et al.* (2008) further investigated gene expression patterns of cumene-induced lung tumours with *K-ras* mutations with those of spontaneously occurring tumours without *K-ras* mutations. The cumene induced tumours with *K-ras* mutations had increased expression for genes involved in the MAPK signalling pathway, inhibition of apoptosis, increased angiogenesis, invasion and metastasis. The authors concluded that these features are indicative of a higher degree of malignancy.

In line with the DS, RAC is of the view that there is no indication for species specificity of the observed *K-ras* mutations seen in mice. This is supported by Hoenerhoff *et al.* (2009) where it is reported that *K-ras*, as well as *p53* mutations, were found in human lung cancer and NTP (2013) where it is pointed out that activation of the proto-oncogen *K-ras* and inactivation of the tumour suppressor gene *p53* were frequently observed in human pulmonary adenocarcinoma.

Overall, it can be concluded that the increased frequency of specific types of mutations observed appear to be characteristic for cumene induced lung tumours. The NTP report suggested that the high frequency of *K-ras* mutation in adenoma was a relatively early event. However, as the sample size was small, it cannot be clearly concluded whether these specific mutations were caused by cumene or its metabolites or if they are a feature of the lung tumours as they developed as a consequence of cumene exposure.

In conclusion, genotoxicity can currently not be excluded as contributing MoA for lung cancer formation.

### b) Secondary genotoxicity via formation of ROS:

Hong *et al.* (2008) mentioned that indirect genotoxicity via ROS formation might be involved in the mutations observed in the lung tumours induced by cumene. Indeed, among others, G to T transversions were noted in the *K-ras* genes. This type of transversion is consistent with 8-OH-G adduct formation (see also section on germ cell mutagenicity). A study to clarify the potential of cumene to induce DNA damage through the formation of ROS was conducted by Kim *et al.* (2008), but RAC considers this study as inadequate to draw a conclusion. In their



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response to comments from the consultation the DS stated that generation of ROS is in general associated with inflammation processes that can be observed in the histopathological examination of the relevant tissues, but in case of the lung of mice, no increased inflammation was observed in exposed mice vs controls.

In conclusion, RAC is of the view that a contribution of ROS to the observed tumours cannot be ruled out.

### c) Epigenetic factors:

Wakamatsu *et al.* (2008) also found indications for the action of possible epigenetic mechanisms in cumene-induced lung cancer. They observed significant alterations in genes associated with the histone deacetylase complex (HDAC) in mouse lung carcinomas. A stronger association was observed between altered genes supposedly associated with HDACs and tumours with *K-ras* mutations compared to tumours without *K-ras* mutations. Therefore, *K-ras* activation may affect histone modification or vice versa.

### d) Involvement of Cyp2f2 expression in Clara cells (Club cells), and human relevance of the observed tumours:

Lung tumours induced by several alkylbenzenes and other aromatic compounds (e.g. styrene) have been observed in mice but not in rats. Cruzan *et al.* (2009, 2012) mainly investigated styrene and postulated that a mouse specific expression of Cyp2f2 in Clara cells (Club cells), which is supposed to catalyse hydroxylation of the aromatic ring (not the side-chain epoxide), leads to the formation of reactive metabolites. It is postulated that these reactive metabolites lead to cytotoxicity, resulting in hyperplasia and finally (at late stage) in tumours. CYP2F2 expression is lower in club cells than the expression of the orthologous enzymes of rat and human, and the frequency of club cells in the lower respiratory tract is much lower in humans compared to rodents. In a workshop by Toxicology Excellence for Risk Assessment, TERA (2013) the relevance of the respective mouse tumours for humans was discussed.

The workshop concluded that the MoA is theoretically possible in humans, if sufficient concentrations of active metabolite were produced, but highly unlikely to occur given the cross-compound evidence of the central role of mouse-specific Cyp2f2 in mediating cytotoxicity. An analogy of cumene to styrene was assumed in recent assessments e.g. AGS (2014); DFG (2016) or SCOEL (2015), but IARC (2013) considered the lung tumours seen with cumene relevant for humans.

The TERA (2013) workshop also developed criteria which need to be fulfilled in order to demonstrate the postulated MoA for other compounds, based on styrene:

- Evaluate the ring oxidation potential of the chemical's structure, looking for demonstration of ring-oxidized metabolites, including *in vitro* Cyp2f2 metabolism studies
- Look at the genetic activity profiles (GAP), to determine if mutation is an early and influential key event in the MoA.
- Look for evidence of acute cytotoxicity in mice and rats (*in vivo*).
- If cytotoxicity response is specific to mice (and not rats), then use Cyp2f2 knockout mouse to demonstrate that the response is dependent upon Cyp2f2 metabolism.
- Lastly, test in the humanised lung tumours in a "susceptible" system (TERA, 2013).

The DS concluded that none of the above tests have been performed for cumene, nor do the available studies support the described MoA.

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Metabolites for cumene from ring-hydroxylation were found only in small quantities in <sup>14</sup>C analysis by Chen *et al.* (2011) and there is no obvious quantitative difference indicating that mice were more prone to this metabolic pathway compared to rat.

Cruzan *et al.* (2009) postulated the Cyp2f2 pathway leads to cytotoxicity as an essential step for subsequent hyperplasia and tumours. However, in the long-term NTP studies on cumene there was no observed cytotoxicity in the lower respiratory tract in mice preceding hyperplasia (NTP, 2009, 2013).

There has been Clara cell loss in bronchioles with styrene exposure. However, this loss has not been observed for cumene (US EPA, 2014).

The observed *K-ras* mutations in tumours from cumene exposure may be part of an alternative MoA, which has not been observed or discussed for styrene.

The postulated analogy of cumene to styrene and other alkylbenzenes is therefore not supported and is in line with a recent US EPA workshop that concluded: "Although structurally related chemicals may cause lung tumours in the B6C3F1 mouse, the mechanism may not be similar" (Pandiri, 2015, US EPA, 2014).

In conclusion, there is insufficient evidence that the MoA observed in other alkylbenzenes is applicable to cumene and some findings (e.g. lack of cytotoxicity or lack of difference with regard to metabolites between rat and mouse) also point against this specific MoA.

Overall, it can be concluded that the relevance of the observed lung tumours in mice for human cannot be excluded. It is most likely that a combination of the described modes of action is the underlying cause of the cumene induced lung tumours.

### Liver tumours

A weak but statistically significant increase in liver tumours was seen in female mice, as demonstrated by the combined incidence of hepatocellular adenoma and carcinoma in exposed females (25/50, 25/50, 29/50, 36/50\* (\*  $p < 0.05$ ; P for trend = 0.024), for further details see the table below). There was a slight trend with dose, and historical controls were exceeded at all doses.

Dose (ppm)	Incidence of liver lesions							
	Males				Females			
	0	250	500	1000	0	125	250	500
Eosinophilic foci	6/50	5/50	16/50 **	14/50 *	8/50	11/50	7/50	14/50
Hepatocellular adenoma, multiple	17/50	20/50	22/50	26/50	9/50	13/50	9/50	10/50
Hepatocellular adenoma (includes multiple)	34/50	33/50	37/50	35/50	18/50	23/50	27/50	29/50 *
Hepatocellular carcinoma, multiple	3/50	1/50	4/50	7/50	2/50	1/50	2/50	0/50
Hepatocellular carcinoma (includes multiple)	13/50	18/50	21/50	17/50	10/50	7/50	6/50	12/50
Hepatocellular adenoma or carcinoma	40/50 a	42/50 a	43/50 a	41/50 a	25/50 b	26/50 b	29/50 b	36/50 b*

<sup>a</sup> Historical control data: 264/449 (58,8% ± 9,6%), range 50% - 80%

<sup>b</sup> Historical control data: 145/447 (32,4% ± 8,8%), range 22% - 50%

\* ≤ 0.05, \*\* ≤ 0.01, \*\*\* ≤ 0.001 (Poly-3 test)

### a) Genotoxicity:

There are only minor indications for direct interaction of cumene or its metabolites with liver

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DNA. Though significant DNA damage was seen in the Comet assay in the liver of male rats, this was not observed in the liver of mouse. For a detailed discussion of genotoxicity see section on genotoxicity and lung tumours above. Overall indications for genotoxicity in the liver are limited, but involvement of primary or secondary genotoxicity in the formation of liver tumours cannot be excluded.

### b) Cytotoxicity:

In the sub-chronic study in mice some minimal focal liver inflammation was observed, though without dose-response. Liver weight was also increased reaching up to 38% in males and females at the top dose. Eosinic foci and necropsy was increased in the top dose males, but tumours were seen in females. Based on these findings it can be concluded that cytotoxicity is not a relevant contributor to the observed tumours.

### c) Possible involvement of nuclear receptor activation in the formation of liver tumours:

In the CLH report the DS concluded that there were no mechanistic data to link cumene induced liver tumours to neither CAR- nor PPAR  $\alpha$ -activation and, in consequence, potentially non-relevance for humans. An increase in liver weights in the absence of considerable liver toxicity might be supportive for this MoA.

In response to comments received during the consultation the DS concluded that a CAR mediated MoA was likely and supported by new data from a pilot *in vivo* study on mouse liver provided during consultation. Observations supporting this MoA were increased liver weight and absence of relevant liver toxicity, strong induction of Cyp2b and increased cell proliferation. Deficiencies in the argumentation for this MoA are that key events (KE) 1 to 3 are only supported for oral exposure and hyperplasia and liver foci, which would support that KE 4 was not consistently shown in the NTP study (only in male animals, while tumours were seen in female animals). For details on the postulated MoA see e.g. Peffer *et al.* (2018). The preliminary study submitted by industry during consultation and the results are summarised under "Supplemental information - In depth analyses by RAC".

RAC concludes that the carcinogenic signal in female mice is not very strong and the proposed CAR/PXR mediated MoA is plausible. However, not all mechanistic studies required to demonstrate this MoA are available and some findings in the newly submitted study do not support the proposed MoA. Importantly, human relevance has not been investigated. In conclusion, the relevance of the observed tumours for humans cannot be completely dismissed.

### Spleen tumours

A slight increase in haemangiosarcoma of the spleen was seen in male mice at the top dose. The DS did not put much weight on these tumours as the increase was only marginally above the historical control values. The DS further mentioned that haemangiosarcomas occur in multiple tissue types and are not specific to or rare in the spleen (NTP, 2013). When taking the haemangiosarcomas of all tissues together, no increase above historical control was evident.

Haemangiosarcoma incidence in spleen: 0/50, P = 0.002, 0/50, 0/49, 4/50\* at 0, 250, 500 and 1000 ppm.; HCD: 6/444, (1.4%  $\pm$  1.5%), range 0% - 4%.

Haemangiosarcoma incidence, all organs: 0/50, P = 0.015, 1/50, 2/50, 4/50\* at 0, 250, 500 and 1000 ppm.; HCD: 21/450, (4.7%  $\pm$  3.7%), range 0% - 12%.

(For the chamber control values the P-value is presented which accounts for differential mortality in animals not reaching terminal sacrifice)

RAC concurs with the DS giving low weight to this tumour type.

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### Follicular-cell adenoma of the thyroid gland

An increase in top dose males was not statistically significant, only the trend was significant (P = 0.01). The increase of 6% was within the historical control range (0 - 6%) for inhalation studies and for all routes. No significant increase in follicular hyperplasia was observed.

Follicular-cell, hyperplasia: 7/50, 7/50, 7/49, 11/50 at 0, 250, 500 and 1000 ppm.

Follicular-cell, adenomas: 0/50, P = 0.010, 0/50, 0/49, 3/50 at 0, 250, 500 and 1000 ppm; HCD: 5/441 (1.1% ± 2.0%), range 0 - 6%.

(For the chamber control values the P-value is presented which accounts for differential mortality in animals not reaching terminal sacrifice)

RAC concludes that the finding is of low biological significance.

### **Rats**

In the carcinogenicity study, survival and body weight changes were similar between the exposed groups and the chamber controls. Body weight changes were not observed. There were no clinical signs related to exposure to cumene. Therefore, RAC noted that no excessive general toxicity was observed in the treated groups. The top dose was chosen based on the kidney histopathological findings (suggestive of kidney toxicity) observed at 1000 ppm in the 3-month inhalation rat study.

### Kidney tumours

In male rats, renal tubule adenoma was increased in all exposed groups above the historical control range of the laboratory (consisting of nine NTP studies performed by inhalation between 1995 and 2005). The incidences of renal tubule carcinoma were increased at ≥ 500 ppm and exceeded the historical control ranges. The increases were not statistically significant, and no dose-response was observed.

A statistically significant dose-related increase in renal tubule hyperplasia was observed in males at 500 ppm. According to NTP, this lesion was distinguished from regenerative epithelial changes and considered as a preneoplastic lesion. Mineralization of the renal papilla was significantly increased in all dose groups of males, consistent with mineralisation associated with α<sub>2</sub>u-globulin nephropathy. Additionally, an increase in the severity of chronic progressive nephropathy was observed in both males and females. A significant increase in kidney weight was noted in all exposed groups in males and in the mid and top dose group in females in the 3-month inhalation rat study of cumene.

RAC considers the increase in kidney renal tumours in males treatment related.

The incidences of kidney tumours in male rats are shown in the table below:

	<b>Kidney tumour incidence (overall rate, %)</b>				
<b>Dose (ppm)</b>	<b>0</b>	<b>250</b>	<b>500</b>	<b>1000</b>	<b>HC</b>
<b>Males: renal tubule</b>					
Adenoma	2	8	10	8	0-2
Carcinoma	2	2	6	6	0-2
Carcinoma or adenoma	4	10	16*	14	0-4

HC: historical control; \*\* p<0.01;

In the table below, selected non-neoplastic kidney findings at termination are provided:

	<b>n=50</b>							
	<b>Males</b>				<b>Females</b>			
<b>Dose (ppm)</b>	<b>0</b>	<b>250</b>	<b>500</b>	<b>1000</b>	<b>0</b>	<b>250</b>	<b>500</b>	<b>1000</b>

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Nephropathy (severity)	47 (2.3)	47 (2.6)	47 (2.9)	50 (2.7)	38 (1.4)	37 (1.5)	41 (1.9)	44 (1.9)
Renal tubule hyperplasia	-	3	8**	6*	-	-	-	-
Pelvis transitional epithelium hyperplasia	3	5	14**	15**	-	-	-	-
Papilla mineralization	5	35**	44**	41**	-	-	-	-

\*\*p≤0.01, \*p≤0.05 (poly-3 test)

The DS postulated that the accumulation of a chemical- $\alpha$ 2u-globulin complex resistant to lysosomal degradation in male rats results in renal tubular cell death and compensatory cell proliferation and neoplasms. This MoA is specific to male rat, as this protein does not exist in humans and to a much lesser extent in female rats (ECHA guidance on CLP criteria, 2017).

The following key events were considered by the DS:

- reversible binding of cumene or cumene metabolite(s) to  $\alpha$ 2u-globulin;
- increased number and size of hyaline droplets in renal proximal tubule cells;
- the hyaline droplets contained  $\alpha$ 2u-globulin;
- histopathological changes in shorter-term studies, renal tubular cell proliferation and induction of tumours.

A treatment-related increase in the amount of  $\alpha$ 2u-globulin, hyaline droplets accumulation in the cortex and medullary granular cast were observed in the 3-month inhalation rat study (NTP, 2009). According to the NTP report, kidney weight and the incidence and severity of renal cortical tubule regeneration were increased. In cell proliferation analysis, kidney cell labelling index was not statistically significantly different from control, but the number of cells in the S-phase was increased. It is noticeable that incidence of kidney nephropathy was very high in controls (97%).

Overall, RAC considered the proposed MoA plausible. RAC notes the absence of specific data on binding affinity to  $\alpha$ 2u-globulin to strengthen the case. Moreover, a clear correlation between chronic nephropathy and renal tubule adenoma or carcinoma is difficult to establish due to the very high background in controls.

### Nasal tumours

An increased incidence of respiratory epithelium adenoma was noted in all treated groups in both male and female rats above the historical control ranges (0-2% in males, not seen in females). The increase was statistically significant in males only. No dose-relation was seen.

The incidences of neoplastic and non-neoplastic lesions in male and female rats are shown in the table below:

	Incidence of nose lesions							
	Males				Females			
	Dose (ppm)	0	250	500	1000	0	250	500
Number of animals	50	50	49	50	50	48	50	50
Respiratory epithelium Adenoma (multiple and all sites)	0	7**	18**	10**	0	5*	4	3
Olfactory epithelium hyperplasia	0	19**	27**	26**	0	14**	25**	31**
Respiratory epithelium hyperplasia	0	15**	16**	23**	0	0	4	6*
Goblet cell hyperplasia	3	11*	7	5	-	-	-	-

\*\*p ≤ 0.01, \*p ≤ 0.05 (poly-3 test)

Overall, although no dose-response was observed, the increase in respiratory epithelium adenoma were clearly above historical control ranges and are considered treatment-related in both males and females. The benign tumour type and the low incidences in females may

decrease the concern.

#### Tumours in testis

A statistically significant increase in top dose males and a positive trend for unilateral and bilateral interstitial cell adenoma of the testis was observed. HCD: 345/449.

Dose (ppm)	Incidence of testis lesions			
	Males			
	0	250	500	1000
Animal number	50	50	50	50
Interstitial cell, hyperplasia	12	18	19	9
Bilateral interstitial cell, hyperplasia	0	0	0	1
Interstitial cell, adenoma	18	14	13	9
Bilateral interstitial cell, adenoma	18	24	27	37
Interstitial cell, adenoma (includes bilateral)	36*	38	40	46**

\*p = 0.006; For chamber control incidence, the p value given is associated with the trend test determined by Poly-3 test (accounts for differential mortality in animals that do not reach terminal sacrifice).

\*\*p ≤ 0.01 (compared to chamber control group determined by Poly-3 test)

As indicated by the historical control data there is a very high background incidence of this tumour type and it is noted that no progression to malignancy is reported (NTP, 2013).

Overall, RAC concludes that the finding is of minor biological relevance.

#### Other studies relevant for carcinogenicity:

The CLH report also reports two *in vitro* transformation assays (Putman, 1987b, Klimisch 1 and Gulf Oil Corporation, 1984, Klimisch 3). The retest by Putman (1987b) did not confirm the positive result from the study by Gulf Oil Corporation. RAC concludes that the two studies have no strong impact on the conclusion on classification.

#### **Overall conclusion on classification and comparison with CLP criteria:**

As there is no evidence of carcinogenicity in humans reported in the dossier, classification in Category 1A is not appropriate.

Based on the results of the NTP (2009) study in mice and rats there is clear evidence for carcinogenicity in animals.

According to the CLP regulation (Annex I: 3.6.2.2.4), additional considerations like human relevance and background incidences as part of a weight of evidence approach have to be taken into account for a classification for carcinogenicity. These are assessed in the following table:

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Factor	Evidence with cumene	Conclusion
Tumour type Considering background incidence and HCD	Lung adenoma or carcinoma in B6C3F1 male and female mice. High spontaneous tumour. Marked dose-related increase above historical control.	Supportive of classification
	Respiratory epithelium adenomas in F344 male and female rats. Above HCD range in both sexes. Low incidences in females.	Supportive of classification
	Tubular kidney adenoma or carcinoma in F344 male rats. Above HCD range.	Supportive of classification
Multi-site responses	Yes	Increased concern
Progression of lesions to malignancy	Yes for kidney and lung tumours. No progression to malignancy for nasal tumours.	Increased concern
Reduced tumour latency	Not investigated.	-
Whether responses are in single sex or both	Both sexes in rats and mice reported tumours.	Increased concern
Whether responses are in a single species or several	Tumour formation occurred in rats and mice.	Increased concern
Structural similarity to a substance(s) for which there is good evidence of carcinogenicity	Several aromatic hydrocarbons have been investigated for their tumorigenic potential, with special focus on the induction of lung tumours. For some substances, with styrene being the reference substance, lung tumours were identified as being mouse specific (Cruzan <i>et al.</i> , 2009, 2012, TERA, 2013, US EPA; 2014: CYP2F2, Clara cells).  The NTP has conducted carcinogenicity studies of ethylbenzene administered by inhalation and reported induction of renal tubule neoplasms in male and female F344/N rats, testicular adenoma in male F344/N rats, alveolar/bronchiolar neoplasms in male B6C3F1 mice, and hepatocellular neoplasms in female B6C3F1 mice in 1999 (reported in NTP, 2009).  Ethylbenzene has an old entry in Annex VI (translated from DSD) and in a RAC opinion from 2012 carcinogenicity was not evaluated.	NI
Routes of exposure	Inhalation is a relevant route of exposure	NI
Comparison of ADME between test animals and humans	No species-specific differences identified in the available toxicokinetics studies.	NI
The possibility of a confounding effect of excessive toxicity at test doses	No excessive toxicity was found in mice or rats	NI
MoA and its relevance for humans	Lung tumours in mice:  a) Genotoxic MoA (Supported by positive results in the COMET assay in the lung and high rate of metabolism in the lung)  b) Metabolism-specific lung tumours in mice (lower relevance to human).  c) Epigenetic MoA	MoA relevant to human plausible.  Not substantiated by data  MoA plausible, supported by related

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		gene expression pattern, possibly more than one MoA active.
	α2u-globulin nephropathy MoA of kidney tumours in male rats. Lower relevance to human	MoA not relevant to human. Decreased concern
	Epithelial nasal tumours: no data to support that the tumours would not be relevant to human	Human relevance plausible

NI - no influence on the concern (neither increase nor decrease)

Cumene was demonstrated to be a multi-site carcinogen in two rodent species via inhalation. No data on other exposure routes are available. A slight genotoxic potential was however demonstrated also for the oral route (COMET assays via gavage application).

As the specific tumour types were either seen in mice or in rats, species specific modes of action as underlying cause and human relevance of the neoplastic findings need to be discussed.

For the cumene induced lung tumours seen in male and female mice, although the background incidence of this type of tumour is high in this strain of mice, the increase was considered treatment related and biologically relevant as the increase was of extensive magnitude and statistically significant and already started at the lowest dose tested. A mouse specific MoA via increased transformation of cumene involving ring oxidation by CYP2F2 in Clara cells of the lower respiratory tract was postulated. However, several findings which are normally seen with this MoA were not demonstrated: lack of cytotoxicity, lack of appreciable amounts of ring oxidation in metabolism studies, absence of differences with regard to these metabolites between rats and mice and lack of Clara cell loss (which was demonstrated for styrene). In addition, the observed *K-ras* mutations might be part of an alternative MoA. In conclusion, the lung tumours have to be regarded as relevant for humans.

For liver tumours in mice there are indications for a CAR/PXR dependent MoA. Support for this MoA comes from a pilot study submitted by industry, but not all relevant mechanistic studies were provided, and alternative MoA cannot be completely ruled out on the basis of the available data. Cytotoxicity does not appear to be a relevant contributor to tumour formation based on the chronic toxicity studies. Overall, the weak increase of benign tumours in female mice is not considered a strong indication for carcinogenicity.

For the renal tumours seen in male rats the formation of α2u-globulin and hyaline droplet formation were discussed as possible underlying MoA. Several aspects of this MoA have been demonstrated but not all of the requirements according to IARC are fulfilled (e.g. insignificant increase of non-malignant nephropathic effects in female rats). On that basis human relevance cannot be completely excluded.

Neoplasms were also seen at several different sites (i.e. testis, spleen, thyroid), but they occurred at low incidences and did not progress to malignancy.

For nasal tumours in male and female rats, human relevance can be assumed, but progress to malignancy is questionable.

In conclusion several tumours were seen in animal studies for which non-human relevance could not be clearly demonstrated.

The highest weight for classification comes from the lung tumours which provide sufficient evidence of carcinogenicity. The induced tumours were clearly malignant and were seen in both male and female mice. In addition, the mechanistic data clearly indicate that the



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postulated mouse specific MoA involving increased metabolism in Clara cells of the lower respiratory tract is unlikely for cumene. The NTP, 2009 study was considered reliable.

In addition, nasal tumours observed in male and female rats provide evidence of carcinogenicity but the evidence can be considered limited as only benign tumours were seen.

Regarding kidney tumours the proposed modes of action without human relevance are likely but, as described above, important elements needed to support these MoAs are missing. Therefore, kidney tumours also provide limited supportive evidence of carcinogenicity.

Liver tumours in female mice are considered of very low weight in the overall WoE for classification as background incidence was high in this strain of rats by inhalation. In addition, the proposed MoA (CAR-mediated tumours) is plausible but RAC notes that the MoA was not sufficiently investigated.

Neoplasms with lower weight due to lack of malignancy or low incidences were also seen at several additional sites (testis, spleen, thyroid). Nevertheless, these tumour types were not considered sufficient for classification.

A contribution of genotoxicity to the observed tumour formation cannot be excluded.

On this basis, RAC is of the opinion that the overall weight of evidence warrants a **classification as Carcinogen Category 1B** based on lung tumours in mice supported by nasal and kidney tumours in the rat.

### **Specific concentration limit**

SCLs were not covered in the CLH proposal, but one MSCA commented in the PC that cumene might fall in the low potency group and might require adequate SCLs, based on T25 calculations.

In line with the EC (1999) guidance RAC calculated the following T25 values based on the alveolar/bronchiolar adenoma or carcinoma observed in male and female mice after inhalation exposure, after correction for background exposure (NTP, 2009).

**NTP, 2009:** Species and exposure route: mouse, inhalation

6h/day, 5 days/week, 105 weeks

<b>Males, endpoint: alveolar/bronchiolar adenoma or carcinoma</b>				
Dose in ppm	0	250	500	1000
Incidence	19/50 (38%)	38/50 (76%)	42/50 (84%)	43/50 (86%)
Background correction	0%	38%	46%	48%

Dose closest to 25%: 250 ppm (38%, after background correction)

$$25/38 * 250 = 164,5 \text{ ppm} = 810 \text{ mg/m}^3 = 0,81 \text{ mg/l}$$

Inhalation volume mouse (EC, 1999): 1,8 l/h (males & females) → 10,8 l/6h

$$0,81 \text{ mg/l} * 10,8 \text{ l/6h} = 8,75 \text{ mg/6h}$$

$$\text{Weight male mouse: } 30\text{g} \rightarrow 8,75 \text{ mg} * 1000\text{g} / 30\text{g} = 292 \text{ mg/kg bw/6h}$$

$$\text{Correction for 5 days per week, 105 weeks vs 104 weeks: } 292 * 5/7 * 105/104 = 210 \text{ mg/kg bw/day.}$$

$$\text{T25} = 210 \text{ mg/kg bw/day} \rightarrow > 100 \text{ mg/kg bw/day} \rightarrow \text{low potency group}$$

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<b>Females, endpoint: alveolar/bronchiolar adenoma or carcinoma</b>				
Dose in ppm	0	125	250	500
Incidence	4/50 (8%)	31/50 (62%)	43/50 (84%)	46/50 (92%)
Background correction	0%	54%	76%	84%

Dose closest to 25%: 125 ppm (54%, after background correction)

$25/54 * 125 = 57,9 \text{ ppm} = 285 \text{ mg/m}^3 = 0,285 \text{ mg/l}$

Inhalation volume mouse (EC, 1999): 1,8 l/h (males & females) → 10,8 l/6h

$0,285 \text{ mg/l} * 10,8 \text{ l/6h} = 3,078 \text{ mg/6h}$

Weight female mouse: 25g →  $3,078 \text{ mg} * 1000\text{g} / 25\text{g} = 123 \text{ mg/kg bw/6h}$

Correction for 5 days per week, 105 weeks vs 104 weeks:  $123 * 5/7 * 105/104 = 88,6 \text{ mg/kg bw/day}$ .

T25 = 88,6 mg/kg bw/day → < 100 mg/kg bw/day → medium potency group

Industry recommended the use of a different inhalation volume than the one proposed by EC (1999), i.e. 3,25 l/h. They considered this value superior to the default value recommended in EC (1999) as it is based on plethysmograph measurements in unanaesthetised mice of the relevant strain, i.e. B6C3F1 (Chang et al., 1981, supported by US EPA, 1988).

In the RAC discussion it was further considered to use the inhalation volume recommended in the REACH guidance document, Chapter R.8 (v2.1, 2012) instead, i.e. males, 2.5 l/h, females 2.2 l/h, as these values are also applied for the derivation of e.g. DNEL values under REACH.

It was, however, noted that the inhalation volume recommended in EC (1999) was the basis for the derived potency classes recommended for classification in the same document. A change of these values would require an in-depth evaluation of all available data relevant for this purpose. A review of the procedure for the derivation of SCLs is currently conducted by the COM expert group on carcinogenicity. In case this group decides that there is a need for revising the currently recommended inhalation volumes, the concentration values could be revisited.

The derived T25 values result in the low potency for males, but in the medium potency group for females. In order to protect the more sensitive sex **RAC recommends the application of the generic concentration limit of 0.1% for the classification of mixtures containing cumene.**

### **Supplemental information - In depth analyses by RAC**

During consultation, industry submitted a mechanistic study to clarify the relevance of CAR/PXR activation to the formation of liver tumours in female mice.

In a 7-day gavage study, female mice either received corn oil (control) or cumene in corn oil at 1000 mg/kg bw/day, with 10 animals per group. The animals were implanted with osmotic mini pumps releasing 5-bromo-2'-deoxyuridine (BrdU) for the detection of cell proliferation in the liver.

Several animals were not included in the analysis e.g. because they were killed due to humane reasons (injury at the site of BrdU pump implantation) or because there was no BrdU staining in the small intestine (internal positive control). Therefore, 7-10 animals from

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the control and 8-10 animals from the treated group were examined for the investigated parameters.

The following parameters were determined:

Clinical chemistry (ALT, AST, ALP), liver histology, protein concentration of liver microsomes, enzyme activities: Ethoxyresorufin-O-deethylation (EROD), Pentoxyresorufin-O-depentylation (PROD), Benzyloxyresorufin-O-debenzylation (BROD), Benzyloxyquinoline-O-debenzylation (BQ), 12-hydroxylauric acid formation (LAH), Cyanide (CN<sup>-</sup>)-insensitive palmitoyl-CoA oxidation (PCoA) and mRNA of Cyp1a1, Cyp1a2, Cyp2b10, Cyp3a11, Cyp4a10, and Cyp4a14.

Terminal mean body weights did not differ statistically significantly between cumene-treated (n=10) and control animals (n=8). There was a statistically significant increase in absolute liver weight in female mice treated with cumene (n=10) with an associated statistically significant increase in liver to body weight ratio of 19% (Figure 1 and Table 5) when compared to control animals (n=8).

Histological examination did not document any signs typical of hepatic hypertrophy despite increased liver weights, suggesting that the initial response to high concentrations of orally administered (divided dose) cumene in this study resulted in increased hepatocyte proliferation.

<b>Parameter</b>	<b>Change at 1000 mg/kg bw/day compared to control</b>	<b>Parameter</b>	<b>Change at 1000 mg/kg bw/day compared to control</b>
Clinical chemistry (ALT, AST, ALP)	No change	<u>mRNA:</u>	
protein concentration of liver microsomes	Increase: mean 1,85 fold, p>0.001)	Cyp1a1	unchanged
EROD	Increase, mean 6.58-fold, p<0.001	Cyp1a2	Increase, mean 1.3-fold, p<0.05
PROD	Increase, mean 31.9-fold, p<0.001	Cyp2b10	Increase, mean 7.6-fold, p<0.05
BROD	Increase, mean 29.49-fold p<0.001	Cyp3a11	Increase, mean 1.6-fold p<0.05
BQ	Unchanged	Cyp4a10	Increase, mean 1.9-fold, p<0.05
LAH	Unchanged	Cyp4a14	Increase, mean 1.8-fold, p<0.05
PCoA	Unchanged	Cell proliferation	Increase, mean 17.7-fold, p<0.001

Though overall the effects of cumene treatment on the enzyme contents and activities were not very strong there are some findings in support of CAR/PXR mediated response. However, as there were also increases in EROD activities, involvement of AhR activation cannot be

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ruled out on the basis of the presented data. In addition, no hypertrophy was reported, which is an associative event in the proposed MoA. However, 7 days might not be the appropriate exposure time to observe hypertrophy with cumene. In addition, it is noted that tumours were seen via the inhalation route and dosing via the oral route (gavage) in the present study might not be directly comparable. However, in the COMET assay (NTP, 2012) an oral dose of 800 mg/kg bw/day induced effects in the liver and the dose of 1000 mg/kg bw/day might therefore be adequate. As tumours were only seen in female mice, it would have been highly relevant to also investigate male mice. Peffer *et al.* (2018) mention that in the interest of animal ethics only one sex should be used if tumours were seen in both sexes, but this is not the case here.

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**10.8 Reproductive toxicity**

**10.8.1 Adverse effects on sexual function and fertility**

**Table 17: Summary table of animal studies on adverse effects on sexual function and fertility**

Method, guideline, deviations if any, species, strain, sex, no/group	Test substance, dose levels duration of exposure	Results	Reference
<p>90d, repeated dose study</p> <p>Rat, F344/N</p> <p>♂,♀,</p> <p>reproductive effects</p> <p>Similar to OECD 413.</p> <p>NTP conducts its studies in compliance with FDA Good Laboratory Practice Regulations</p> <p>Reliability according to disseminated database: study not reported</p> <p>Reliability according to authors of this evaluation: 1</p>	<p>Cumene,</p> <p>Purity: 99.9 %</p> <p>Inhalation exposure:</p> <p>3 months exposure (n=10/sex and concentration)</p> <p>0, 62.5, 125, 250, 500, and 1000 ppm</p> <p>(0, 306, 612, 1225, 2450, 4900 mg/m<sup>3</sup>)</p> <p><u>Males:</u></p> <p>Rats (0, 250, 500, and 1000 ppm) examined for</p> <p>-weight changes of cauda epididymis, epididymis and testes,</p> <p>-spermatid parameters (spermatid heads and spermatid counts), and</p> <p>-epididymal spermatozoal parameters (sperm motility, sperm concentration)</p> <p>Complete histopathology performed on 0 and 1000 ppm (core study) rats.</p> <p>Reproductive tissues examined: testis (with epididymis and seminal vesicle)</p> <p><u>Females:</u></p> <p>Rats (0, 250, 500, and 1000 ppm) examined for - Oestrus cycle length</p> <p>- Oestrus stages (% of cycle in dioestrus, prooestrus, oestrus, metoestrus)</p> <p><b>Complete histopathology performed on 0, 500, 1000 ppm (core study) rats. Reproductive tissues examined: clitoral gland, mammary gland, and uterus.</b></p>	<p><u>Males:</u></p> <p>No dose dependent significant differences between exposed and chamber control males in reproductive tissue evaluations.</p> <p>No histological changes in examined tissues from reproductive organs.</p> <p>For example:</p> <p>Testis weight (g): 1.41 ± 0.03, 1.46 ± 0.01, 1.43 ± 0.03, 1.45 ± 0.02</p> <p>Spermatid count (10<sup>6</sup>/cauda epididymis): 100.28 ± 5.52, 88.53 ± 4.55, 95.54 ± 3.36, 90.53 ± 2.32</p> <p>Sperm motility (%): 85.45 ± 3.10, 81.28 ± 2.83, 84.10 ± 2.03, 87.62 ± 1.30</p> <p>→ negative</p> <p><u>Females:</u></p> <p>Exposed female groups differ significantly from the chamber control females in the relative length of time spent in the oestrous stages. Exposed females spent more time in oestrus and less time in proestrus than chamber control females (not dose dependent).</p> <p>No histological changes in examined tissues from reproductive organs.</p> <p>Estrous stages (% of cycle):</p> <p>Diestrus: 49.2, 41.7, 41.7, 44.2</p> <p>Proestrus: 19.2, 14.2, 9.2, 11.7</p> <p>Oestrus: 15.8, 25.8, 28.3, 25.0</p> <p>Metestrus: 15.8, 18.3, 20.8, 19.2</p> <p>→ positive with questionable relevance</p> <p>Other toxicological endpoints assessed after three months of exposure: relative liver and kidney weights were significantly decreased in female rats at 250 ppm and above.</p> <p>Relative kidney weight (g):</p> <p>62.5 ppm: 3.322 ± 0.057</p> <p>125 ppm: 3.439 ± 0.057*</p>	<p>(NTP, 2009)</p>

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Method, guideline, deviations if any, species, strain, sex, no/group	Test substance, dose levels duration of exposure	Results	Reference
		250 ppm: 3.486 ± 0.044** 500 ppm: 3.612 ± 0.040**  Relative liver weight (g): 62.5 ppm: 30.094 ± 0.634* 125 ppm: 31.289 ± 0.412** 250 ppm: 32.286 ± 0.386** 500 ppm: 36.958 ± 0.724**  *:p≤0.05, **: p≤0.01  Markers of hepatocyte injury and hepatobiliary function were also altered at or above 250 ppm. No cross lesions and other persistent effects were observed.	
90d, repeated dose study  Mice, B6C3F <sub>1</sub> , ♂,♀,  reproductive effects  Similar to OECD 413. NTP conducts its studies in compliance with FDA Good Laboratory Practice Regulations  Reliability according to disseminated database: study not reported  Reliability according to authors of this evaluation: 1	Cumene, Purity: 99.9 %  Inhalation exposure: 3 months exposure (n=10/ sex and concentration)  <u>Males:</u> 0, 62.5, 125, 250, 500, and 1000 ppm (0, 306, 612, 1225, 2450, 4900 mg/m <sup>3</sup> ) Mice (0, 250, 500, and 1000 ppm) examined for -weight changes of cauda epididymis, epididymis and testes, -spermatid parameters (spermatid heads and spermatid counts), and -epididymal spermatozoal parameters (sperm motility, sperm concentration)  Complete histopathology was performed on 0 and 1000 ppm (core study) mice. Reproductive tissues examined: testis (with epididymis and seminal vesicle)  <u>Females:</u> 0, 62.5, 125, 250, 500, and 1000 ppm (0, 306, 612, 1225, 2450, 4900 mg/m <sup>3</sup> ) Mice (0, 125, 250, and 500 ppm) examined for -Oestrus cycle length - Oestrus stages (% of cycle in dioestrus, prooestrus, oestrus, metoestrus)  Complete histopathology was performed on 0 and 1000 ppm (core study) mice. Reproductive tissues examined: clitoral gland, mammary gland, and uterus.	<u>Males:</u>  At 1000 ppm significant reduction in cauda epididymis weight (p≤0.05) and in spermatid counts (p≤0.05), no other significant differences between exposed and exposed and chamber control mice.  No histological changes in examined tissues from reproductive organs.  For example: Cauda epididymis weight (g): 0.0196 ± 0.001, 0.019 ± 0.0007, 0.0173 ± 0.0006, 0.0171 ± 0.0006* (p≤0.05)  Spermatid count (10 <sup>6</sup> /cauda epididymis): 18.05 ± 0.95, 17.62 ± 1.11, 17.53 ± 1.04, 14.70 ± 0.87* (p≤0.05)  Sperm motility (%): 85.44 ± 1.96, 82.75 ± 2.41, 79.95 ± 2.13, 83.65 ± 2.43  → positive with questionable relevance  Other toxicological endpoints assessed after 3 months of exposure: Final mean body weights and body weight gains of males exposed to ≥250 ppm generally less than of chamber controls. Significant increases in absolute liver weights in mice exposed to ≥500 ppm, significant increases in relative liver weights at ≥125 ppm. Minimal to mild liver necrosis significantly increased in mice at 1000 ppm.  <u>Females:</u>  Exposed females do not differ significantly from the chamber control females in the relative time spent in the oestrous stages  No histological changes in examined	(NTP, 2009)

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<b>Method, guideline, deviations if any, species, strain, sex, no/group</b>	<b>Test substance, dose levels duration of exposure</b>	<b>Results</b>	<b>Reference</b>
		tissues from reproductive organs. → negative	
13w inhalations study, rats ♂, ♀ According to OECD 413  Reliability according to disseminated database: 1  Reliability according to authors of this evaluation: 3	Cumene, Purity: ≥99.94% ♂, ♀ - F344 rats, N=21 rats/sex/ group, exposure concentration: 0, 100, 500, 1200 ppm (0, 490 – 5880 mg/m <sup>3</sup> ), 13 weeks, 6h/d, 5d/w.  <u>Males:</u> Epididymides of 15 male animals per group were excised, and evaluated for sperm count and sperm morphology. Right testis of the 1200 ppm-group and control group was evaluated for stages of spermatogenesis.  <u>Females:</u> Necropsic examination of ovaries, uteri, cervix, vagina, oviducts and mammary tissue	<u>Males:</u> No cumene-related differences in the counts of testicular sperm heads or epididymal spermatozoa. Morphology and stages of spermatogenesis in the testes of 1200 ppm-group normal (1 rat exposed to 1200 ppm with diffuse testicular atrophy). For epididymal spermatozoa no abnormalities involving head portion of sperms. Only at 500 ppm-group increased frequency of sperm head abnormalities, but relatively infrequent and no dose response pattern observed.  Sperm anomalies (%): 1.4, 1.6, 3.4, 2.3 Sperm head anomalies (%): 0.5, 0.5, 1.1, 0.7 Sperm tail anomalies (%): 0.5, 0.6, 1.5, 1.0  → negative  <u>Females:</u> No cumene exposure related weight differences for ovaries compared to control were found. Lack of (adverse) findings for female rats.  → negative	(CMA, 1989a; Cushman <i>et al.</i> , 1995)  Study also reported in (ECHA Dissemination, 2018, toxicity to reproduction)

**Table 18: Summary table of human data on adverse effects on sexual function and fertility**

<b>Type of data/report</b>	<b>Test substance,</b>	<b>Relevant information about the study (as applicable)</b>	<b>Observations</b>	<b>Reference</b>
No human data on adverse effects on sexual function and fertility from cumene exposure are available				

**Table 19: Summary table of other studies relevant for toxicity on sexual function and fertility**

<b>Type of study/data</b>	<b>Test substance,</b>	<b>Relevant information about the study (as applicable)</b>	<b>Observations</b>	<b>Reference</b>
No other relevant studies on cumene are available, which are relevant for reproductive toxicity classification assessment				

**10.8.2 Short summary and overall relevance of the provided information on adverse effects on sexual function and fertility**

There are no data on adverse effects on sexual function and fertility from human studies. No experimental studies on fertility following OECD guidelines are available. Therefore, observations on endpoints such as fertility impairment in either male or female experimental animals are not reported. However, subchronic

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repeated dose studies specifically addressed some indicative parameters, relevant for the assessment of adverse effects on sexual function and fertility. From those data there is only very limited indication of some adversity in reproductive parameters in experimental animals:

- In male B6C3F<sub>1</sub> mice at high concentrations (1000 ppm inhalation exposure) a significant reduction in cauda epididymis weight was observed and spermatid count was reduced (NTP, 2009). This concentration caused already significant reductions in body weight gain and was a hepatotoxic exposure concentration.
- In male F344 rats at high concentrations (1200 ppm inhalation exposure) one rat showed diffuse testicular atrophy. At 500 ppm increased frequency of sperm head abnormalities have been found, but not in a dose-related manner and relatively infrequent (Cushman *et al.*, 1995). However, at (and below) 1000 ppm no adverse effects on testes or sperm parameters were seen in the study by NTP (2009).
- In female F344 rats the relative length of time spent in the oestrus stages were shifted compared to chamber control (NTP, 2009). This effect was not dose-related. Female rats in this study showed signs of hepatotoxicity and kidney weight changes at exposure concentrations leading to changes in oestrus cycle.

No other possible impairments of reproductive function were reported. This includes a study by Darmer *et al.* (1997) on developmental toxicity (see Table 20 for details) with some examinations on reproductive parameters of pregnant does.

### 10.8.3 Comparison with the CLP criteria

The available data were compared with the CLP criteria. In general, any effect of substances that has the potential to interfere with sexual function and fertility, is addressed. This includes, but is not limited to, alterations to the female and male reproductive system, adverse effects on onset of puberty, gamete production and transport, reproductive cycle normality, sexual behaviour, fertility, parturition, pregnancy outcomes, premature reproductive senescence, or modifications in other functions that are dependent on the integrity of the reproductive systems. For potential classification of cumene, classification criteria were analysed accordingly:

- *Category 1: Known or presumed human reproductive toxicant. Substances are classified in Category 1 for reproductive toxicity when they are known to have produced an adverse effect on sexual function and fertility, or when there is evidence from animal studies, possibly supplemented with other information, to provide a strong presumption that the substance has the capacity to interfere with reproduction in humans. The classification of a substance is further distinguished on the basis of whether the evidence for classification is primarily from human data (Category 1A) or from animal data (Category 1B).*

There are no data on adverse effects on sexual function and fertility associated with cumene exposure supporting category 1 classification.

- *Category 2: Suspected human reproductive toxicant. Substances are classified in Category 2 for reproductive toxicity when there is some evidence from humans or experimental animals, possibly supplemented with other information, of an adverse effect on sexual function and fertility, and where the evidence is not sufficiently convincing to place the substance in Category 1. If deficiencies in the study make the quality of evidence less convincing, Category 2 could be the more appropriate classification. Such effects shall have been observed in the absence of other toxic effects, or if occurring together with other toxic effects the adverse effect on reproduction is considered not to be a secondary non-specific consequence of the other toxic effects.*

For cumene, only minor effects on male or female reproductive capacity were observed. These effects are not sufficient for classification to Category 2 (see also below).

- *No classification: If, in some reproductive toxicity studies in experimental animals the only effects recorded are considered to be of low or minimal toxicological significance, classification may not necessarily be the outcome.*

For cumene, the observed effects are of low or minimal toxicological significance and human relevance is questionable:



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- Changes in relative length of time spent in the oestrous stages, as indicated in a three month repeated dose study (NTP, 2009), were not dose-related and no other adverse effects on reproductive parameters were shown in female rats. Indication of liver and kidney toxicity occurred at equivalent concentrations. Another study with pregnant rats did not provide any indication on changes in gestational parameters from exposure to cumene (Darmer *et al.*, 1997).
- Testicular atrophy in a male rat at high inhalation exposures (1200 ppm) (Cushman *et al.*, 1995) may not have been causally related to cumene exposure, because this effect occurred in just one animal and is not supported by the outcome of a second study with male rats with cumene inhalation exposure (NTP, 2009) and occurred at otherwise toxic exposure concentrations.
- Similarly, sperm head abnormalities in one dose group of the above mentioned repeated dose study (Cushman *et al.*, 1995) were not dose related and only minor in degree. Moreover this isolated effect was not confirmed in the other repeated dose study (NTP, 2009).
- Another isolated effect in cauda epididymis weight and in spermatid counts occurred in mice at the highest exposure concentration (NTP, 2009). However, this effect was accompanied by significant other indications of toxicity in the male mice.

### 10.8.4 Adverse effects on development

**Table 20: Summary table of animal studies on adverse effects on development**

Method, guideline, deviations if any, species, strain, sex, no/group	Test substance, dose levels, duration of exposure	Results	Reference
Developmental toxicity study, ♀, CD Sprague-Dawley rat  Guideline study OECD 414; compliant with GLP  Reliability according to disseminated database: 1  Reliability according to authors of this evaluation: 2	Cumene Purity: >99.9%  25 pregnant ♀ rats/concentration; mated with unexposed males  6h/d; GD 6-15 0, 100, 500, 1200 ppm vapour (490-5880 mg/m <sup>3</sup> ), sacrificed at GD 21	Maternal effects: increased relative liver weight, perioral wetness and incrustation at 1200 ppm, NOAEC 100 ppm, because of reduced food consumption at and above 500 ppm on GD 6-15  NOAEC: developmental effects ≥1200 ppm  Gestational parameters not affected, no changes in number of live litters and litter size.  F1: sex ratio: not affected; no significantly increased frequencies in malformations or external variations.	(CMA, 1989b; Darmer <i>et al.</i> , 1997)  Study also reported in (ECHA Dissemination, 2018, developmental toxicity/teratogenicity, #001, key)
Developmental toxicity study, ♀, New Zealand White rabbits  Guideline study OECD 414 compliant	Cumene Purity: >99.9%  15 pregnant rabbits/concentration; mated with unexposed males  6h/d; GD 6-18 0, 500, 1200, 2300 ppm vapour (2450-11270 mg/m <sup>3</sup> ), post exposure	Maternal effects: increased relative liver weight, perioral wetness at 2300 ppm, NOAEC <500 ppm, because of reduced food consumption at and above 500 ppm on GD 6-18  NOAEC: developmental effects ≥ 2300 ppm  Gestational parameters not affected, no changes in number of live litters and litter size. Nonviable implants were found in one doe at	(CMA, 1989c; Darmer <i>et al.</i> , 1997)  Study also reported in (ECHA Dissemination, 2018, developmental

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<b>Method, guideline, deviations if any, species, strain, sex, no/group</b>	<b>Test substance, dose levels, duration of exposure</b>	<b>Results</b>	<b>Reference</b>
with GLP  Reliability according to disseminated database: 1  Reliability according to authors of this evaluation: 2	observation until GD29	500 and 1200 ppm, respectively  F1: sex ratio: not affected; no treatment related significantly increased frequencies in malformations or variations. An external variation (ecchymosis) was statistically significant only in the 500-ppm group (incidence of this variation 35.7% vs. 0% in control, but historical incidence 0-66.7% in unexposed rabbits).  → negative	toxicity/teratogenicity, #002, key)

**Table 21: Summary table of human data on adverse effects on development**

<b>Type of data/report</b>	<b>Test substance,</b>	<b>Relevant information about the study (as applicable)</b>	<b>Observations</b>	<b>Reference</b>
No human data on adverse effects on development from cumene exposure are available				

**Table 22: Summary table of other studies relevant for developmental toxicity**

<b>Type of study/data</b>	<b>Test substance,</b>	<b>Relevant information about the study (as applicable)</b>	<b>Observations</b>	<b>Reference</b>
No other relevant studies on cumene are available, which are relevant for reproductive toxicity classification assessment				

**10.8.5 Short summary and overall relevance of the provided information on adverse effects on development**

There is no available data on adverse effects on development from exposure to cumene in humans. There were no developmental effects observed in either New Zealand White Rabbits or CD Sprague Dawley rats in an OECD guideline study (OECD 414) as published by Darmer *et al.* (1997). This publication is consistent with an earlier internal study report (CMA, 1989b; c).

**10.8.6 Comparison with the CLP criteria**

Cumene is negative in developmental toxicity studies. CLP criteria for developmental toxicity do not apply to cumene.

**10.8.7 Adverse effects on or via lactation**

**Table 23: Summary table of animal studies on effects on or via lactation**

<b>Method, guideline, deviations if any, species, strain, sex, no/group</b>	<b>Test substance, dose levels, duration of exposure</b>	<b>Results</b>	<b>Reference</b>
No relevant studies on cumene are available, which are relevant for “effects on or via lactation” assessment			

**Table 24: Summary table of human data on effects on or via lactation**

Type of data/report	Test substance,	Relevant information about the study (as applicable)	Observations	Reference
No human data on adverse effects on or via lactation are available				

**Table 25: Summary table of other studies relevant for effects on or via lactation**

Type of study/data	Test substance,	Relevant information about the study (as applicable)	Observations	Reference
No other relevant studies on cumene are available, which are relevant for “effects on or via lactation” assessment				

**10.8.8 Short summary and overall relevance of the provided information on effects on or via lactation**

No data available.

**10.8.9 Comparison with the CLP criteria**

Not applicable, as no data are available for this endpoint.

**10.8.10 Conclusion on classification and labelling for reproductive toxicity**

As outlined in Section 10.8.3, a weight of evidence approach indicates that the minor observed effects on sexual function and fertility are not sufficient to meet the criteria for classification of cumene for this endpoint. However, it should be noted that adequate studies to examine fertility and other endpoints of reproductive toxicity have not been performed.

There is no evidence of adverse effects from cumene exposure on developmental toxicity (see Section 10.8.5). There are no data to assess adverse effects on or via lactation from cumene exposure. Overall the information is conclusive, but not sufficient for classification of cumene as a reproductive toxicant.

**RAC evaluation of reproductive toxicity**

**Summary of the Dossier Submitter’s proposal**

***Sexual function and fertility***

The evaluation of sexual function was based on two 90-day inhalation repeated-dose toxicity studies in rats and mice (NTP, 2009). These studies were similar to OECD TG 413 and GLP-compliant. A third rat 90-day study, investigating some reproductive parameters, published by Cushman *et al.*, 1995 was also considered but rated unreliable by the DS. In addition, some reproductive parameters examined in a developmental rat toxicity study (Darmer *et al.*, 1997) were taken into account by the DS.

No studies on fertility following OECD TG were available. In these studies, the observed effects on oestrous cycle, testicular atrophy, epididymis weight and sperm findings were considered of low or minimal toxicological significance and of questionable human relevance. Therefore, no classification was proposed by the DS for toxicity on sexual function and fertility.

**Developmental toxicity**

The assessment of developmental toxicity was based on two studies in rats and rabbits published by Darmer *et al.*, 1997. The study was performed according to OECD TG 414 (reliable with limitations) and was GLP compliant. No classification was proposed by the DS as no developmental toxicity was observed.

**Comments received during consultation**

One MS commented that in absence of adequate fertility study and based on the available 90-day studies, data are inconclusive and insufficient to conclude on classification for fertility. The DS responded that although reproductive organs were examined in 90-day studies, the absence of screening of generational studies might indeed lead to the conclusion of "data lacking".

Two industry representatives and one individual agreed with the DS that findings are insufficient to classify cumene as a reproductive toxicant (sexual function, fertility and development).

**Assessment and comparison with the classification criteria****Sexual function and fertility**

The observed findings that may indicate potential effects on sexual function and fertility, highlighted by the DS, are discussed below.

Estrous cycle findings

In the 90-day repeated-dose toxicity studies (inhalation exposure) in rats (NTP, 2009), exposed females spent significantly ( $p \leq 0.05$ ) more time in the estrous stage than chamber control females and less time on proestrus. At the top dose, mean body weight and body weight gains in all tested groups were similar to controls. Liver toxicity consisted of biochemical changes and increased liver relative weight at  $\geq 250$  ppm. In addition, kidney relative weight was slightly increased in female rats at  $\geq 250$  ppm. No necropsy findings were observed in these organs. The table below presents the estrous cycle characterisation in the 3-month NTP study.

Dose (ppm), n=10	0	250	500	1000
Estrous cycle length	5.06	4.85	4.80	4.90
Estrous stage (% of cycle)				
- Diestrus	49.2	41.7	41.7	44.2
- Proestrus	19.2	14.2	9.2	11.7
- Estrus	15.8	25.8	28.3	25
- Metestrus	15.8	18.3	20.8	19.2

Estrous cycle disruption, indicated by an extended vaginal estrous is of concern and may indicate a potential effect on ovulation. Nevertheless, among the exposed groups, no clear dose-response was observed. The increased duration of estrous had no impact on the lengthening of the cycle and acyclicity was not reported. In addition, no histopathological findings in ovary (e.g. atrophy, absence of *corporea lutea*) were noted in this study. In another study using pregnant rats (Darmer *et al.*, 1999), and in the 90-day study published by Cushman *et al.*, 1995, no findings in ovary (weight, necropsy) or other investigated reproductive parameters were noted. Overall, RAC agrees with the DS that the shift in the

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relative length of time spent in the oestrus stages is insufficient for classification. Nevertheless, RAC notes that a study investigating fertility is missing and would have been necessary to clarify this concern.

### Testicular findings in male rats

Testicular atrophy was reported by Cushman *et al.*, 1995 in one out of 15 male rats at the top dose (1200 ppm). RAC agrees with the DS that this finding was only observed in one rat and may thus not be treatment-related. Moreover, this finding was not observed in the NTP, 1999 studies. Cushman *et al.*, 1995 also observed at the mid-dose an increase in the frequency of sperm head abnormalities (0.5%, 0.5%, 1.1%, 0.7% at 0, 100, 500, 1200 ppm, respectively). RAC agrees with the DS that the finding provide only very limited indication of some adversity as the effect was not dose-related, not statistically significant and infrequent. No sperm head abnormalities were seen in the NTP, 2009 study.

### Epididymides findings in male mice

In the 90-day NTP study (exposure by inhalation), a significant reduction in cauda epididymis weight was observed at the top dose. A dose-related reduction in spermatid counts was also noted, reaching statistical significance at the top dose only. A summary of these effects is presented in the table below (mean values presented).

Dose (ppm), n=10	0	250	500	1000
Absolute cauda epididymis weight (g)	38.3	36.1	36.3	34.7** (↓9,4% vs control)
Spermatid count (10 <sup>6</sup> /cauda epididymis)	18.05	17.62	17.53	14.70* (↓19% vs control)

\*\*p≤0.01; \*p≤0.05

Testis and epididymis weights and epididymal spermatozoal measurements were not affected by treatment. At the top dose, mean body weight in males exposed to 500 and 1000 ppm were significantly less than those of the controls. Liver toxicity (weight, necrosis) was also observed at the top dose. As the effect occurs in presence of toxicity in males, RAC agrees with the DS that the observed effects are of minimal toxicological significance.

### Conclusion on fertility and sexual function

Overall, RAC agrees with the DS that the findings in the 90-day studies investigating reproductive parameters are not sufficient to classify cumene for reproductive toxicity. Nevertheless, **due to the lack of data on fertility and sexual function (e.g. a generational study), RAC was unable to evaluate this hazard class.**

### ***Developmental toxicity***

As no relevant toxicological findings were observed in the two developmental toxicity studies performed with cumene, RAC agrees with the DS that **no classification is warranted for cumene for developmental toxicity.**

### ***Adverse effects on or via lactation***

**RAC was unable to evaluate on this hazard class due to lack of data.**

**10.9 Specific target organ toxicity-single exposure**

Evaluation not performed for this substance

**10.10 Specific target organ toxicity-repeated exposure**

Evaluation not performed for this substance

**10.11 Aspiration hazard**

Evaluation not performed for this substance

**11 EVALUATION OF ENVIRONMENTAL HAZARDS**

Evaluation not performed for this substance

**12 EVALUATION OF ADDITIONAL HAZARDS**

Evaluation not performed for this substance

**13 ADDITIONAL LABELLING**

No additional labelling relevant for this substance.

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**15 ANNEXES**

*[Please add ANNEX I to the CLH report and potential other annexes.]*

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**Tabular overview on the different tumours and relevant modes of action prepared by the DS as response to the comments received during consultation (see page 11 of RAC Opinion)**

**Overview of arguments in favour/against a specific MoA for specific tumour localisations observed after inhalation exposure to cumene in the study of NTP (2009)**

The following table summarises the current view on the evidence available for specific MoAs for the various tumour types observed in the NTP studies on cumene.

Tumour location	Characteristics of tumours observed	MoA under discussion	Arguments in favour of this MoA	Arguments against this MoA	Conclusions
Lung tumours in B6C3F1 mice	Benign and malign tumours  Significant increase in both sexes  dose-dependent, mice only	CYP2F2-dependent metabolism in Clara cells	<ul style="list-style-type: none"> <li>- Direct genotoxicity unlikely MoA</li> <li>Lung tumours observed only in mice, not in rats</li> <li>- Increased retention of radioactivity in female mouse lung after repeated cumene exposure (Chen <i>et al.</i>, 2011)</li> <li>- Higher rates of metabolism to AMS and related metabolites in mouse lung microsomes <i>in vitro</i> compared to mouse liver or rat liver or lung (Chen <i>et al.</i>, 2011)</li> </ul>	<p>Analogy to other substances claimed, but no evidence for key events as shown for styrene (Cruzan <i>et al.</i> 2018):</p> <p>Study with CYP2F2-KO and CYP2F1-TG mice in comparison to B6C3F1 lacking to show:</p> <ul style="list-style-type: none"> <li>- Lung metabolism by CYP2F2</li> <li>- Differences in gene expression</li> <li>- Gene proliferation only in 2F2 +/- strains</li> </ul>	MoA with low human relevance possible but yet unproven additional work on lung tumour MoA announced by ACC
Liver tumours in female B6C3F1 mice	Benign and malign tumours  (Borderline) significant increase in females only  Slight trend with dose, mice only, but highest dose above historical control range	CAR-dependent	<p>(new data on mouse liver pilot study provided during consultation):</p> <p>Observations (see text):</p> <ul style="list-style-type: none"> <li>- Increased liver weight</li> <li>- Absence of liver toxicity</li> <li>- Strong induction of CYP2B</li> <li>- Increased cell proliferation</li> </ul>	<ul style="list-style-type: none"> <li>- KE1, KE2 and KE3 shown for oral pathway only (inhalation study planned);</li> <li>- KE4 (hyperplasia, liver foci) not consistently shown in NTP study</li> </ul>	MoA with low human relevance likely
Renal tumours in male F344/N	Benign and malign tumours	$\alpha$ 2u-globulin accumulation	According to NTP 2009 lesions observed in male rats characteristic	Cell proliferation: labelling index not significantly increased (but mean numbers of proximal tubule cells in	MoA with low human relevance likely

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rats	(Borderline) significant increase in males only  dose-dependent, rats only, but exposure groups above historical control range		of α2u-globulin accumulation: - α2u -globulin and protein concentrations in kidney cells increased - mineralization of the renal papilla - exposure-related increase in hyaline droplets (findings in 3-month study) - high conc. of radioactivity in male kidneys (Chen <i>et al.</i> 2011)	S-phase significantly increased at 500 and 1000 ppm; hyperplasia in chronic study)	
Nasal tumours in F344/N rats	Benign tumours only (with unclear progression to malignancy)  Significant increase in both sexes  dose-dependent in males only, rats only	CYP2F2-dependent metabolism	- Higher activity of CYP2F enzymes in rat olfactory epithelium, - No progression to malignancy observed in 2 year study	- Evidence for link to CYP2F-mediated metabolism lacking - no known MoA for induction of hyperplasia in olfactory and respiratory epithelium	MoA for benign tumours in one species unknown; additional work on MoA announced by ACC

**Direct genotoxicity**

Overall, mutagenicity tests with cumene were mostly negative (Ames tests and *in vitro* CHO/HGPRT tests, see the detailed description in the BD). Weakly positive results were observed for cumene in *in vivo* Comet assays (weakly positive for liver, male rats and positive for lung, female rats) (NTP, 2012). As pointed out in the response to comment #14, high variability was observed in the sparsely available historical control data for the selected animal strain. Therefore, the reliability of the positive results includes some uncertainty.

Also, the metabolite AMS was mostly negative in *in vitro* mutagenicity tests. Only in two SCE assays (weakly) positive results were observed with AMS (Norppa and Vainio, 1983; NTP, 2007) in cultured human lymphocytes and Chinese hamster ovary cells. The assay does not offer the possibility to distinguish between direct or indirect genotoxicity. However, based on the observations, direct genotoxicity remains a possibility for the observed positive effects. AMS can be metabolised to AMS oxide, but this substance has not been confirmed as a metabolite of cumene *in vivo* so far.

Lung tumours in mice found in the 2-year carcinogenicity study by NTP were analysed for mutations of *K-ras* and *p53* (Hong *et al.*, 2008). A detailed discussion of the conclusions that can or cannot be drawn from the occurrence of these mutations is given below in the section "Lung tumours in mice – Cyp2f2-dependent MoA".



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In conclusion, direct genotoxicity is unlikely for cumene, but cannot be excluded completely.

### **Indirect genotoxicity**

In a FLARE test (Fragment Length Analysis with Repair Enzyme) to clarify the DNA damage from reactive oxygen species, some indications for oxidative DNA damage from cumene exposure were found (Kim *et al.*, 2008). However, no clear duration-response relationship was observed and the study is qualified as being insufficient in reporting of methods and results by NTP (2013).

Generation of ROS is in general associated with inflammation processes that can be observed in the histopathological examination of the relevant tissue. In case of the lung of mice no increased inflammation was observed on exposed mice compared to control.

Therefore, indirect genotoxicity mediated by the production of reactive oxygen species does not seem a satisfying explanation for the carcinogenic activity observed in animal studies.

Wakamatsu *et al.* (2008) studied the potential involvement of epigenetic mechanisms in cumene-induced lung cancer. They observed significant alterations in genes associated with the histone deacetylase complex (HDAC) in mouse lung carcinomas. A stronger association was observed between altered genes supposedly associated with HDACs and tumours with *K-ras* mutations compared to tumours without *K-ras* mutations. Therefore, *K-ras* activation may affect histone modification or vice versa (NTP, 2013).

There is not enough information to conclude on either of these MoA.

### **Lung tumours in mice – CYP2F2-dependent MoA**

A CYP2F2-dependent metabolism in lung Clara cells (club cells) was postulated and, consequently, the observed lung tumours in mice were considered species-specific by some commentators. As listed in the table, limited information is available to confirm this MoA. Structural similarity to substances, for which this MoA was confirmed (e.g. styrene) is not sufficient to assume that the same MoA is active for cumene as well. Specifically, information is required that metabolism in lung cells, altered gene expression and cell proliferation (key events for this MoA according to Cruzan *et al.* (2018) are CYP2F2 dependent. One commentator pointed out that respective studies are under way.

Further, increased occurrence of *K-ras* and/or *p53* mutations in cumene-induced mouse lung tumours compared to spontaneous lung tumours were found. The mutational spectra of *K-ras* and *p53* in these lung tumours differed from those observed in spontaneous lung tumours and the molecular alterations resemble those found in human lung and other cancers. From the available data it is not clear whether this increase in specific mutations is caused by cumene or is a feature of the lung tumours as they developed in consequence of cumene exposure. Therefore, and because the metabolite  $\alpha$ -methylstyrene (AMS) was shown to be formed by mice lung microsomes *in vitro* (Chen *et al.*, 2011), which might give rise to the genotoxic metabolite AMS oxide, genotoxicity cannot be completely ruled out, although, based on the existing evidence from genotoxicity studies, direct genotoxicity as the dominant MoA is considered unlikely. Nevertheless, due to the uncertainty regarding the MoA, a threshold for the carcinogenic activity cannot be determined.

### **Liver tumours in mice – CAR-dependent MoA**

During consultation, preliminary findings from an *in vivo* mouse study investigating the MoA for the observed liver tumours in female B6C3F1 mice was submitted by the American Chemistry Council (document ACC Comments on Cumene CLH Proposal 11 22 19.pdf, additional details on this study are reported in a separate, confidential document, ACC Cumene Research Cons Final Int Report Liver Pilot

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Excerpt.pdf). In this study, female C57BL/6 mice were treated by gavage for 7 days with 1000 mg/kg/day (daily dose split in two doses of 500 mg/kg each). Animals had mini-pumps filled with BrdU implanted to study cell proliferation. Main observations were:

- Increased liver weights
- No clinical signs of hepatotoxicity (ALT, ALP, AST not increased)
- No histopathological changes in liver, no hypertrophy observed
- Increased P450 concentrations, increased activity of CYP2B and CYP1A enzymes
- Increased mRNA levels for CYP2B10, CYP1A2, 3A11, 4A10, and 4A14
- Increase in S-phase labelling, indicating cell proliferation.

These observations are largely in support of Key events 1, 2, and 3 of the MoA framework for substances inducing liver tumours in rodents by a constitutive androstane receptor (CAR) MoA, as proposed recently by Cruzan *et al.* (2018). Although the evidence for Key event 4 (hyperplasia, liver foci) is limited (only shown in male animals), this needs to be discussed in connection to the borderline significance of the liver tumours observed. Overall, these results are indications for a CAR-like MoA for liver tumours observed in female mice in the NTP study. In a recent workshop on non-genotoxic MoA for rodent liver tumours “most, but not all, participants considered the CAR [...] MoA as not relevant to humans” (Felter *et al.*, 2018).

### **Renal tumours in male F344/N rats**

Information from the NTP (2009) study with rats regarding a  $\alpha$ 2u-globulin-associated MoA:

- $\alpha$ 2u-globulin and protein concentrations in kidney cells increased
- mineralization of the renal papilla observed
- exposure-related increase in hyaline droplets (findings in 3-month study)
- High concentration of radioactivity in male kidneys (Chen *et al.* 2011)
- mean numbers of proximal tubule cells in S-phase increased (indicating cell proliferation)

As discussed by NTP (2013) not all criteria for confirming  $\alpha$ 2u-globulin nephropathy as the dominant MoA (due to slight nephropathy in female animals, no evidence of sustained cell proliferation in the renal cortex, uncertainty about potential genotoxic metabolites such as AMS oxide) are fulfilled, but together with the fact that renal tumours were observed only in rats and only in males point to  $\alpha$ 2u-globulin-induced nephropathy as the most likely MoA.

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### **Nasal tumours in F344/N rats**

Clear dose-related increased incidences of benign nasal tumours (adenoma of the respiratory epithelium) were observed in rats of both sexes. According to NTP (2013) „this type of tumour typically does not progress to malignancy“, although the basis for this conclusion is not completely clear. No malignant forms were observed in the NTP study after 2 years. A Cyp2f2-mediated MoA was suggested by commentators, based on high content in the olfactory epithelium, but no further information is available to substantiate the role of this MoA. A commentator announced further mechanistic investigations on these tumours.

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