

European Union Risk Assessment Report

TRIS (NONYLPHENYL) PHOSPHITE

CAS No: 26523-78-4
EINECS No: 247-759-6

RISK ASSESSMENT

GENERAL NOTE

This report contains different documents:

- **Environment**

Version August 2007 (pages 76 - conclusions resumed on the last one)

- **Human Health**

Version February 2006 (pages 88 - conclusions resumed on the last one)

European Union Risk Assessment Report

TRIS (NONYLPHENYL) PHOSPHITE

CAS-No.: 26523-78-4
EINECS-No.: 247-759-6

RISK ASSESSMENT

DRAFT

LEGAL NOTICE

Neither the European Commission nor any person acting on behalf of the Commission is responsible for the use which might be made of the following information

A great deal of additional information on the European Union is available on the Internet.
It can be accessed through the Europa Server (<http://europa.eu.int>).

DRAFT

Cataloguing data can be found at the end of this publication

Luxembourg: Office for Official Publications of the European Communities, 2002
ISBN ([insert number here](#))

© European Communities, 2002 ([insert correct year here](#))
Reproduction is authorised provided the source is acknowledged.

Printed in Italy

TRIS(NONYLPHENYL) PHOSPHITE

CAS No.: 26523-78-4
EINECS No.: 247-759-6

RISK ASSESSMENT

Draft of August 2007

France

Rapporteur for the risk assessment of tris(nonylphenyl) phosphite is the Ministry of Ecology and Sustainable Development as well as the Ministry of Employment and Social Affairs in co-operation with the Ministry of Public Health. Responsible for the risk evaluation and subsequently for the contents of this report is the rapporteur.

The scientific work on this report has been prepared by the National Institute for Industrial Environment and Risks (INERIS) and by the Chemical Risk Assessment Bureau (BERPC).

Contact point:
BERPC
60-62 rue d'Hauteville
75010 Paris
FRANCE

Date of Last Literature Search :

[please insert year]

Review of report by MS Technical Experts finalised:

[please insert month and year]

Final report:

[please year]

DRAFT

Foreword

This Draft Risk assessment Report is carried out in accordance with Council Regulation (EEC) 793/93¹ on the evaluation and control of the risks of “existing” substances. “Existing” substances are chemical substances in use within the European Community before September 1981 and listed in the European Inventory of Existing Commercial Chemical Substances. Regulation 793/93 provides a systematic framework for the evaluation of the risks to human health and the environment of these substances if they are produced or imported into the Community in volumes above 10 tonnes per year.

There are four overall stages in the Regulation for reducing the risks: data collection, priority setting, risk assessment and risk reduction. Data provided by Industry are used by Member States and the Commission services to determine the priority of the substances which need to be assessed. For each substance on a priority list, a Member State volunteers to act as “Rapporteur”, undertaking the in-depth Risk Assessment and recommending a strategy to limit the risks of exposure to the substance, if necessary.

The methods for carrying out an in-depth Risk Assessment at Community level are laid down in Commission Regulation (EC) 1488/94², which is supported by a technical guidance document³. Normally, the “Rapporteur” and individual companies producing, importing and/or using the chemicals work closely together to develop a draft Risk Assessment Report, which is then presented at a Meeting of Member State technical experts for endorsement. The Risk Assessment Report is then peer-reviewed by the Scientific Committee on Toxicity, Ecotoxicity and the Environment (CSTEE), now renamed Scientific Committee on Health and Environmental Risks (SCHER) which gives its opinion to the European Commission on the quality of the risk assessment.

This Draft Risk Assessment Report has undergone a discussion in the Competent Group of Member State experts with the aim of reaching consensus by interpreting the underlying scientific information, or including more data, but this work has not yet been totally finalised. The information contained in this Draft Risk Assessment Report does not, therefore, necessarily provide a sufficient basis for decision making regarding the hazards, exposures or the risks associated with the priority substance.

This Draft Risk Assessment Report is under the responsibility of the Member State rapporteur. In order to avoid possible misinterpretations or misuse of the findings in this draft, anyone wishing to cite or quote this report is advised to contact the Member State rapporteur beforehand.

¹ O.J. No L 084, 05/04/199 p.0001 – 0075

² O.J. No L 161, 29/06/1994 p. 0003 – 0011

³ Technical Guidance Document, Part I – V, ISBN 92-827-801 [1234]

0

OVERALL RESULTS OF THE RISK ASSESSMENT

CAS Number: 26523-78-4
EINECS Number: 247-759-6
IUPAC Name: Phenol, nonyl-, phosphite (3:1)

Environment

Section not updated, see section 3.3.

Human exposed via the environment

This section will be updated taking into account the refinement of the other parts of the risk assessment.

DRAFT

CONTENTS

1	GENERAL SUBSTANCE INFORMATION	5
1.1	IDENTIFICATION OF THE SUBSTANCE.....	5
1.2	PURITY/IMPURITIES, ADDITIVES	5
1.2.1	Purity	5
1.2.2	Additives.....	6
1.3	PHYSICO-CHEMICAL PROPERTIES	6
1.3.1	Physical state (at ntp).....	6
1.3.2	Melting point	6
1.3.3	Boiling point.....	6
1.3.4	Relative density	6
1.3.5	Vapour pressure.....	6
1.3.6	n-octanol / water partition coefficient.....	8
1.3.7	Water solubility	9
1.3.8	Flash point	10
1.3.9	Autoflammability.....	10
1.3.10	Explosivity.....	10
1.3.11	Oxidising properties.....	10
1.3.12	Viscosity	10
1.3.13	Henry's Law constant.....	10
1.3.14	Summary of physico-chemical properties	12
1.4	CLASSIFICATION	13
1.4.1	Current classification	13
1.4.2	Proposed classification	13
2	GENERAL INFORMATION ON EXPOSURE	14
2.1	PRODUCTION.....	14
2.1.1	Production process.....	14
2.1.2	Production capacity	15
2.2	USES 16	
2.2.1	Introduction	16
2.2.1.1	Industrial use	17
2.2.1.2	Production of Polyvinylchloride (PVC) film.....	18
2.2.1.3	Production of Polyolefins linear low density polyethylene (LLDPE)	18
2.2.1.4	Production of High density polyethylene (HDPE)	18
2.2.1.5	Production of rubber	18
2.2.2	Other applications.....	19
2.2.2.1	Use of end-products.....	19
2.2.2.2	Recovery and disposal.....	19
3	ENVIRONMENT RISK ASSESSMENT	20
3.1	ENVIRONMENTAL EXPOSURE	20
3.1.1	Environmental fate.....	20
3.1.1.1	Degradation in the environment	20
3.1.1.1.1	Atmospheric degradation.....	20
3.1.1.1.2	Aquatic degradation - hydrolysis.....	21
3.1.1.1.3	Aquatic degradation - oxidation.....	25
3.1.1.1.4	Aquatic degradation - biodegradation.....	26
3.1.1.1.5	Degradation in soil.....	27
3.1.1.1.6	Summary of environmental degradation.....	27
3.1.1.2	Distribution.....	28

3.1.1.2.1	Adsorption	28
3.1.1.2.2	Precipitation.....	28
3.1.1.2.3	Volatilisation.....	28
3.1.1.2.4	Distribution in waste water treatment plants.....	29
3.1.1.2.5	Accumulation and metabolism	30
3.1.2	Environmental releases.....	30
3.1.2.1	General information.....	30
3.1.2.2	Use of the OECD ESD for plastic additives with TNPP	32
3.1.2.3	Use of the OECD ESD for additives used in rubber industry with TNPP.....	33
3.1.2.4	Calculation of NP releases due to the processing of TNPP	34
3.1.2.5	Sensitivity analysis of the exposure assessment for ranges of log Kow and water solubility (and subsequent associated parameters)	36
3.1.2.6	Regional releases	38
3.1.2.7	Regional concentrations.....	38
3.1.2.8	Releases during TNPP production.....	39
3.1.2.9	Releases during the use in PVC films or LLDPE films	39
3.1.2.10	Releases during the use in rubber	39
3.1.2.11	Releases during the use in HDPE films	40
3.1.2.12	Releases during the use in other applications	40
3.1.3	Aquatic compartment (incl. sediment).....	40
3.1.3.1	Predicted environmental concentrations in water	40
3.1.3.1.1	PEC _{local(water)} at TNPP production sites	40
3.1.3.1.2	PEC _{local(water)} at sites using TNPP.....	41
3.1.3.1.3	PEC _{local(water)} for emissions during products service life	42
3.1.3.2	Calculation of Predicted Environmental Concentration for Sewage Treatment Plants (PEC _{stp})	42
3.1.3.3	Calculation of PEC _{sediment}	43
3.1.4	Marine compartment.....	43
3.1.5	Terrestrial compartment.....	43
3.1.5.1	Calculated PEC for soil	43
3.1.5.1.1	PEC _{local soils} at TNPP production sites	44
3.1.5.1.2	PEC _{local soils} at sites using TNPP	44
3.1.5.1.3	PEC _{local soils} during products service life	45
3.1.6	Atmospheric compartment.....	45
3.1.6.1.1	PEC _{local air} at TNPP production sites	46
3.1.6.1.2	PEC _{local air} at sites using TNPP.....	46
3.1.6.1.3	PEC _{local air} during products service life	46
3.1.7	Non compartment specific effects relevant for the food chain (Secondary poisoning)	46
3.2	EFFECTS ASSESSMENT: HAZARD IDENTIFICATION AND DOSE (CONCENTRATION) - RESPONSE (EFFECT ASSESSMENT)	48
3.2.1	Aquatic compartment (including sediment).....	48
3.2.1.1	Fish	48
3.2.1.1.1	Acute toxicity.....	48
3.2.1.1.2	Long-term toxicity	50
3.2.1.2	Aquatic invertebrates	50
3.2.1.2.1	Acute toxicity.....	50
3.2.1.2.2	Long-term toxicity	53
3.2.1.3	Algae	53
3.2.1.4	Micro-organisms.....	54
3.2.1.5	Calculation of Predicted No Effect Concentration (PNEC).....	55
3.2.1.5.1	Surface water	55
3.2.1.5.2	Sewage treatment plants	55
3.2.1.5.3	Sediment	55
3.2.2	Marine compartment.....	56
3.2.3	Terrestrial compartment.....	56
3.2.3.1	Terrestrial effect data.....	56
3.2.3.2	Calculation of PNEC _{soil}	56
3.2.4	Atmosphere.....	56
3.2.5	Non compartment specific effects relevant to the food chain (secondary poisoning).....	56

3.3	RISK CHARACTERISATION	57
3.3.1	Aquatic compartment (including sediment).....	57
3.3.2	Marine compartment.....	60
3.3.3	Terrestrial compartment.....	60
3.3.4	Atmosphere.....	61
3.3.5	Non compartment specific effects relevant for the food chain (Secondary poisoning)	62
3.3.6	PBT assessment	63
3.3.6.1	PBT assessment for TNPP.....	63
3.3.6.2	PBT assessment for NP	63
4	HUMAN HEALTH.....	64
4.1.1	Human exposed via the environment.....	64
4.1.1.1	Indirect exposure via the environment.....	64
4.1.1.2	Human exposed via the environment.....	64
4.1.1.2.1	Summary of risk characterisation for exposure via the environment.....	64
5	RESULTS.....	64
5.1	ENVIRONMENT	64
5.2	HEALTH	64
6	REFERENCES.....	65

TABLES

Table 1.1:	Vapour pressure data for TNPP	7
Table 1.3:	log Kow calculations for TNPP along with the validity domain of the calculation method used.....	8
Table 1.5:	Physical and chemical properties of the TNPP	12
Table 2.1:	Typical quantitative breakdown of TNPP Uses	16
Table 2.3:	Industrial uses of TNPP in the Nordic Countries: Denmark, Norway and Sweden (in Tonnes).....	16
Table 2.5:	Use pattern of TNPP in the Nordic Countries: Denmark, Norway and Sweden (in Tonnes)	17
Table 2.7:	Global food contact regulations specific to TNPP	18
Table 3.1:	hydrolysis half-lives of three TNPP grades.....	22
Table 3.3:	Calculated partition coefficients for TNPP	28
Table 3.5:	Estimation of removal of TNPP in STPs according to EUSES.....	29
Table 3.7:	Industrial Categories (IC), Use Categories (UC) and Main categories (MC) used in EUSES calculations	31
Table 3.11:	main characteristics of nonylphenol used for the risk assessment	35
Table 3.13:	Parameters used for the calculation of local releases due to formulation and processing of TNPP	37
Table 3.15:	regional releases of TNPP.....	38
Table 3.16:	regional concentrations for TNPP.....	38
Table 3.17:	regional concentrations for NP (E.C., 2002).....	38
Table 3.18:	Local TNPP and NP (in brackets) emissions due to use in PVC films or LLDPE films	39
Table 3.20:	Local TNPP and NP (in brackets) emissions due to use in rubber.....	39
Table 3.21:	Local TNPP and NP (in brackets) emissions due to use in HDPE films.....	40
Table 3.27:	PECllocal for water for sites using TNPP	41
Table 3.29:	PECllocal for emission during products service life	42
Table 3.30:	PECllocal for sediment for sites using TNPP	43
Table 3.32:	PECllocal for the three European TNPP production sites.....	44
Table 3.34:	PECllocal,soil for European TNPP processing sites	44
Table 3.36:	PECllocal,soil for emission during the service life of products	45
Table 3.37:	PECllocal for the three European TNPP production sites.....	46
Table 3.39:	PECllocal for air for sites using TNPP.....	46
Table 3.41:	PECllocal for emissions during the service life of products	46
Table 3.42:	Predicted Concentrations for secondary poisoning	47
Table 3.44:	Summary of acute toxicity tests with fish	48
Table 3.46:	Summary of acute toxicity tests with aquatic invertebrates	50

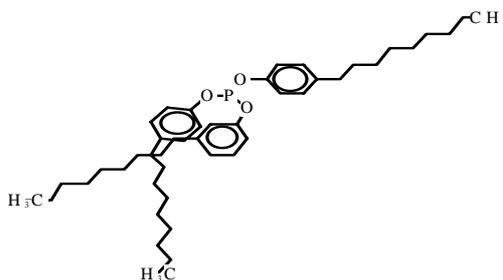
Table 3.48: Summary of toxicity tests with algae	53
Table 3.50: RCR for STP for the three European TNPP production sites	57
Table 3.52: RCR for STP and water for sites using TNPP	57
Table 3.54: RCR for STP and water for the service life of plastics containing TNPP.....	57
Table 3.55: PEClocal for sediment for sites using TNPP	58
Table 3.57: PEClocal for sediment for the service life of plastics containing TNPP.....	58
Table 3.58: PEClocal for soil for sites using TNPP	60
Table 3.60: PEClocal for soil for the service life of plastics containing TNPP	61
Table 3.61: RCRs for secondary poisoning	62

DRAFT

1 GENERAL SUBSTANCE INFORMATION

1.1 IDENTIFICATION OF THE SUBSTANCE

CAS No: 26523-78-4
EINECS No: 247-759-6
IUPAC Name: Phenol, nonyl-, phosphite (3:1)
Molecular formula: $C_{45}H_{69}O_3P$
Structural Formula:



Molecular weight: $689 \text{ g}\cdot\text{mol}^{-1}$
Synonyms and tradenames: Alkanox TNPP, Doverphos, DP4, DP4HP, Lowinox TNPP, Irgafos TNPP, Tris(monononylphenyl)phosphite, Tri(nonylphenyl)phosphite, Weston 399, Weston TNPP, Irgastab CH 55, Naugard TNPP, Polygard, Polygard HR, Polygard LC, TNPP, Trisnonylphenylphosphit.

In this assessment, the name Tris(nonylphenyl)phosphite (TNPP) will be used for the substance as this is the most common name.

1.2 PURITY/IMPURITIES, ADDITIVES

1.2.1 Purity

There are two grades of TNPP that are sold in the marketplace. The purity of the standard TNPP is reported as ca. 95 – 100% w/w.

The following impurities may be found in standard TNPP :

- Nonylphenol (CAS 25154-52-3) < 5% w/w,
- Phenol (CAS 108-95-2) < 0.1% w/w,
- Di(nonylphenyl)phenylphosphite (CAS 25417-08-7) 0.05% w/w,

A high purity grade of TNPP was introduced into the market in the late 1990s. The impurities found in the high purity TNPP are:

- Nonylphenol (CAS 25154-52-3) < 0.1% w/w,
- Phenol (CAS 108-95-2) < 0.1% w/w,
- Di(nonylphenyl)phenylphosphite (CAS 25417-08-7) 0.05% w/w,

1.2.2 Additives

1,1',1''-nitrilotripropan-2-ol (CAS No: 122-20-3), also known as tri-isopropanol amine, or TIPA, is an additive commonly found in TNPP in the proportion of 0.5 to 1% w/w. TIPA acts as an acid scavenger and increases the hydrolytic stability of TNPP.

1.3 PHYSICO-CHEMICAL PROPERTIES

1.3.1 Physical state (at ntp)

TNPP is a viscous liquid at room temperature.

1.3.2 Melting point

Instead of a melting point, a pour point of $6^{\circ}\text{C} \pm 3^{\circ}\text{C}$ was determined (Reimer&Associates, 2001c). A melting point could not be observed using the differential scanning calorimetric (DSC) method because an endothermic event was not observed in the heat flow vs temperature plot. The pour point (the lowest temperature at which the test substance is first observed to flow on warming) is an appropriate measurement for viscous liquid substances. The test was conducted according to ASTM Method D97, as recommended in the OECD 102 guideline.

1.3.3 Boiling point

The boiling point was reported as $>303^{\circ}\text{C}$ (Reimer&Associates, 2001a). The test method was based on OECD 103 guideline. Bubbling was observed for the first 1 to 2 seconds of heating, and then stopped. This was probably due to the boiling of a minor component ($<0.1\%$) present in the test substance. Consequently a new study was undertaken to assess the true boiling point. The TNPP producers have determined that TNPP will begin to degrade before boiling. According to a Thermal Gravimetric Analysis (TGA) of TNPP, the phosphite has an onset of degradation at 322°C under nitrogen.

1.3.4 Relative density

The relative density has been quoted at 0.98 g.cm^{-3} at 20°C (Crompton, 2003).

1.3.5 Vapour pressure

A vapour pressure was estimated using structure activity relationships models developed by the U.S. Environmental Protection Agency and Syracuse Research Corporation (EPIWIN, v. 3.10,

US EPA and Syracuse Research Corporation, 2001). The vapour pressure was estimated to 5×10^{-12} Pa (Staples, 2001).

A much higher value of 0.047 Pa at 20°C was extrapolated from results obtained by isoteniscope (method ASTM D2879) at temperatures ranging from 125 to 375 °C (Phoenix_Chemical_Laboratory, 1997). These measured values are displayed in Table 1.1.

Table 1.1: Vapour pressure data for TNPP

Temperature °C	Vapour Pressure (Pa)
125	22.7
150	65.3
175	160
200	373
225	747
250	1533
275	2800
300	4666
325	8133
350	15330
375	65330

A strong relation between the temperature ($1/T$) and the vapour pressure is found. Excluding the last value measured at 375°C, the plot of the above results gives a linear regression with a good reliability (see Figure 1-1). Vapour pressures of respectively 0.039 Pa and 0.058 Pa at 20°C and 25°C could be derived from this equation. These results are consistent with the extrapolated value of 0.047 Pa at 20°C found in the study summary in the IUCLID file and for which no information on the extrapolation method was available.

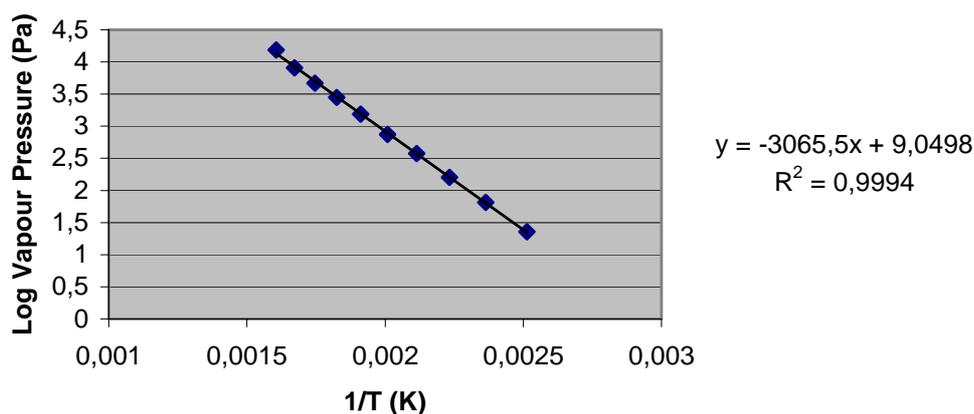


Figure 1-1: linear regression between the temperature ($1/T$) and the measured vapour pressures

The isoteniscope method is recommended for the measurement of vapour pressures between 10^2 and 10^5 Pa. The extrapolated value is three orders of magnitude below this range however almost all of the data used for the extrapolation were included in the valid range for vapour pressure.

There is speculation that the higher volatility reported in the 1997 study by Phoenix Chemical is due to the presence of high levels of nonylphenol as an impurity. To more accurately determine the vapour pressure of high purity TNPP, Industry commissioned the conduct of a new vapour pressure determination with standard TNPP and Hi Pure TNPP. The results of this evaluation should be available in early September. Until such time as newer information is obtained, the value of 0.058 Pa at 25°C will be retained for this assessment.

1.3.6 n-octanol / water partition coefficient

The n-octanol-water partition coefficient was estimated using structure activity relationships models developed by the U.S. Environmental Protection Agency and Syracuse Research Corporation (EPIWIN, US EPA and Syracuse Research Corporation, 2001). The log P_{ow} was estimated to 20.05 (US EPA and Syracuse Research Corporation, 2001).

According to Reimer & Associates, 2001c, it was not appropriate to conduct the partition coefficient measurement because the solubility of TNPP in water was too low (see section 1.3.7). Moreover, the reaction of TNPP with alcohol and consequently with octanol does not allow the measurement of adequate TNPP concentrations in octanol. The n-octanol / water partition coefficient was therefore calculated using the software from Advanced Chemistry Development Inc. ("ACD/LogP DB"). The result of the calculation was found to be 21.6 ± 0.6 (Reimer&Associates, 2001d).

The annex of the OECD guideline 117 presents some K_{ow} calculation methods that can be used to "provide an estimate when experimental methods cannot be applied". However there are some limitations to the use of such methods. First, the reliability of calculation methods decreases as the complexity of the compound under study increases. Here, TNPP could be classified as a rather complex molecule with a high molecular weight and several functional groups. The domain of application of K_{ow} calculation methods is characterised in terms of chemical structures. For example, some calculation programs cannot be applied to the estimation of Kow for phosphorus compounds including phosphites. Second, the validity domain of the models is also restricted by the log K_{ow} range of their applicability. In general, clear estimates can be expected in the region of log K_{ow} 0-5. Some programs have shown good estimates for compounds with log $K_{ow} > 5$ but estimates for log K_{ow} around 10 or above should be considered rather as qualitative than quantitative information (TGD, Part III, Chapter 4, E.C., 2003). As an example, Table 1.2 presents calculation results for log Kow of TNPP obtained using several models.

Table 1.2: log Kow calculations for TNPP along with the validity domain of the calculation method used

Program	Validity range*	TNPP value	Remark
CLOGP	0-5 (clear estimates in this range most of the time)	19.918	Very high LogP unrealistic in nature (this remark was associated to the result of the model) In general, CLOGP gives also better estimates with log Kow < 0.
LOGKOW (KOWWIN) Version 1.67	0-5 (clear estimates in this range most of the time)	20.05	
SPARC	>5	19.02	Better than KOWWIN and CLOGP for Kow >5.

* all estimates for Kow around 10 or above should be considered rather as qualitative than quantitative information.

Considering the high hydrophobic potential of TNPP which contains 27 aliphatic and 18 aromatic carbons, a high log Kow value could be expected for this compound. However, in the absence of valid data for this endpoint, a sensitivity analysis has been performed considering a range for log Kow between 6 and 20 (see Annex 1). This range takes into account both the highest result obtained using QSARs and the fact that this substance is expected to have a high log Kow based on its structure.

Consideration of further testing for the log Kow determination

OECD guideline n°117 gives guidance for Kow measurement using HPLC method (Kow is a function of the retention time in the HPLC column). Initially, this method is designed to measure log Kow between 0 and 6. But this method can be extended to measure log Kow between 6 and 10 using a modified mobile phase (or by shortening of the column length or decreasing the polarity of the eluent – more methanol or acetonitrile for example).

It has to be considered if a Kow for TNPP could be measured using the HPLC method with acetonitrile (aprotic solvent) as eluent for example. A measured Kow would be helpful for exposure assessment part. Indeed, if it can be shown that Kow is below 10 for example, the exposure concentrations for soil and sediment calculated with EUSES would be lower than if a worst case is taken for physico-chemical properties that are not known.

1.3.7 Water solubility

A water solubility was estimated using structure activity relationships models developed by the U.S. Environmental Protection Agency and Syracuse Research Corporation (EPIWIN, US EPA and Syracuse Research Corporation, 2001). The water solubility was estimated to 1.3×10^{-15} mg.L⁻¹ (Staples, 2001). Other estimations have been obtained using a more recent version of EPI suite software (US-EPA and Syracuse Research Corporation, 2004): 3×10^{-16} and 6.9×10^{-7} mg/L calculated with a water solubility estimate from log Kow (WSKOW v1.41 with a log Kow of 20.05) and a water solubility estimate from fragments, respectively.

Experimental water solubility was determined by (Reimer&Associates, 2001e). The flask method based on OECD Guideline 105 was used. TNPP was not detected in the saturated aqueous test solution. Therefore it is concluded that the water solubility of TNPP is below the detection limit of the substance. This detection limit was estimated to be 0.6 mg.L⁻¹, the lowest TNPP concentration that produced a signal that is reliably distinguished from the background signal as determined from chromatograms of TNPP solutions. Therefore, the water solubility of TNPP would be < 0.6 mg.L⁻¹ at 24°C.

The TNPP Industry commissioned a laboratory to develop a more sensitive analytical method for measuring TNPP so as to better approximate the true water solubility limit. Preliminary efforts were able to establish a new LOQ of 0.05 mg.L⁻¹. Solubility measurements have been attempted using this new analytical method. Considering the first results of this experiment, it seems that water solubility is still around or below this analytical limit (TNO, personal communication). This value has been used in the risk assessment but still need to be confirmed and validated.

Here again, in the absence of valid data for this endpoint, a sensitivity analysis will be made (see Annex 1) considering a range for water solubility between ~0.05 and 3×10^{-16} mg/L. This range takes into account both the highest result obtained using QSARs and the fact that this substance is expected to have a water solubility below the detection limit actually available for TNPP.

1.3.8 Flash point

Values of 183°C (internal reference, Great Lakes Chemical, Italia, Milan) and 195°C (Ciba MSDS) were reported using closed cup methods.

Besides, a value of 207°C was reported using the Pensky-Martin apparatus (closed cup) (Pittsburgh_Testing_Laboratory, 1978). This last value will be retained in this risk assessment because the analytical report was available.

1.3.9 Autoflammability

In a MSDS by Uniroyal, a value of 268°C was quoted. Moreover, using the Setchkin method, a result of 440°C was found (United States Testing Company, 1990).

1.3.10 Explosivity

No result could be found in the literature on any explosion limit. However, on the basis of its chemical structure, TNPP is not expected to have explosive properties.

1.3.11 Oxidising properties

No oxidising property was reported for TNPP (internal reference, Great Lakes Chemical, Milan, Italia).

1.3.12 Viscosity

In a product information sheet, a value of 6000 cps at 25°C is quoted (Crompton, 2003). Other values are also presented in this document showing that the viscosity goes from 15000 cps at 15°C to 18 cps at 120°C. The value at 25°C will be retained for the risk assessment.

1.3.13 Henry's Law constant

The Henry's law constant was estimated using structure activity relationships models developed by the U.S. Environmental Protection Agency and Syracuse Research Corporation (EPIWIN, v. 3.10, sub-model HENRYWIN, US EPA and Syracuse Research Corporation, 2001). At 25°C, a value of 66.1 Pa.m³.mol⁻¹ was calculated (US EPA and Syracuse Research Corporation, 2001).

The Henry's law constant can also be estimated from the ratio of the vapour pressure to the water solubility (E.C., 2003):

$$HENRY = \frac{VP \cdot MOLW}{SOL}$$

Using a vapour pressure of 0.058 Pa, a molecular weight of 689 g.mol⁻¹ and a water solubility of <0.05 mg.L⁻¹ the Henry's Law constant would be >799 Pa.m³.mol⁻¹.

In the risk assessment, the sensitivity analysis performed with log Kow and water solubilities will influence the value of the Henry's Law constant. This value will consequently range between 799 and 7.99x10¹⁶ Pa.m³.mol⁻¹ (the highest value obtained using the QSAR result for

the water solubility of 3×10^{-16} mg/L). Additional comments on the Henry's Law constant will be found in section 3.1.1.2.3 on environmental behaviour and fate.

DRAFT

1.3.14 Summary of physico-chemical properties

The physico-chemical properties of TNPP used in this risk assessment are summarised in the following table:

Table 1.3: Physical and chemical properties of the TNPP

Property	Value	Comments
Physical state at ntp	Viscous liquid	
Molecular weight	689 g.mol ⁻¹	
Melting Point	6°C ± 3°C	Instead of a melting point, a pour point (more appropriate to viscous liquids) was determined
Boiling Point	322°C	Degradation
Relative density	0.98 g.cm ⁻³	
Vapour pressure	0.058 Pa at 25°C*	extrapolated from results obtained by isoteniscope (method ASTM D2879)
Partition coefficient	Log Kow = 19* A sensitivity analysis will be performed in this risk assessment taking into account a log Kow ranging between 6 and 20	Calculated with SPARC
Water solubility	Upper value: <0.05 mg.L ⁻¹ * Lower value: 3.10 ⁻¹⁶ mg/L A sensitivity analysis will be performed in this risk assessment taking into account a water solubility ranging between 0.05 and 3.10 ⁻¹⁶ mg/L	Upper value: a saturated solution was not obtained and the water solubility result corresponds to the detection limit of the analytical method. Lower value: value obtained using QSAR calculation
Flash point	207°C	Pensky Martin apparatus (closed cup)
Autoflammability	440°C	Setchkin method
Explosivity	TNPP is not expected to have explosive properties	On the basis of chemical structure
Oxidising properties	No oxidising property	
Viscosity	6000 cps at 25°C	
Henry's law constant	Between 799 and 1.3.10 ¹⁷ Pa.m ³ .mol ⁻¹ (Cf. sensitivity analysis on water solubility)	TGD calculation

* Works are currently undertaken in order to better define this figure.

1.4 CLASSIFICATION

1.4.1 Current classification

TNPP chemical is not classified under Annex I of Directive 67/547 EEC.

1.4.2 Proposed classification

Waiting for additional information on the toxicity of TNPP.

DRAFT

2 GENERAL INFORMATION ON EXPOSURE

2.1 PRODUCTION

TNPP is produced all over the world: Unites States, Europe, India, Korea, Russia, China, etc. (Chemical Information Services, 2002). Three facilities are currently producing TNPP in Europe⁴. On the other hand, the major source of TNPP to Europe is from the United States.

2.1.1 Production process

The basic manufacturing processes used to produce TNPP are reasonably similar in the various plants in the US and Europe, except that not all plants strip out nonylphenol to the same degree. Figure 2-1 is providing an overview of a typical production process.

TNPP production is carried out in a closed system where nonylphenol (NP) and phosphorus trichloride (PCl_3) are added to the reactor (ca. 3 :1) and held at greater than 110°C to ensure all the PCl_3 is consumed. The HCl by-product is vented to an absorber. The HCL by-product can be filtered and stored for sale or use in other processes. Excess nonylphenol is stripped from the product. The stripped nonylphenol can be recycled. The product TNPP in the reactor after stripping is pumped to a storage tank for packaging and sale. The product may be packaged into drums, isotoners, rail cars, or tank trunks.

Environmental release and exposure

The process is fully automated (computer controlled) in a closed system. The reactor is operated under 3-5 lbs (1.4 – 2.3 kg) of pressure. The vacuum pump vent is the only potential process release to the atmosphere, and it is passed through a carbon filter. The storage tank is kept under nitrogen preventing release to the atmosphere. Nitrogen is also used during transfer and packaging.

⁴ In this report, Europe will correspond to 15 member states as data on exposure were gathered before the enlargement of the European Union in 2004.

Trisnonylphenyl Phosphite (TNPP) Process Overview

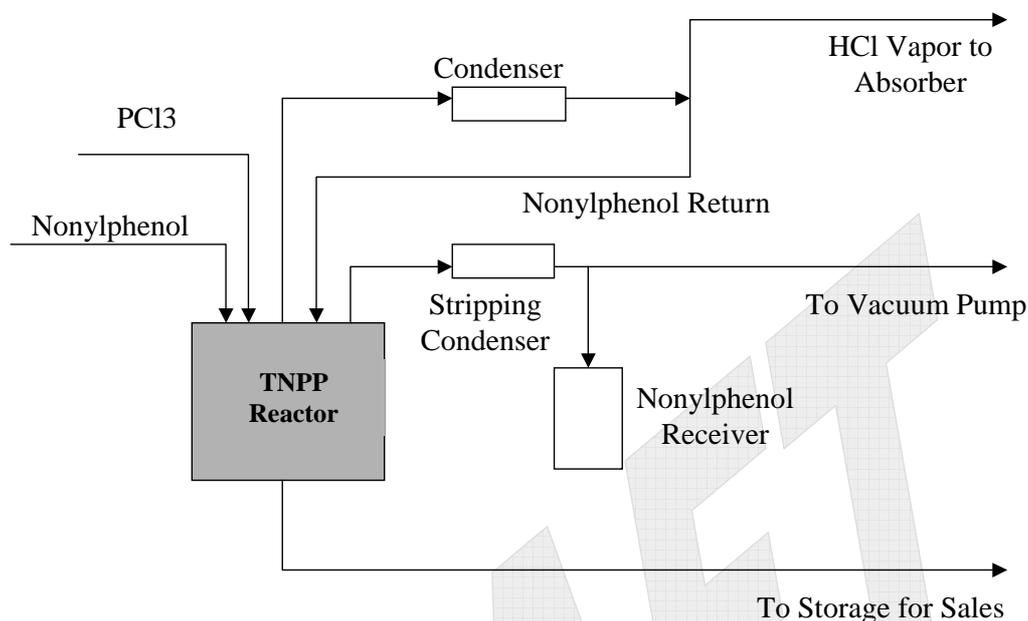


Figure 2-1: Process overview of tris(nonylphenyl)phosphite (TNPP) production

2.1.2 Production capacity

European and North American TNPP producers are organised under the TNPP consortium, a not-for-profit trade association, whose members have commercial interest in TNPP. Information on production and imports of TNPP in EU₁₅ were provided by the TNPP Consortium. Hardly any individual volume was provided for each producer/importer.

Three facilities are currently producing TNPP in EU₁₅. A fourth facility ceased TNPP production in 2001. Between 1990 and 1997, the production + import volumes were around 5,000 – 10,000 t/year.

Information is available on the combined estimate of TNPP produced within EU₁₅ and imported into EU₁₅ over the last three years:

- 1999 – approximately 5,565 tonnes
- 2000 – approximately 5,700 tonnes
- 2001 – approximately 6,800 tonnes

As this information is provided by the TNPP Consortium, it cannot be excluded that these volumes do not take into account shipments of product from producers in other parts of the world than Europe and North America. However, according to the TNPP Consortium, the quantity of TNPP from non-TNPP Consortium companies are not expected to be significant.

European production plants have also reported their production volumes for the year 2001. Imported volume for the same year is also available. Consequently, a total volume in EU₁₅ of 8,000 t. calculated with all 2001 data will be used in this report.

2.2 USES

2.2.1 Introduction

TNPP is used as a stabiliser in the processing of various plastic and rubber products. They are used with hindered phenolic antioxidants in plastic food packaging. In the stabilisation process, TNPP is gradually oxidised and nonylphenol is released (Building Research Establishment Ltd., 2001).

TNPP is also used as a secondary antioxidant in polymer formulations (Ullmann, 1985).

An estimate of the breakdown of TNPP uses was developed based on an informal survey of North American and European manufacturers. Quantitative breakdown of TNPP uses are given in Table 2.1. The information pertains to sales of TNPP in 1999. It is expected that the breakdown of uses from the 1999 sales statistics is typical for the current year. Corresponding volumes are calculated using the total tonnage of 8,000 t.

Table 2.1: Typical quantitative breakdown of TNPP Uses

	Percentage of tonnage	Volume (tonnes)	Industrial Category / Use Category
Polyvinylchloride (PVC) film	35%	2,800	IC 11 / UC 49
Polyolefins linear low density polyethylene (LLDPE)	15%	1,200	IC 11 / UC 49
High density polyethylene (HDPE)	10%	800	IC 11 / UC 49
Rubber	37%	2,960	IC 11 / UC 49
Other/Unknown	3%	240	IC 55 / UC 0
TOTAL	100%	8,000	

In the SPIN Database (SPIN, 2007), the following industrial uses are described:

Table 2.2: Industrial uses of TNPP in the Nordic Countries: Denmark, Norway and Sweden (in Tonnes)

	2000	2001	2002	2003	2004	2005
Manufacture of chemicals and chemical products	27	62	4	17	20	9.5
Manufacture of rubber and plastic products	105	88	471.1	466.1	62	40
Manufacture of furniture; manufacturing n.e.c.	0.4	0.1	0.3	0.1	0.1	0.1
Manufacture of fabricated metal products, except machinery and equipment	0.2	0.1	0	0	0	0
Construction	0.2	0.1	0.2	0.2	0.2	0.2
Manufacture of wood and products of wood and cork, except furniture; manufacture of articles of straw and plaiting materials	0	0.1	0			
Total (including all other uses)	133.5	578.7	479.2	483.6	82.4	49.9

TNPP is also mentioned in the following industrial categories: publishing, printing and reproduction of recorded media / sale, maintenance and repair of motor vehicles and motorcycles; retail sale of automotive fuel / manufacture of other transport equipment n.e.c. However, the volumes used in such industries and reported in this database could be considered as negligible (< 0.1 t/y in each country).

Besides, the following use pattern is described in the SPIN database:

Table 2.3: Use pattern of TNPP in the Nordic Countries: Denmark, Norway and Sweden (in Tonnes)

	2000	2001	2002
Process regulators	n. i.	n. i.	421.3
Stabilizers	120	91	46
Intermediates	1	n. i.	n. i.
Others	1	3	7
Adhesives, binding agents	0.5	1	1.7
Paints, lacquers and varnishes	0.3	0	0.1
Fillers	0	0.2	0.2
Total	122.8	95.2	476.3

n.i.: not indicated

TNPP is also mentioned in the following use categories: lubricants and additives / reprographic agents. However, the volumes used in such applications could be considered as negligible (< 0.1 t/y in each country).

From these tables, it could be stated that TNPP is mainly used as a stabiliser for the manufacture of rubbers and plastic products. The breakdown of TNPP uses described in Table 2.1 will be used in this risk assessment.

2.2.1.1 Industrial use

Formulation and processing steps are necessary to manufacture plastic and rubber products. Formulation could be defined as the stage where TNPP is combined in a process of blending and mixing into a polymer or into another material while during the processing step, the TNPP containing material is formed. It is not known to what extent formulation and processing may occur at the same site. In the rubber industry, these two steps can often not be viewed separately (E.C., 2003, Emission Scenario Document for IC 15: others: rubber industry).

Therefore, as a worst assumption, formulation and processing stages will be assumed to occur at one site for every use.

Without any specific information, it could be considered that TNPP is used for polymer processing, in the sub-category “processing of thermoplastics” as a processing aid. This categorisation will be used in the risk assessment for the determination of the default releases factors.

Besides, for plastic and rubber products, stages of private use and recovery may be considered. However, no specific information is available on the possible releases of TNPP during these stages.

All calculations will be performed using EUSES default parameters and, when available, emission factors issued from the emission scenario document on plastics additives (OECD, 2004).

2.2.1.2 Production of Polyvinylchloride (PVC) film

PVC containing TNPP may be used in many products like shower curtains, floorings and wall coverings.

2.2.1.3 Production of Polyolefins linear low density polyethylene (LLDPE)

LLDPE films containing TNPP are used for the manufacture of bags and food packaging. Many national regulations are covering the use of TNPP in food contact materials (Table 2.4)

Table 2.4: Global food contact regulations specific to TNPP

Country	Regulation
USA	Food and Drug Administration (FDA) – 21 CFR Part 178.2010
Japan	Self-restrictive Requirements on Food-Contact Articles Japan, Hygienic Olefin and Styrene Plastics Association (JHOSPA) (March 1996), Section A4-2, maximum 1.2%
European Union	Plastics Directive 2002/72/EC, pm/ref. No. 74400, specific migration limit 30 mg/kg
Germany	BfR Recommendation VI, maximum 2.0% total of all stabilisers BGA: maximum 6% in plastics
Netherlands	Food Packaging and Utensils Decree of 01.10.1979 as amended Chapter 1
France	Brochure 1227 (Avril 1990) maximum 1.0%
Italy	Min. Decree of 21.03.1973 maximum 0.3% Min. Decree of 0.04.1985
Spain	Royal Decree 125/1982 of 30.04.1982 Resolution of 4.11.1982
Belgium	Royal Decree of 11.05.1992, specific migration limit 30 mg/kg
United Kingdom	BIBRA/BBF Code of Practice (1991) Rec. No. C.159, maximum 1.0%

2.2.1.4 Production of High density polyethylene (HDPE)

HDPE containing TNPP is used in the manufacture of many products like blow-molded plastic drums or outer wrapping (film) of cigarette boxes or tea boxes.

2.2.1.5 Production of rubber

Rubber containing TNPP are used for example in tires and shoes soles.

2.2.2 Other applications

TNPP is used in other applications than plastic and rubber productions. Using the information provided in the SPIN database, it could be supposed that these other applications include the use of TNPP in publishing, printing and reproduction activities, in the manufacture of products of wood, of fabricated metal products, of furniture and in the construction activities. However, no more specific information is available.

2.2.2.1 Use of end-products

Shower curtains, flooring and wall coverings, bags and food packaging, blow-molded plastic drums, outer wrapping films, tires and shoes soles are examples of plastic and rubber end-products containing TNPP. For all these products, both private and professional end-uses may happen. As a worst case, private use of end-products made with plastics which might contain TNPP will be considered for all uses in the EUSES program (E.C., 2004b). However, it could be expected that TNPP or NP releases due to the use of end-products are low.

2.2.2.2 Recovery and disposal

No information on recovery has been submitted. In view of the end-products containing TNPP that are manufactured, it could be assumed that products containing TNPP may be either recycled into new products, disposed in landfill or incinerated. Therefore, this stage could be considered in the EUSES calculation (E.C., 2004a). However, no default value is currently available for this stage in version 2.0 of the software.

3 ENVIRONMENT RISK ASSESSMENT

3.1 ENVIRONMENTAL EXPOSURE

3.1.1 Environmental fate

3.1.1.1 Degradation in the environment

3.1.1.1.1 Atmospheric degradation

In the atmosphere, TNPP may be degraded by reactions with photochemically generated species like hydroxyl radicals. This atmospheric photo-oxidation potential can be estimated using structure activity relationships models developed by the U.S. Environmental Protection Agency and Syracuse Research Corporation (EPIWIN, v. 3.10, sub-model AOPWIN US EPA and Syracuse Research Corporation, 2001). The corresponding specific rate constant with OH radicals (k_{OH}) was estimated to be $50.6 \times 10^{-12} \text{ cm}^3 \cdot \text{molec}^{-1} \cdot \text{s}^{-1}$. Then the pseudo first order rate constant for degradation in air ($k_{deg_{air}}$) is determined with the following equation:

$$k_{deg_{air}} = k_{OH} \cdot OH \cdot CONC_{air} \cdot 24 \cdot 3,600$$

Using the global annual average OH radicals in the atmosphere ($5 \times 10^5 \text{ molec} \cdot \text{cm}^{-3}$), $k_{deg_{air}}$ is calculated to 2.19 d^{-1} and the half life for the reaction of hydroxyl radical with TNPP in the atmosphere is calculated as 0.32 days (7.6 hours).

Assuming another value for the OH radicals in the atmosphere ($1.5 \times 10^6 \text{ molec} \cdot \text{cm}^{-3}$) and 12 hours of daylight, $k_{deg_{air}}$ is calculated to 3.28 d^{-1} and the half life for the reaction of hydroxyl radical with TNPP in the atmosphere is calculated as 0.21 days (5.07 hours) (Staples, 2001).

The $1.5 \times 10^6 \text{ molec} \cdot \text{cm}^{-3}$ value for daylight hours is based on recent experimental observations (Leifer, 1993; Mount and Eisele, 1992 in US EPA and Syracuse Research Corporation, 2001).

The use of the 12 hours daylight period is justified by the U.S. EPA considering that OH radicals were existing only during sunlight hours. Therefore, the 12 hours period was chosen as an average daylight for a whole year and the pseudo first order rate constant was calculated with this new information. This last result (half life of 5.07 hours) will be used in the risk assessment.

Anyway, there are chances that TNPP will not be transported far from its emission source when it is emitted to the atmosphere. With such a low half life, TNPP will be rapidly degraded in the air and it is therefore not expected that TNPP will contribute to ozone depletion in the stratosphere.

3.1.1.1.2 Aquatic degradation - hydrolysis

TNPP can be hydrolyzed to form NP (Nonylphenol) in the presence of water. The ability to resist this decomposition is called “hydrolytic stability”.



Hydrolytic stability can occur in several different ways, depending upon where the actual hydrolysis is predicted to take place:

- Bulk storage and handling hydrolytic stability (during the production, shipment and storage of TNPP)
- In-polymer hydrolytic stability (what occurs to TNPP in the polymer during the processing)
- Hydrolytic stability of TNPP in an aqueous compartment (small amounts of TNPP in an aqueous solution)

The information available to quantify the hydrolytic stability of TNPP is presented hereafter in order to show thoroughly degradation properties of this substance. However, only results originating for the last test can be used in the risk assessment in order to estimate the behaviour and fate of TNPP in the environment (surface water).

Bulk storage and handling hydrolytic stability of TNPP

The bulk storage and handling hydrolytic stability of a phosphite can be measured by placing the neat phosphite in a Petri dish, and exposing it to elevated temperatures and humidity. This is an accelerated test. This accelerated test uses a temperature of 35°C (95°F) and 85% humidity. The stability was measured by analyzing for nonylphenol with HPLC, since nonylphenol is formed as the product hydrolyzes.

TNPP may contain a residual amount of nonylphenol that is left over from the synthesis/reaction. Dover Chemical offers two grades of TNPP, regular DP4 and a high purity grade called DP4HP that contains less than 0.1% free/residual nonylphenol.

Some hydrolysis rate constant calculations have been performed using a product datasheet from a producer (Dover Chemical Corporation, 2001). Degradation constants for three different TNPP formulated products have been determined based on the measurement of free nonylphenol formed from TNPP formulated products at 35°C and with 85% relative humidity (see Figure 3-1 **Error! Reference source not found.**).

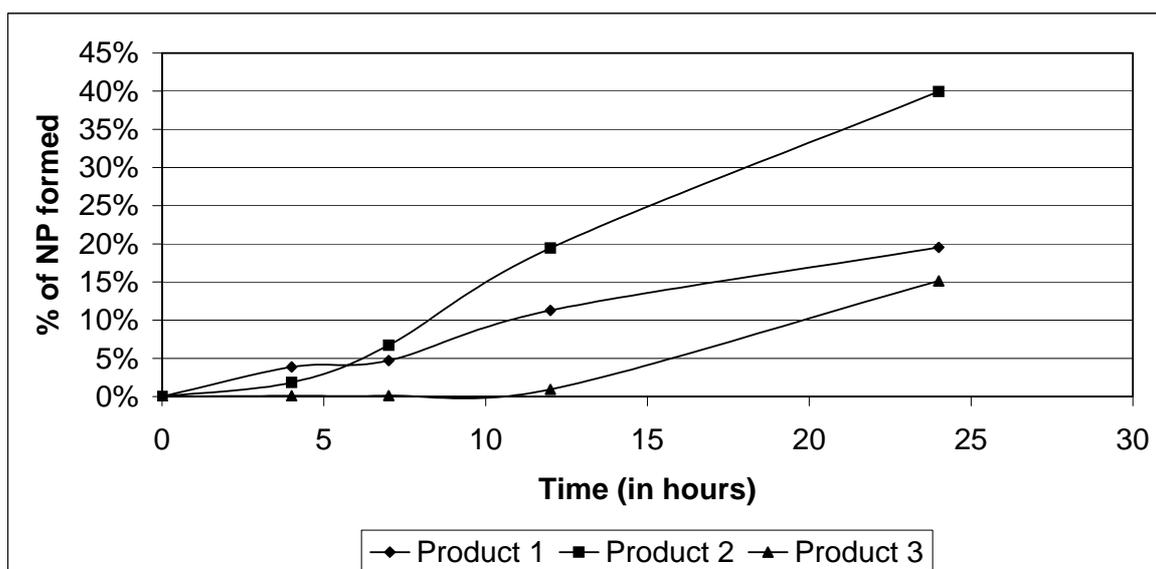


Figure 3-1: hydrolysis of different TNPP grades at 35°C and 85% relative humidity

Products 1 and 2 are “classical” TNPP formulation used in processing operations whereas product 3 contains an additive that enhances the stability of the product during its use. Calculated half-lives range between 1.6 and 4.4 days at 35°C (see Table 3.1 **Error! Reference source not found.**). This information could be used as supportive data in the risk assessment report particularly in order to assess to which extent NP could be formed during the use of TNPP for the processing of TNPP.

Table 3.1: hydrolysis half-lives of three TNPP grades

	Degradation constants at 35°C, 85% hum.	Half-lives at 35°C, 85% hum. (d.)
Product 1	0,0082	3,5
Product 2	0,0177	1,6
Product 3	0,0065	4,4

Other examples have been provided by Dover to illustrate the hydrolytic stability of TNPP during storage or handling and this is illustrated by Figure 3-2 (Dover, unpublished). TNPP is usually sold with an additive present at 0.5-1.0%. The additive, tri-isopropanol amine, or TIPA (CAS #122-20-3), acts as an acid scavenger and increases the hydrolytic stability of TNPP. ~78% of the TNPP Dover Chemical sells in Europe contains TIPA (DP4HR and DP4HPHR). Some applications such as PVC do not need TIPA since the polymer contains other types of acid scavengers. DP4HP with TIPA is very stable, with less than 3% hydrolyzed even after 80 days.

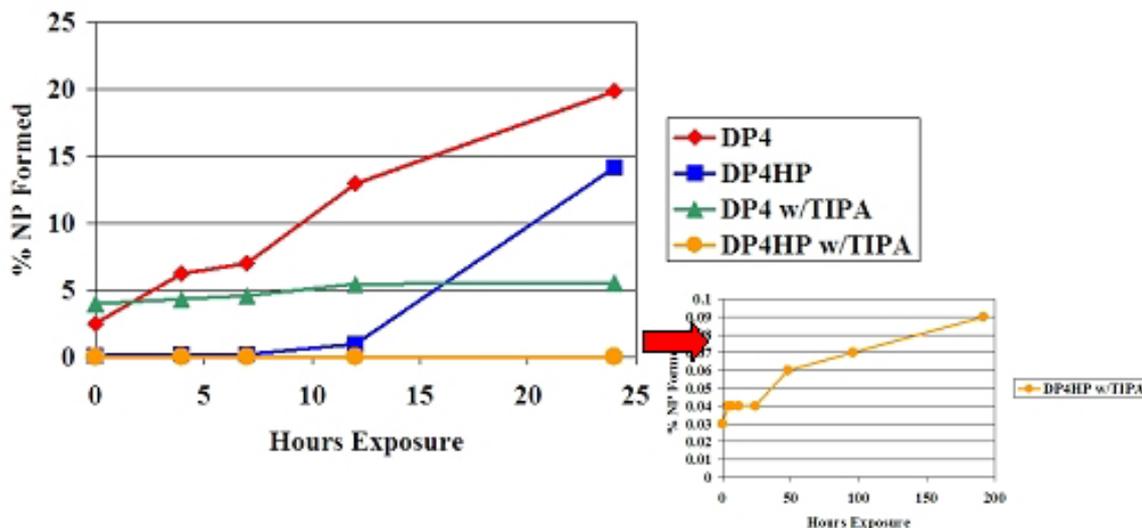


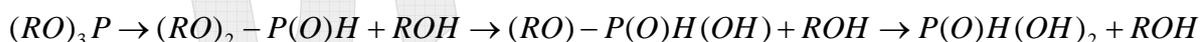
Figure 3-2: hydrolytic stability of TNPP (different purities and with/without TIPA) exposed at 35°C with 85% humidity

These test results can be used to support the fact that TNPP has a potential to hydrolyse during handling and storage. However this information cannot be used for the quantitative estimation of the hydrolysis potential of TNPP in the environment. Indeed, temperature conditions are different and the very low solubility of TNPP will influence its behaviour in the aquatic compartment.

Instead of measuring the hydrolytic stability of TNPP in the neat state, you can also measure the hydrolytic stability of TNPP when placed into an aqueous solution or environment (TNPP by itself is essentially insoluble in water).

Hydrolysis of TNPP in an aqueous compartment

In the literature, it is indicated that some organic phosphites are hydrolytically unstable with the general pathway described below (Goghova M. et al., 1989; Stevenson D.R., 1997 in Reimer&Associates, 2001b):



With R = organic group; alkyl or aryl.

For TNPP, the final hydrolysis products are nonylphenol (NP) and phosphorous acid. Theoretically, with excess of water, the hydrolysis should be complete, yielding 3 molecules of NP and 1 molecule of phosphorous acid for each molecule of TNPP (see Figure 3-3). However, laboratory data reveal that TNPP resists hydrolysis.

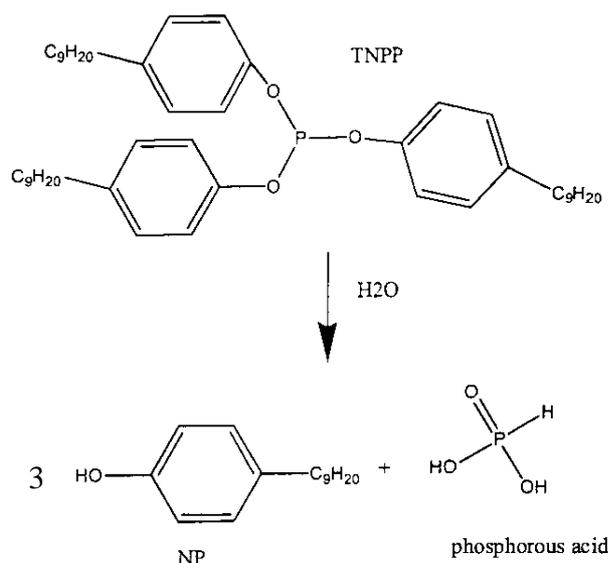


Figure 3-3: complete hydrolysis of TNPP

Any further breakdown of nonylphenol would be biodegradation.

① First hydrolysis study (Reimer&Associates, 2001b) – invalid study (included for information)

An experiment was conducted on TNPP to determine its hydrolysis potential as a function of pH (Reimer&Associates, 2001b): TNPP was dissolved in buffers (pH 4, 7, and 9) with CH₃CN co-solvent (1/1; v/v). These solutions were placed at 22°C and analysed repeatedly for NP as the hydrolysis product.

During the hydrolysis experiment, nonylphenol was detected but concentrations remained small and constant. This constant, low concentration of nonylphenol during hydrolysis was explained by the authors by the surfactant properties of NP which may have resulted in its adsorption to the glass surface of the vial (the hydroxyl group(-OH) of NP is responsible for hydrogen bonding with the glass). These adsorption properties of nonylphenol to the walls of vessels were also reported in some adsorption isotherm studies on nonylphenol (Roy F. Weston Inc, 1990d in Building Research Establishment Ltd., 2001). Due to its high K_{ow} and low water solubility TNPP probably also adsorbed to the glass surface of the vial and/or aggregated at the water surface. The measured decrease in TNPP could also be due to these phenomena. Another plausible explanation for the constant concentration of NP observed during the experiment could be the initial contamination of the test substance with NP. Indeed the test material in this study contained about 5% of nonylphenol.

For this hydrolysis experiment on TNPP, the apparent half-life of TNPP in solutions of buffer/CH₃CN (1/1; v/v) was estimated between 13 and 14 h at 22 °C. The hydrolysis was also found to be independent from pH conditions, i. e. TNPP concentrations decreased identically at pH 4, 7 and 9, which is different from what is usually observed with other alkyl phosphites (see Annex 2, OECD, unpublished). As described further, in subsequent experiments, these initial half-life determinations, were likely not accurately measuring the degradation of TNPP.

We should also consider that TNPP concentration used in this hydrolysis study (589 mg/L) is well above its water solubility (~0.5 µg/L based on the limit of quantification and a QSAR value of 3x10⁻¹⁶ mg/L has been calculated). The truly dissolved TNPP could hydrolyse faster but the resulting NP concentration would be well below the detection limit.

② *Second hydrolysis study (TNO, 2004)*

TNO has confirmed a very low limit of quantification for NP of 23 ng/L. The TNPP Industry therefore undertook a modified hydrolysis study to measure the rate of formation of NP from low concentration solutions of TNPP.

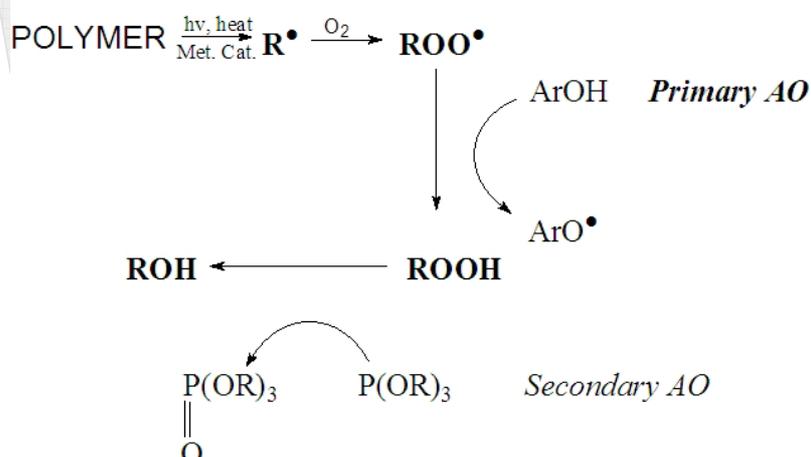
Since it was difficult to measure TNPP at low concentrations (see section 1.3.7 on water solubility), the TNPP Industry therefore undertook a modified 24-hour hydrolysis study where the rate of formation of NP was measured from water samples containing TNPP at 0.1 and 1 µg/L. During this test the level of NP was below the level of quantification (23 ng/L) in all the samples over the 24-hour period indicating no formation of NP.

Under the assumption that the half-life of TNPP was between 13-14 hrs, TNO should have been able to detect NP formed at the two (0.1 and 1 µg/L) concentrations used for the hydrolysis experiment. Also it has been established that NP is likely not adsorbing to the glass containers since TNO was able to find a very good recovery of NP in the calibration solutions prepared similar to the TNPP hydrolysis samples. Based on this experiment, it can be concluded that under ambient conditions TNPP in the aquatic environment will not hydrolyze to any meaningful degree to NP.

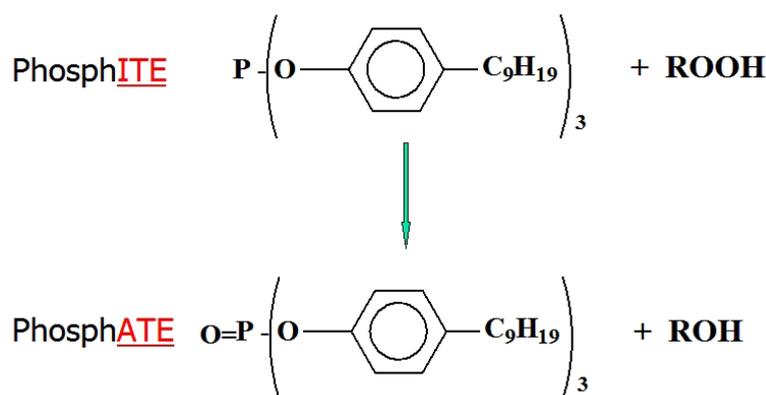
In conclusion, although it cannot be totally ruled out that there might be environmental conditions where hydrolysis could occur, hydrolysis of TNPP in the aquatic environment will not be considered as an important phenomenon. This is based on the expected very low water solubility of the substance that would not enable hydrolysis to occur in large amount. Furthermore, the high hydrophobicity of TNPP (high log Kow) will contribute to a large adsorption of the substance on sediment when entering the aquatic compartment thus reducing its availability for hydrolysis.

3.1.1.1.3 Aquatic degradation - oxidation

TNPP is used as a secondary antioxidant in polymers. It is placed into the polymer to decompose hydroperoxides, thus partially transforming TNPP into TNPP-phosphate while the polymer is being processed. In the polymer the following oxidation reaction occurs:



The following figure shows the chemical structures of TNPP and TNPP-phosphate:



Bulk storage and handling hydrolytic stability of TNPP-phosphate

During the processing of the polymers, TNPP is oxidised partly and TNPP-phosphate is produced. TNPP-phosphate is more hydrolytically stable than TNPP as illustrated by Figure 3-4:

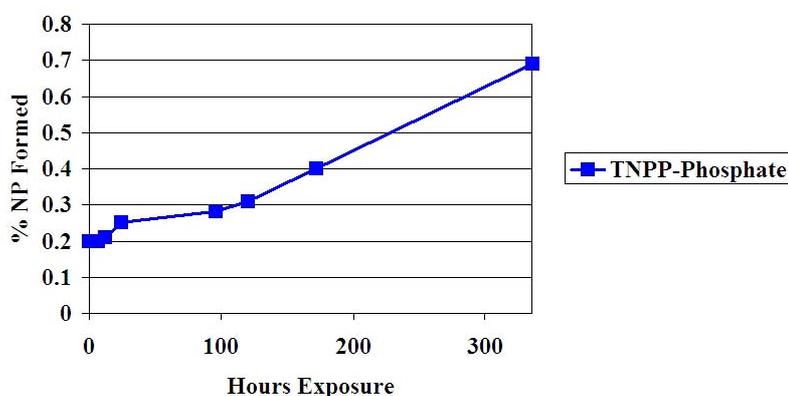


Figure 3-4: hydrolytic stability of TNPP-phosphate exposed at 35°C with 85% humidity

In an experiment (Dover, unpublished) with TNPP-phosphate, it has been shown that less than 1% of this substance is hydrolyzed after two weeks (without TIPA).

3.1.1.1.4 Aquatic degradation - biodegradation

Test #1: the ready biodegradability of TNPP was studied in the closed bottle test (OECD 301D) (Hydroqual Laboratories Ltd, 2001c). TNPP solutions were inoculated with a commercial bacterial preparation (Polysseed) and incubated at $20 \pm 1^\circ\text{C}$.

The biological oxygen demand (BOD) of the test substance was measured at 0, 7, 14, 21 and 28 days and compared to the theoretical oxygen demand (ThODNO₃) of the nominal concentration of TNPP. TNPP concentration was 15.4 mg.L⁻¹ which was theoretically corresponding to a Chemical Oxygen Demand of 13.2 mg. Beside the TNPP solution, there were 3 controls: a test control (inoculated mineral medium), a procedure control (degradation of a reference substance) and a toxicity control (degradation of the reference substance in the simultaneous presence of TNPP).

All controls passed the acceptability criteria of the test: oxygen depletion in the test control did not exceed 1.5 mg per litre after 28 days incubation, biodegradation of the sodium acetate

reference substance met the criterion of > 60% of the ThODNO₃ within 14 days. TNPP did not inhibit the degradation of the reference substance by more than 25% after 14 days. Finally, the variance amongst duplicate test bottles was less than 20%.

Less than 4% of TNPP was biodegraded after 28 days experiment. According to this test, TNPP is not readily biodegradable.

Test #2: the ready biodegradability of TNPP was studied in another test following the OECD guideline 301B (CIBA-Geigy, 1994). Test substance has been tested in duplicates at a concentration of 18.1 mg/L which corresponds to 15.3 mg ThOC/L. The inoculum was constituted with activated sludge collected from the sewage treatment plant of Reinach (Switzerland). The pH after collection was 7.0. Before application, the inoculum was pre acclimated to the test medium overnight. The test was performed at a temperature of 22 +/- 2°C with a carbon dioxide free air supply.

To take into account the very low solubility of the test substance, its preparation was as follow: a stock solution was prepared dissolving 1.36 g. of test substance in 10 mL dichloromethane. From this stock solution, for each replicate, 27.2 mg (200 µL) were applied onto a filter paper as small drops. After the filter paper was completely dry (no remaining of dichloromethane was present), it was cut to small pieces (10-15) and added to the test medium. Thereafter, the medium volume was completed to 1.5L with 300 mL water and the flasks were immediately connected to the CO₂ scrubber. Within a few hours the filter paper was homogeneously distributed in the test medium (so that it could not be seen anymore).

A deviation from the guideline should be noted. Indeed, only one CO₂ scrubber was used during the test. However, theoretically, a solution of 0.05 M NaOH is sufficient to trap at least twice more CO₂ than the maximum ThCO₂ which can be produced in each test bottle (including CO₂ possibly evolved from the bacteria, e.g. endogenous respiration). Moreover, experimentally it was confirmed that no measurable CO₂ carry over has ever occurred with the scrubbers used.

A test has also been conducted with sodium benzoate as reference substance, at a concentration corresponding to 15 mg DOC/L. After 7 days and 20 days, the biodegradation of the reference substance reaches respectively 71 and 86%.

Only 1% of TNPP was biodegraded after 29 days experiment. According to this test, TNPP is not readily biodegradable.

TNPP will be considered as non-readily biodegradable in this risk assessment.

3.1.1.1.5 Degradation in soil

No result could be found on degradation processes of TNPP in soils.

3.1.1.1.6 Summary of environmental degradation

TNPP released to the atmosphere is expected to degrade by reaction with hydroxyl radicals. A rate constant for degradation in air of 3.28 d⁻¹ with a corresponding half-life of 5.07 hours were estimated.

TNPP is not biodegradable in aquatic environments and the corresponding rate constant for biodegradation is 0 d⁻¹ with an infinite corresponding half-life.

As far as hydrolysis of TNPP in the aquatic environment is concerned, different sources indicate a potential of TNPP to hydrolysis. However TNPP hydrolysis will not be considered as an important degradation phenomenon in the risk assessment. This is based on the expected very low water solubility of the substance that would not enable hydrolysis to occur in large amount. Furthermore, the high hydrophobicity of TNPP (high log Kow) will contribute to a large adsorption of the substance on sediment when entering the aquatic compartment. However it should be taken into account that during the processing of polymers using TNPP as antioxidant, TNPP will undergo hydrolysis resulting in the release of nonylphenol in the environment. This hydrolysis during processing is due to the conditions of temperature in the process reactor.

The hydrolysis of TNPP leading to the formation of NP during processing will be considered in the risk assessment.

3.1.1.2 Distribution

3.1.1.2.1 Adsorption

The partition coefficients for TNPP have been calculated using EUSES (E.C., 2004a) based on log Kow of 6 and 20. They are presented as an example in Table 3.2. However, in the risk assessment, a range of values between 6 and 20 is used so as to assess the influence of the Kow on the assessment. Consequently, the use of each value retained for Kow will result in the calculation of appropriate partition coefficients for TNPP.

Table 3.2: Calculated partition coefficients for TNPP

	Log Kow 6	Log Kow 20	
K _{oc}	9.13x10 ⁴	2x10 ¹⁶	Partition coefficient organic carbon-water (L.kg ⁻¹)
K _p _{susp}	9.13x10 ³	2x10 ¹⁵	Partition coefficient solid-water in suspended matter (L.kg ⁻¹)
K _p _{sed}	4.56x10 ³	9.98x10 ¹⁴	Partition coefficient solid-water in sediment (L.kg ⁻¹)
K _p _{soil}	1.83x10 ³	3.99x10 ¹⁴	Partition coefficient solid-water in soil (L.kg ⁻¹)
K _{soil-water}	2.74x10 ³	6.1x10 ¹⁴	Soil-water partition coefficient (m ³ .m ⁻³)
K _{susp-water}	2.28x10 ³	4.99x10 ¹⁴	Suspended matter-water partition coefficient (m ³ .m ⁻³)
K _{sed-water}	2.28x10 ³	4.99x10 ¹⁴	Sediment-water partition coefficient (m ³ .m ⁻³)

3.1.1.2.2 Precipitation

Based upon the reaction with hydroxyl radicals, a half-life of 5.07 hours was calculated (see section 3.1.1.1.1). With such a short half-life, TNPP is unlikely to be transported a long distance from its point of emission and therefore neither enter the atmosphere in large amounts. Besides, TNPP being insoluble in water, concentrations in rainwater could be assumed to be negligible.

3.1.1.2.3 Volatilisation

A Henry's law constant between 799 and 1.33.10¹⁷ Pa.m³.mol⁻¹ was calculated from TGD (see section 1.3.13) taking into account the range of values chosen for water solubility. Indeed, in the risk assessment, the sensitivity analysis performed with log Kow and water solubility will influence the value of the Henry's Law constant.

The resulting air-water partition coefficient ($K_{air-water}$) would then range between 0.337 and $5.62 \times 10^{13} \text{ m}^3 \cdot \text{m}^{-3}$ by EUSES (E.C., 2004a). These values should be interpreted with care as it should also be taken into consideration that with the increase of hydrophobicity (lower solubility), higher adsorption of the substance on organic matter will occur. In that conditions, the property that will effectively limit the volatility of the substance will be the strong adsorption onto sediment and soil rather than the Henry's law constant.

Thus, volatilisation of TNPP from water is not expected to be a major phenomenon.

3.1.1.2.4 Distribution in waste water treatment plants

The distribution of TNPP in sewage treatment plants have been calculated using the model SIMPLETREAT integrated to EUSES (E.C., 2004a) based on different log Kow and Henry's law constants. They are presented as an example in Table 3.3. However, in the risk assessment, a range of values between 6 and 20 is used for Kow so as to assess the influence of the Kow on the assessment (see Annex 1). A range of values is also used for water solubility which impacts the calculation of the Henry's Law constant ($H > 799 \text{ Pa} \cdot \text{m}^{-3} \cdot \text{mol}^{-1}$).

Table 3.3: Estimation of removal of TNPP in STPs according to EUSES

	Log Kow 6 H = 799 Pa.m ⁻³ .mol ⁻¹ (calculated using a solubility of 0.05 µg/L)	Log Kow 20 H 1.33x10 ¹⁷ Pa.m ⁻³ .mol ⁻¹ (calculated using a solubility of 3x10 ⁻¹⁶ µg/L)
% to air	21.8	4.32x10 ⁻¹⁰
% to water	6.24	8
% to sludge	71.9	92
% degraded	0	0
% removal	93.7	92

TNPP being insoluble, not volatile and considered as not biodegradable, releases through production or processing will mainly go to sludge.

3.1.1.2.5 Accumulation and metabolism

Measured data on bioaccumulation of TNPP are not available. A calculated BCF has been obtained using EpiWin: 3.162 L/kg.

Bioaccumulation potential of TNPP could also be evaluated through EUSES calculation.

Based on the high Kow expected for TNPP, there are indications that TNPP may be bioaccumulated through trophic chains. Using EUSES calculation a bioconcentration factor of 4.62×10^4 L/kg could be calculated for fish taking into account a log Kow of 6.85 (worst case).

Besides, bioaccumulation of nonylphenol due to TNPP releases into the aquatic compartment will also have to be considered (BCF for NP: 1,280 L/kg for fish - E.C., 2002).

For earthworms, a partition coefficient earthworm-porewater is calculated using EUSES model (E.C., 2004a): $K_{\text{worm-porewater}} = 8.5 \times 10^4$ L/kg taking into account a log Kow of 6.85 (the same value as the one used for the calculation of the BCF for fish). Concerning the calculation of the BCF for earthworm, it should be noticed that contrary to QSARs available for fish, there is only one formula for the derivation of BCF for earthworm based on the log Kow. This leads to the determination of high BCFs for high Kow values (with a validity domain between 1 and 8) whereas for fish a maximum is calculated for the log BCF at a log Kow of 6.85.

The bioaccumulation factors calculated for TNPP based on a log Kow of 6.85 as a worst case indicate a high bioaccumulation potential. Nevertheless, the bioaccumulation potential of TNPP based on these calculations should be considered with precaution for the following reasons:

- molar weight is near 700 g/mol (689 g/mol) and certain classes of substances with molecular mass greater than this threshold are not readily taken up by fish and are unlikely to bioaccumulate significantly.
- Information on the molecular size of TNPP is also available (personal communication, Kazumi Kawahara, CERi, 20th October 2005). Based on this study, it seems that, taking into account the calculated molecular size of TNPP, the bioaccumulation potential is negligible. The calculation of the mean diameter for six different three dimension structures of TNPP has led to a lowest value of 13.9 angstrom. This conclusion has been reached based on a cut-off value for the ability of a chemical to pass through fish gill membrane has been established at 9.5 angstrom (Opperhuizen *et al.*, 1985). However, it should also be considered that the current cut-off value proposed by the PBT subgroup is a mean diameter higher than 17 angstroms.
- A worst case value of 7 has been taken into account for the calculation of BCFs for TNPP. However, there are some indications that the Kow of TNPP could be much higher than this value (a QSAR calculation gives a log Kow of 19 for example).

3.1.2 Environmental releases

3.1.2.1 General information

Releases of TNPP and/or NP (nonylphenol) to the environment occur during production, transport, storage, formulation and processing of plastic and rubber products. In addition,

releases may also take place through the uses of the end-products. Finally, waste disposal of the end-products may also release TNPP or NP into the environment.

The different industry categories (IC), use categories (UC) and main categories (MC) used in the EUSES calculations are described in Table 3.4.

Table 3.4: Industrial Categories (IC), Use Categories (UC) and Main categories (MC) used in EUSES calculations

Life cycle stages		IC	UC	MC
Production		11	49	I b
PVC films (2,800 t)	Formulation	11	49	III
	Processing	11	49	II
LLDPE films (1,200 t)	Formulation	11	49	III
	Processing	11	49	II
HDPE films (800 t)	Formulation	11	49	III
	Processing	11	49	II
Rubber (2,960 t)	Formulation	11	49	III
	Processing	11	49	II
Others (240 t)	Formulation	15	55	III
	Processing	15	55	II

About 25 to 35 facilities are processing TNPP in EU₁₅. Their consumption ranges from a few tonnes to around 700 tonnes/year. The highest value comes from an identified site where near 10% of the total TNPP used in EU₁₅ is processed. As the type of use taking place at this site is not known, this worst case will be taken into account for each category of use identified. In the exposure assessment, it will also be considered by default that only one site processes TNPP in a region with an annual tonnage of 700 t.

Table 2.1 shows the order of magnitude of the sizes of European sites where TNPP is used. These data come from a survey where 21 sites were identified using a total of 4,269 t. of TNPP.

Table 3.5: order of magnitude of TNPP volumes processed in identified sites

Use	Number of sites identified in the specified volume range		
	< 45 t. ("small" site)	45-227 t. ("medium" site)	>227 t. ("large" site)
Polyolefins linear low density polyethylene (LLDPE)		1	1
High density polyethylene (HDPE)		3	
Rubber	2	6	1
Polyvinylchloride (PVC) film		1	3
Other	1	1	1

The regional tonnage will be defined in the following way:

- Where a “large” site has been identified for a type of use, the regional volume will be set at 700 t. (the highest site identified), except for the other uses to which only 240 t. have been attributed. It will be considered that there is only one site in a region.
- When only medium sites have been identified, e. g. for TNPP used for HDPE, only one site will be considered in a region with a volume of 227 t.

Consequently, for tonnage input in the B tables, regional tonnage of TNPP was set to 700 t for the uses for PVC, LLDPE and rubber (maximum reported consumption range for TNPP processing facilities). For the uses in HDPE and other uses, the regional tonnage was respectively set to 227 t and 240 t.

A default fraction of TNPP in formulation is suggested in TGD (E.C., 2003) Emission Scenario Document for rubber Industry: up to 1.5 % (wt) for processing aids used as stabilisers. However, TNPP manufacturers have submitted better approximations of this value, for different formulated products (Personal communication from TNPP consortium, 1st April 2004):

- | | |
|---------------|-----------|
| ▪ PVC film | 0.8-1.5 % |
| ▪ Polyolefins | 0.1-0.2 % |
| ▪ Rubber | 0.4-1.0 % |

As a worst case, the upper limit of these intervals will be used for the exposure assessment. Then, in the absence of more specific information, fractions of the main source and number of days are derived from Tables B using the tonnage as such for each use.

Releases of TNPP have to be estimated during the production of the substance and during its uses. TNPP has several applications in Polymer Industry (Industrial Category 11) where it is mainly used as a stabiliser (Use Category 49). Both formulation and processing steps have to be considered in the risk assessment.

Releases due to the use of TNPP containing products have also to be considered.

Local releases due to formulation and processing of TNPP are calculated using default scenarios presented in the TGD (E.C., 2003) and other available Emission Scenario Documents. Details of calculation parameters are given in Table 3.7.

3.1.2.2 Use of the OECD ESD for plastic additives with TNPP

TNPP is used as an antioxidant and volatility of antioxidants is the same as the substances used in plasticizers. Considering its vapour pressure, TNPP should be classified in the high volatility group (NOTE: By mid-September, Industry is expected to have available an updated vapour pressure determination for high purity TNPP. If the updated value results in a change to the volatility group, revised emissions estimates will be prepared. Depending on the vapour pressure of TNPP, emission factors would be lowered by a factor of 5 or 25). Emission factors are detailed hereafter. Worst case emission factors available in the ESD for plastic additives have been used (OECD, 2004).

- Raw material's handling (formulation)

The initial state of TNPP is liquid. To estimate TNPP releases, antioxidants are analogous to plasticizers with respect to handling. Thus, default emission factors of the scenario for antioxidants cannot be applied in its case. The scenario only considered losses from the handling

of solid materials. Plasticizers are almost in a liquid form. They are usually transported and handled in bulk through enclosed storage systems. So minimal loss by spillage can be assumed.

Consequently, $F_{\text{handling,water}} = 0.01\%$. It is considered that loss by volatilisation should be minimal.

- Compounding (formulation)

Two cases should be taken into account depending on the method used for the compounding. Here again, the factors for plasticizers are used. Release factors used for dry blending and Banbury blending are used as a worst case: $F_{\text{compounding,water}} = 0.025\%$ and $F_{\text{compounding,air}} = 0.025\%$ (emission factors for the high volatility group).

For the formulation stage, the following emission factors will be used: $F_{\text{air}} = 0.00025$ and $F_{\text{water}} = 0.00035$

- Conversion (processing)

For high volatility group, losses during conversion are estimated using the following emission factors. As a worst case, the processing performed in open processes (solid articles) will be applied: $F_{\text{conversion,water}} = 0.125\%$ and $F_{\text{conversion,air}} = 0.125\%$.

- Service life

The following emission factors have been chosen for the service life of products containing TNPP:

- Indoor service, leaching to liquid waste: $F_{\text{service life,water}} = 0.05\%$ over lifetime
- Indoor service, volatility to atmosphere: $F_{\text{service life,air}} = 0.05\%$ over lifetime
- Outdoor service, leaching to environment: $F_{\text{service life,water}} = 0.16\% \times T_{\text{service}}$ (T_{service} is the service life of product in years)
- Outdoor service, volatility to atmosphere: $F_{\text{service life,air}} = 0.05\%$ over lifetime

This results in fractions of emission to water of 0.016 (taking into account, as a worst case, a lifetime of 10 years for TNPP containing products) and 0.0005 to air (the worst case of outdoor service is taken into account).

- Disposal

In landfill, leaching losses to water will depend on many factors, relating to the type of landfill as well as to the properties of TNPP and the nature of the polymer in which it has been used. The maximum potential loss could be calculated from the amount of additive remaining in the plastic at disposal, but it is very unlikely that this amount would be released. The volatilisation loss from landfill is also likely to be limited. Consequently, in this assessment, releases during disposal are assumed to be negligible.

3.1.2.3 Use of the OECD ESD for additives used in rubber industry with TNPP

For this scenario, the production of two main product types identified in this assessment will be considered: tyres and soles.

- Formulation and processing

The emission rate into waste water for formulation and processing is calculated using the following equation:

Equation 3-1: calculation of daily releases into wastewater during formulation and processing of TNPP used in rubber industry

$$E_{local\ water} = Q_{prod} \cdot \frac{Q_{additive}}{100 \cdot F_{recipe}} \cdot (1 - F_{remaining})$$

For TNPP, the amounts of the product types produced per day (Q_{prod}) are taken by default as 26,400 kg/d for tyres and 550 kg/d for soles. The part of additive introduced into the rubber is 0.25 phr (parts per hundred rubber parts) for tyres (default value) and 3 phr in soles (default for other rubber products). The fraction of TNPP remaining in the rubber product ($F_{remaining}$) is set at 0.943 (the default value, 0.99, minus the fraction of NP formed during processing) for tyres and 0.933 for soles (the default value, 0.98, minus the fraction of NP formed during processing). Finally, F_{recipe} is set at 2 when $Q_{additive}$ is given in phr.

For the releases in air and soil, the calculations are made as follow:

Equation 3-2: calculation of daily releases to air and soil during formulation and processing of TNPP used in rubber industry

$$E_{local\ air} = Q_{prod} \cdot \frac{Q_{additive}}{100 \cdot F_{recipe}} \cdot F_{air} \quad \text{or } F_{soil}$$

With F_{air} and F_{soil} set at 5×10^{-4} and 1×10^{-4} respectively.

- Service life: for this stage of the life cycle, only the emission resulting from the abrasion of tyres will be considered. The emission calculated using the equation proposed in the Emission Scenario Document for Rubber Industry will be allocated to the region.

Equation 3-3: calculation of the annual releases to water at the regional level due to the abrasion of tyres containing TNPP

$$E_{region\ water} = Q_{ann_prod} \cdot F_{abrasion} \cdot \frac{MM_{abrasion}}{MM_{additive}}$$

With Q_{ann_prod} the amount of additive used per year in the region (by default, the tenth of the amount of TNPP used in EU₁₅ in rubber products will be used, 296 t/a). $F_{abrasion}$ the emission factor for abrasion from tyres (0.12). The ratio of molecular weights will be set at 1 since here the additive is considered (TNPP).

- Disposal: this stage is not taken into account due to a lack of data to estimate the emission rates and leachate processes.

3.1.2.4 Calculation of NP releases due to the processing of TNPP

During the use of TNPP, nonylphenol is present as an impurity and could, to some extent and under the conditions of temperature and humidity occurring in the process, be formed due to the

hydrolysis of TNPP. It should be noted that NP formation during the processing of TNPP is limited as much as possible, using very pure TNPP or employing stabilisers in TNPP formulation for example, because it has a negative impact on the production of plastics. TNPP is used as a stabiliser in the processing of various plastic and rubber products. The primary mechanism of polymer stabilisation of phosphites is oxidation, not hydrolysis. It is extremely critical during the process to minimise the generation of NP from TNPP, as this will destroy the stabiliser properties.

Information that can be used to quantify the amount of NP formed during the manufacture of plastics can be estimated from a study by Howe *et al.*, 2001. This assessment was performed using two grades of TNPP differing with respect to the level of residual NP. These grades have been chosen to be representative for the TNPP available on the market. The concentrations of residual NP were ~1.5% (sample 1) and <0.1% (sample 2). Using these samples of TNPP grades resulting in the measurement of the following concentrations in plastics:

Plastic sample	p-nonylphenol		TNPP		TNPP phosphate	
	ppm	% of total TNPP _{total}	ppm	% of total TNPP _{total}	ppm	% of total TNPP _{total}
LLDPE Resin 1	58	4.7	819	66.2	360	29.1
LLDPE Film 1	10	0.9	631	54.1	526	45.1
LLDPE Resin 2	14	1.2	847	69.9	350	28.9
LLDPE Film 2	5	0.4	891	70.4	370	29.2
PVC film 1	160	2.4	3170	47.4	3360	50.2
PVC film 2	120	1.7	3390	47.3	3650	60.0

It has to be noticed that this source of information is considered relevant to estimate the fraction of NP in plastics. Indeed, the initial residual NP concentrations in TNPP samples were known and these TNPP grades have been taken as representative for the TNPP on the market.

Taking the maximum value for NP concentration, it will be assumed that during the process, 4.7%⁵ of TNPP is hydrolysed leading to a similar amount of NP (in mass). Indeed, considering the hydrolysis pathway (three molecules of NP formed for each molecule of TNPP hydrolysed) and the molecular weights of the substances, it can be assumed that 1 mg of TNPP would yield to 0.96 mg of NP.

The following properties have been taken for the risk assessment of NP due to the use of TNPP. They have been taken from the EU RAR for nonylphenol (E.C., 2002).

Table 3.6: main characteristics of nonylphenol used for the risk assessment

Molecular weight	220,34 g/mol	F _{STP-air}	0.0669
Melting point	-8°C	F _{STP-water}	0.35
Boiling point	290°C (degradation)	F _{STP-sludge}	0.344
Relative density	0.95	K _{psusp}	5.36.10 ²
Vapour pressure	0.3 Pa	K _{psed}	2.68.10 ²
Log Kow	4.48	K _{psoil}	1.07.10 ²

⁵ This fraction of TNPP converted into NP during the process is based on limited information on the fraction of NP in product samples taken during processing. Further information is required concerning this issue.

Water solubility	6 mg/L	K _{oc}	5.36.10 ³
Henry's Law constant	11.02 Pa.m ⁻³ .mol ⁻¹	BCF	1,280
Half-life for biodegradation in soil	300 days		

3.1.2.5 Sensitivity analysis of the exposure assessment for ranges of log Kow and water solubility (and subsequent associated parameters)

A sensitivity analysis has been performed for the exposure assessment due to the lack of data on log Kow and water solubility. Several calculations were run taking into account the following ranges for the parameters tested:

- between 6 and 20 for log Kow
- between 0.05 and 3.10⁻¹⁶ mg/L for water solubility

Details on the sensitivity analysis are available in annex 1. Minimum and maximum values obtained for exposure concentrations are reported all along the exposure assessment up to the risk characterisation.

Table 3.7: Parameters used for the calculation of local releases due to formulation and processing of TNPP

Life cycle stages	Tonnage				Stage	Emission factors				Local characteristics			
	Uses	Fraction	Tonnage for application	Regional tonnage of substance		Fraction of TNPP in formulation	A-Table or scenario	F released to air	F released to waste water	F released to industrial soils	B-Tables	Number of days of emission	Fraction of main source
PVC films		0.35	2,800	700	0.015	Formulation	PA ESD ²	0.00025	0.00035	0	B 2.3	300	1
						Processing	PA ESD ²	0.00125	0.00125	0	B 3.9	300	1
						Service life ¹	PA ESD ²	0.0005	0.016	0	B 4.1	365	0.002
LLDPE films		0.15	1,200	700	0.002	Formulation	PA ESD ²	0.00025	0.00035	0	B 2.3	300	1
						Processing	PA ESD ²	0.00125	0.00125	0	B 3.9	300	1
						Service life ¹	PA ESD ²	0.0005	0.016	0	B 4.1	365	0.002
HDPE films		0.1	800	227	0.002	Formulation	PA ESD ²	0.00025	0.00035	0	B 2.3	300	1
						Processing	PA ESD ²	0.00125	0.00125	0	B 3.9	300	1
						Service life ¹	PA ESD ²	0.0005	0.016	0	B 4.1	365	0.002
Rubber		0.37	2,960	700	0.01	Formulation	RI ESD ³	Specific calculations for this scenario are explained in section 3.1.2.3					
						Processing	RI ESD ³						
						Service life ¹	RI ESD ³						
Others		0.03	240	240	0.015	Formulation	PA ESD ²	0.00025	0.00035	0	B 2.3	300	1
						Processing	PA ESD ²	0.00125	0.00125	0	B 3.14	300	1

¹ Emission considered at the regional level

² OECD Emission Scenario Document on plastic additives

³ OECD Emission Scenario Document on additives in rubber industry

3.1.2.6 Regional releases

Based on the exposure scenarios introduced previously, the following regional releases can be estimated:

Table 3.8: regional releases of TNPP

	TNPP regional releases (kg/d)
Air	15.2
Waste water	111
Surface water	125
Industrial soil	0.688

3.1.2.7 Regional concentrations

The following regional concentrations for TNPP have been calculated using EUSES and taking into account the different releases identified in the previous section of the exposure assessment.

The regional concentration of NP are also reported in .

Table 3.9: regional concentrations for TNPP

	TNPP regional PEC Log Kow 6; sol. 50 µg/L	TNPP regional PEC QSARs values
Surface water (µg/L)	4.95×10^{-2}	1.87×10^{-12}
Sea water (µg/L)	1.77×10^{-3}	1.83×10^{-13}
Freshwater sediment (µg/kg ww)	195	1630
Seawater sediment (µg/kg ww)	6.89	159
Agricultural soil (mg/kg ww)	0.75	0.078
Natural soil (mg/kg ww)	0.308	0.023
Air (mg/m ³)	6.34×10^{-7}	6.17×10^{-7}

Table 3.10: regional concentrations for NP (E.C., 2002)

	NP regional PEC
Surface water (µg/L)	0.60
Freshwater sediment (µg/kg ww)	103
Agricultural soil (mg/kg ww)	0.265
Natural soil (mg/kg ww)	1.44×10^{-5}
Air (mg/m ³)	3.14×10^{-6}

Since the completion of the risk assessment report, risk reduction measures have been applied to reduce the amounts of NP released in the environment. Consequently, this should have had an impact on the regional concentrations calculated initially. The contribution of these regional

concentrations to the risk calculated in this risk assessment will be taken into account when conclusions will be drawn.

3.1.2.8 Releases during TNPP production

Three facilities are currently producing TNPP in EU₁₅. A fourth facility ceased TNPP production in 2001. In general, companies do not routinely monitor TNPP or NP and hence meaningful monitoring data from most facilities are likely not available. To estimate the daily releases of TNPP, TGD Tables A1.1. and B1.4. were used (E.C., 2003).

Site specific information was used to complete the exposure assessment of the three facilities manufacturing TNPP in EU₁₅. To ensure the confidentiality of the data, detailed calculations for real production sites are not included at this stage. PEC calculated using site specific information will appear at point 3.1.3.1.1.

One production facility conducted periodic monitoring for nonylphenol of the waste stream leaving their waste treatment facility prior to entering the municipal treatment plant. In 2002, the NP concentrations were non-detectable, with a sensitivity of 1 mg/m³. In 2003, the results were also non-detectable with a sensitivity of 0.1 mg/m³. Consequently, at the production step, releases of NP from the production of TNPP will not be taken into account.

3.1.2.9 Releases during the use in PVC films or LLDPE films

Remark: both scenarios are compiled since there are driven by the same parameters.

Table 3.11: Local TNPP and NP (in brackets) emissions due to use in PVC films or LLDPE films

	Emission to waste water	Emission to Air
	kg/d	kg/d
Formulation	0.82 (0.0384)	0.58 (0.0274)
Processing	2.91 (0.137)	2.91 (0.137)
Service life	0.06 (2.97x10 ⁻³)	0.002 (1.8x10 ⁻⁴)
Disposal / Recovery	Not considered	

3.1.2.10 Releases during the use in rubber

Table 3.12: Local TNPP and NP (in brackets) emissions due to use in rubber

	Emission to waste water	Emission to Air
	kg/d	kg/d
Formulation and processing		
- tyres	- 1.881 (0.088)	- 0.017 (8x10 ⁻⁴)
- soles	- 0.553 (0.026)	- 4x10 ⁻³ (2x10 ⁻⁴)
Service life		
- tyres	Considered at the regional scale	
Disposal / Recovery	Not considered	

3.1.2.11 Releases during the use in HDPE films

Table 3.13: Local TNPP and NP (in brackets) emissions due to use in HDPE films

	Emission to waste water	Emission to Air
	kg/d	kg/d
Formulation	0.27 (0.0124)	0.19 (8.89x10 ⁻³)
Processing	0.95 (0.044)	0.95 (0.044)
Service life	0.02 (9.62x10 ⁻⁴)	0.001 (5.83x10 ⁻⁵)
Disposal / Recovery	Not considered	

3.1.2.12 Releases during the use in other applications

Table 3.14: Local TNPP and NP (in brackets) emissions due to use in other applications

	Emission to waste water	Emission to Air
	kg/d	kg/d
Formulation	0.28 (1.32x10 ⁻²)	0.20 (9.4x10 ⁻³)
Processing	1.00 (4.7x10 ⁻²)	1.00 (4.7x10 ⁻²)

3.1.3 Aquatic compartment (incl. sediment)

3.1.3.1 Predicted environmental concentrations in water

The Predicted Environmental Concentrations (PECs) for local water are calculated using the environmental releases detailed in section 3.1.2 using the equations set out in the TGD. By default and in the absence of specific information, the following values are used: $EFFLUENT_{STP} = 2000 \text{ m}^3/\text{d}$ (flow of the STP), $DILUTION = 10$ (dilution factor).

In calculating the local PEC, the regional PEC is added to the local concentrations.

3.1.3.1.1 $PEC_{\text{local(water)}}$ at TNPP production sites

Final results of the PECs calculations for the three European TNPP production sites are presented in Table 3.15. The calculations have been performed using specific information available (e.g. flows of the STPs and receiving waters). Although these specific data are kept confidential, the type of data available for each site is indicated in Table 3.15.

 Table 3.15: PEC_{local} for the three European TNPP production sites

	Site-specific information available	PEC_{STP} for TNPP (µg/L)	PEC_{local} for TNPP (µg/L)	PEC_{STP} for TNPP (µg/L)	PEC_{local} for TNPP (µg/L)
		Log Kow 6; sol. 50 µg/L		QSAR values	
Site A	Production volume (2001) Waste water treatment plant flow Flow of receiving waters Releases of TNPP	64.4	0.106	84.1	4.68x10 ⁻¹²

	Site-specific information available	PEC _{STP} for TNPP (µg/L)	PEC _{local} for TNPP (µg/L)	PEC _{STP} for TNPP (µg/L)	PEC _{local} for TNPP (µg/L)
	Processing takes place on-site (included)				
Site B	Production volume (2001) Waste water treatment plant flow Flow of receiving waters	104	3.7	136	1.83x10 ⁻¹⁰
Site C	Production volume (2001)	655	57.7	857	2.86x10 ⁻⁹

3.1.3.1.2 PEC_{local(water)} at sites using TNPP

Site specific information for releases during the use of TNPP for LLDPE films production

In 2003, water samples were collected at one TNPP processing facility. These samples were analysed for nonylphenol concentrations by High Performance Liquid Chromatography. The highest concentration of 82.7 µg/L was measured in the process wastewater.

At the same site, nonylphenol concentrations up to 10.3 µg/L were measured in sewer. At this site, the stream further undergoes primary treatment prior to discharge but it is not subjected to secondary treatment. Therefore, as a worst case estimation (no further removal), nonylphenol concentration in effluent of 10.3 µg/L could be considered at this site as a concentration resulting from TNPP processing.

This monitoring result shows that nonylphenol releases due to TNPP uses in polymer industry could not be neglected. However, as on one hand the end-products processed at this site are not known and on the other hand the TNPP volumes used are not specified, these values could not be used as a general scenario for estimating nonylphenol releases due to TNPP processing.

Therefore, default releases estimations will have to be used in the risk assessment for each use pattern.

Default releases estimation

It is assumed that formulation and processing stages could occur at the same site. Therefore, in calculating local concentrations, added releases due to both stages are considered.

Table 3.16: PEC_{local} for water for sites using TNPP

Uses	PEC _{local} (µg/L)			PEC _{STP} (µg/L)		
	TNPP Log Kow 6; sol. 50 µg/L	TNPP QSAR values	NP	TNPP Log Kow 6; sol. 50 µg/L	TNPP QSAR values	NP
PVC films LLDPE films	10.1	5x10 ⁻¹⁰	3.67	114	150	30.9
Rubber						
- tyres	5.11	2.53x10 ⁻¹⁰	0.871	57.6	75.2	2.73
- soles	1.54	7.57x10 ⁻¹¹	0.736	16.9	22.1	1.37
HDPE films	3.31	1.63x10 ⁻¹⁰	1.59	37	48.4	10.0
Other uses	3.49	1.73x10 ⁻¹⁰	1.65	39	51.2	10.6

3.1.3.1.3 **PEC_{local(water)} for emissions during products service life**Table 3.17: PEC_{local} for emission during products service life

Uses	PEC _{local} (µg/L)			PEC _{STP} (µg/L)		
	TNPP Log Kow 6; sol. 50 µg/L	TNPP QSAR values	NP	TNPP Log Kow 6; sol. 50 µg/L	TNPP QSAR values	NP
PVC films	0.214	1x10 ⁻¹¹	0.65	1.87	2.45	0.52
LLDPE films						
HDPE films	0.103	4.52x10 ⁻¹²	0.62	0.61	0.79	0.17

3.1.3.2 **Calculation of Predicted Environmental Concentration for Sewage Treatment Plants (PEC_{stp})**

For the risk characterisation of a substance upon micro-organisms in the STP, it can be assumed that homogeneous mixing in the aeration tank occurs which implies that the dissolved concentration of a substance is equal to the effluent concentration. In addition, no intermittent release is assumed for production or uses of TNPP. Therefore, PEC_{STP} are equal to the concentration of TNPP in STP effluents (C_{local,eff}). See above section 3.1.3.1 for the results.

3.1.3.3 Calculation of PEC_{sediment}

The concentration in bulk sediment can be derived from the corresponding water body concentration, assuming a thermodynamic equilibrium (see equation 50 in TGD).

In calculating the local PEC, the regional PEC is added to the local concentrations.

Results of $PEC_{\text{local sediment}}$ are given in Table 3.18.

Table 3.18: PEC_{local} for sediment for sites using TNPP

Uses	PEC_{sed} for TNPP (mg/kg ww) Log Kow 6; sol. 50 µg/L	PEC_{sed} for TNPP (mg/kg ww) QSAR values	PEC_{sed} for NP (mg/kg ww)
Production site A	0.211	2.03	-
Production site B	7.35	79.5	-
Production site C	115	1.24×10^3	-
PVC films, LLDPE films (formulation and processing)	20	217	0.53
PVC films, LLDPE films (service life)	0.425	4.36	0.18
Rubber			
- tyres	10.1	110	0.102
- soles	3.05	32.9	0.086
(formulation and processing)			
HDPE films (formulation and processing)	6.56	71	0.28
HDPE films (service life)	0.204	1.96	0.17
Other uses	6.93	75	0.27

3.1.4 Marine compartment

This section will be added when the exposure part for the aquatic compartment (freshwater and freshwater sediment) will be refined.

3.1.5 Terrestrial compartment

3.1.5.1 Calculated PEC for soil

PECs can be calculated for natural soil, agricultural soil and grassland using equations 51 to 67 in the TGD. These predicted environmental concentrations in soils take into account possible direct release of TNPP to soils, application of sewage sludge in agriculture and dry and wet deposition from the atmosphere.

Direct releases of TNPP to soils are expected to be negligible (see Table 3.7). In a same way, soil concentrations due to air deposition could be expected to be negligible because of low amount of TNPP released to air and a rapid degradation of TNPP in the atmosphere (see section 3.1.1.1.1).

The main contribution to TNPP concentration in soils is then expected to come from the application of sewage sludge (Cf. the high log Kow).

In calculating the local PEC, the regional PEC for natural soil is added to the local concentrations.

3.1.5.1.1 PEC_{local}soils at TNPP production sites

Final results of the PECs calculations for the three European TNPP production sites are presented in Table 3.19.

Table 3.19: PEC_{local} for the three European TNPP production sites

	PEC _{local} agr.soil mg.kg ⁻¹ wet wt. (averaged over 30 days) Log Kow 6; sol. 50 µg/L	PEC _{local} agr.soil mg.kg ⁻¹ wet wt. (averaged over 30 days) QSAR values	PEC _{local} agr.soil mg.kg ⁻¹ wet wt. (averaged over 180 days) Log Kow 6; sol. 50 µg/L	PEC _{local} agr.soil mg.kg ⁻¹ wet wt. (averaged over 180 days) QSAR values	PEC _{local} grassland mg.kg ⁻¹ wet wt. (averaged over 180 days) Log Kow 6; sol. 50 µg/L	PEC _{local} grassland mg.kg ⁻¹ wet wt. (averaged over 180 days) QSAR values
Site A	18	0.545	17.6	0.091	4.86	0.018
Site B	29.1	0.879	28.5	0.147	7.84	0.029
Site C	9.77x10 ⁻³	2.56x10 ⁻⁵	9.87x10 ⁻³	2.56x10 ⁻⁵	0.013	2.56x10 ⁻⁵

3.1.5.1.2 PEC_{local}soils at sites using TNPP

It is assumed that formulation and processing stage could occur at the same site. Therefore, in calculating local concentrations, added releases due to both stages are considered.

Table 3.20: PEC_{local,soil} for European TNPP processing sites

	Uses	PEC _{local} agr.soil mg.kg ⁻¹ wet wt. (averaged over 30 days)	PEC _{local} agr.soil mg.kg ⁻¹ wet wt. (averaged over 180 days)	PEC _{local} grassland mg.kg ⁻¹ wet wt. (averaged over 180 days)
TNPP Log Kow 6; sol. 50 µg/L	PVC films	32	31.3	8.62
	LLDPE films			
	Rubber	16.1	15.8	4.35
	- tyres			
	- soles	4.74	4.64	1.28
HDPE films	10.4	10.2	2.8	
Other uses	11	10.7	2.96	
TNPP QSAR values	PVC films	0.965	0.161	0.032
	LLDPE films			
	Rubber	0.487	0.081	0.016
	- tyres			
- soles	0.143	0.024	0.005	

	Uses	PEC _{local,agr.soil} mg.kg ⁻¹ wet wt. (averaged over 30 days)	PEC _{local,agr.soil} mg.kg ⁻¹ wet wt. (averaged over 180 days)	PEC _{local,grassland} mg.kg ⁻¹ wet wt. (averaged over 180 days)
	HDPE films	0.313	0.052	0.011
	Other uses	0.331	0.055	0.011
NP	PVC films LLDPE films	0.451	0.422	0.326
	Rubber			
	- tyres	0.281	0.279	0.270
	- soles	0.273	0.272	0.268
	HDPE films	0.325	0.316	0.285
	Other uses	0.317	0.309	0.282

3.1.5.1.3 PEC_{local,soils} during products service life

Table 3.21: PEC_{local,soil} for emission during the service life of products

	Uses	PEC _{local,agr.soil} mg.kg ⁻¹ wet wt. (averaged over 30 days)	PEC _{local,agr.soil} mg.kg ⁻¹ wet wt. (averaged over 180 days)	PEC _{local,grassland} mg.kg ⁻¹ wet wt. (averaged over 180 days)
TNPP Log Kow 6; sol. 50 µg/L	PVC films LLDPE films	0.525	0.514	0.142
	HDPE films	0.171	0.167	0.046
TNPP QSAR values	PVC films LLDPE films	0.016	2.67x10 ⁻³	5.55x10 ⁻⁴
	HDPE films	5.16x10 ⁻³	8.83x10 ⁻⁴	1.97x10 ⁻⁴
NP	PVC films LLDPE films	0.268	0.268	0.266
	HDPE films	0.266	0.266	0.265

3.1.6 Atmospheric compartment

In the calculation of PEC_{local} for air, emission from a point source as well as emission from a STP are taken into account.

For the generic TNPP production site, the concentration calculated at 100 m distance from the STP represents the major input for the PEC_{local} calculation whereas for all uses, TNPP concentration in air is mainly due to direct emission. Annual average predicted concentrations in air are calculated below.

In calculating the local PEC, the regional PEC is added to the local concentrations.

3.1.6.1.1 PEC_{local,air} at TNPP production sites

Final results of the PECs calculations for the three European TNPP production sites are presented in Table 3.22.

Table 3.22: PEC_{local} for the three European TNPP production sites

	PEC _{local,air,ann.} ($\mu\text{g}\cdot\text{m}^{-3}$) Log Kow 6; sol. 50 $\mu\text{g}/\text{L}$	PEC _{local,air,ann.} ($\mu\text{g}\cdot\text{m}^{-3}$) QSAR values
Site A	0.225	0.226
Site B	0.618	6.88×10^{-4}
Site C	1.26	6.88×10^{-4}

3.1.6.1.2 PEC_{local,air} at sites using TNPP

Table 3.23: PEC_{local} for air for sites using TNPP

Uses	PEC _{air} for TNPP ($\mu\text{g}\cdot\text{m}^{-3}$) Log Kow 6; sol. 50 $\mu\text{g}/\text{L}$	PEC _{air} for TNPP ($\mu\text{g}\cdot\text{m}^{-3}$) QSAR values	PEC _{air} for NP ($\mu\text{g}\cdot\text{m}^{-3}$)
PVC films	0.799	0.799	0.041
LLDPE films			
Rubber			
- tyres	0.096	0.439	0.003
- soles	0.029	0.156	0.003
HDPE films	0.260	0.260	0.015
Other uses	0.275	0.275	0.014

3.1.6.1.3 PEC_{local,air} during products service life

Table 3.24: PEC_{local} for emissions during the service life of products

Uses	PEC _{air} for TNPP ($\mu\text{g}\cdot\text{m}^{-3}$) Log Kow 6; sol. 50 $\mu\text{g}/\text{L}$	PEC _{air} for TNPP ($\mu\text{g}\cdot\text{m}^{-3}$) QSAR values	PEC _{air} for NP ($\mu\text{g}\cdot\text{m}^{-3}$)
PVC films			
LLDPE films	4.39×10^{-3}	1.22×10^{-3}	3.19×10^{-3}
HDPE films	1.85×10^{-3}	8.6×10^{-4}	1.53×10^{-2}

3.1.7 Non compartment specific effects relevant for the food chain (Secondary poisoning)

EUSES has been used to calculate the concentrations of TNPP in fish and earthworms. As a worst case values of 0.05 mg/L and 6.85 have been used for water solubility and log Kow,

respectively. Considering the chosen value for log Kow, a BMF of 10^6 is applied to the calculation of the PECs for TNPP and a BMF of 1 is taken for NP concentrations.

Table 3.25: Predicted Concentrations for secondary poisoning

Life Cycle Stage	Concentration of TNPP in fish from surface water for predators (mg.kg ⁻¹)	Concentration of TNPP in earthworms from agricultural soils (mg.kg ⁻¹ wet weight)	Concentration of NP in fish from surface water for predators (mg.kg ⁻¹)	Concentration of NP in earthworms from agricultural soils (mg.kg ⁻¹ wet weight)
TNPP use in PVC films and LLDPE films (formulation and processing)	1530	266	2.38	0.76
TNPP use in PVC films and LLDPE films (Service life)	45.8	10.4	0.80	0.49
TNPP used in rubber				
- tyres	781	137	0.91	0.51
- soles	241	44.7	0.84	0.50
(formulation and processing)				
TNPP use in HDPE films (formulation and processing)	508	90.4	1.29	0.57
TNPP use in HDPE films (Service life)	25.3	7.57	0.78	0.49
TNPP use in other applications (formulation and processing)	536	95.2	1.22	0.56

⁶ There are indications that the log Kow is higher than 7 (a QSAR value of 19 is available for example). Thus, taking into account a higher value for this parameter, BMFs of 3 (log Kow >8-9) or 1 (log Kow >9) could be applied.

3.2 EFFECTS ASSESSMENT: HAZARD IDENTIFICATION AND DOSE (CONCENTRATION) - RESPONSE (EFFECT ASSESSMENT)

3.2.1 Aquatic compartment (including sediment)

3.2.1.1 Fish

3.2.1.1.1 Acute toxicity

Table 3.26 shows a summary of the acute toxicity tests that were performed with fish species. The toxicity limits reported are above the upper limit of the estimated water solubility (solubility < 50 µg/L).

Table 3.26: Summary of acute toxicity tests with fish

Test #	Species	References	Comment	Validity*
1	Species: <i>Oncorhynchus mykiss</i> LC ₅₀ (96 hours) > 100 mg/L Method: OECD GL 203	Guterson, 2001	Concentrations tested were far above the solubility of the substance. No effect was seen at the highest concentration tested although no analytical monitoring was performed.	2
2	Species: <i>Brachydanio rerio</i> LC ₅₀ (96 hours) = < 10 mg/L LC ₅₀ (48 hours) = 16 mg/L Method: Dir. 84/449/EEC C.1	CIBA-Geigy, 1992a	The tested concentrations were probably very far above the actual water solubility of the substance. No analytical follow-up of the test concentrations was performed. As there was no equilibration time to allow dissolution of the substance during the preparation of the test concentration, it is not even clear that the maximum solubility in the test medium was achieved. The report mentions that undissolved substance was observed at all test concentrations.	3
3	Species: <i>Leuciscus idus</i> LC ₅₀ (48 hours) = 7.1 mg/L Method: DIN 38412-L15	CIBA-Geigy, 1988a	Concentrations tested were above the solubility of the substance and the results show no effect below the estimated upper limit of the water solubility of TNPP.	3

* 1 = valid; 2 = valid with restrictions; 3 = invalid; 4 = not assignable

Detailed descriptions of the tests are presented hereafter.

Test #1: the acute toxicity of an hydrolysed solution of TNPP (purity 99.8%) has been tested on *Oncorhynchus mykiss* according to the OECD guideline 203 (Guterson, 2001).

The fish were held 33 days before initiating the test on TNPP. Mortality in the stock culture was less than 0.1 % the week prior to test initiation. The fish were fed a daily ration of trout chow equal to 5 % of their body weight but were not fed 24 h prior to test initiation or during the test. The dilution water was dechlorinated City of Calgary tap water (charcoal filtered and aerated) and had a hardness of 198 mg CaCO₃/L, alkalinity of 140 mg CaCO₃/L, pH of 7.6, and a conductance of 446 ms/cm.

The test solutions were prepared from a stock solution initially containing 100.0 mg/L of TNPP. The solutions were gently aerated for 78 h at room temperature (20 ± 2 °C). The supernatants containing the hydrolysis products of TNPP were then decanted for preparation of the test solutions. The stock solutions and 200 L of dilution water were cooled to the test temperature overnight in a controlled environment chamber (15 °C with aeration).

At test initiation, dissolved oxygen, temperature, and pH ranged from 8.7 to 9.2 mg/L (98% to 100% saturation), 14 to 16 °C, and 7.7 to 8.0 units, respectively. At test termination, the temperature and pH of the test solutions were 15 °C and 7.8, respectively. Dissolved oxygen levels ranged from 6.2 to 6.8 mg/L (69 to 75 % saturation). The test solutions were only analysed for nonylphenol but nonylphenol was not detected in any of the test solutions collected at test initiation and termination.

There were no signs of stress or unusual behaviour exhibited by the fish in any of the treatment concentrations. No fish died at any concentration at any time point. The highest non-lethal concentration tested was set as greater than or equal to the 100 mg/L of TNPP hydrolysis products. LC₅₀ was > 100 mg/L after 24, 48, 72 and 96h.

This study should be considered as valid with restrictions. Indeed, tested concentrations were far above the water solubility of TNPP. Nonylphenol has been measured but not detected in any sample. The way test solutions were prepared should have enable the observation of effects triggered off by metabolites (nonylphenol). The result from this test could be used to support the fact that no toxicity of TNPP is expected above its water solubility (< 50 µg/L).

Test #2: the acute toxicity of TNPP (purity >94%) has been tested on *Brachydanio rerio* according to Directive 84/449/EEC, C.1 (CIBA-Geigy, 1992a). Five concentrations plus one control were tested (10, 18, 32, 58 and 100 mg/l). The control was performed in the test medium, i.e. dechlorinated tap water with an hardness of 171 mg CaCO₃/L. Other test parameters were as follow: pH between 7.3 and 7.9, temperature = 22 +/- 1°C. During the test, 10 fish were disposed per aquarium. They were acclimated 125 days prior the test and adapted to test medium 24 hour prior testing and no food was delivered 24 hours prior to exposure. A gentle aeration was started after 48 hours exposure. The test was conducted under a fluorescent light, 16 hours daily. The stock solution contained a mixture of 4 g. test substance and 160 mg Alkylphenol-Polyglycol-Ether (ARKOPAL) completed to 2 L with water.

During the test, the oxygen saturation ranged from 89-97% at 24 hours, 68-83% at 48 hours, and 60-76% at 72 hours. In the preliminary test, 10 mg/L TNPP had no effect to the fish after 96 hours of exposure. In the main test, 10 mg/L showed no effect to the fish after 48 hours. However, the oxygen concentration in the water was determined to be low at 48 hours and a gentle aeration was started at this time. After 72 hours of exposure with the test substance, all fish were dead. It is also important to notice that a small part of the test substance was swimming at the surface of the test vessels at all test times and concentrations.

No LC₅₀ could be estimated after 96h but some results were calculated at intermediate times: LC₅₀(48h)=16 mg/L (95% CL 12-19 mg/L) ; LC₅₀(24h)=29 mg/L (95% CL 23-36 mg/L). No mortality occurred in blank and in the vehicle controls.

Test #3: a static test was performed with *Leuciscus idus* (CIBA-Geigy, 1988a). Test organisms were acclimated 22 days with no food distribution three days prior to testing and for the test, mean fish size and weight were respectively 44 mm (35-50 mm) and 0.59 g. (0.29-0.85 g.). This led to a loading of 0.39 g/L in the test aquariums (test volume = 15 L.). 10 fish were disposed per concentration and control and dechlorinated tap water was used as dilution water. A hardness of 254 mg CaCO₃/L (Ca/Mg = 4/1) was measured. During the test, dissolved oxygen, pH and

temperature were measured at 0, 24 and 48 hours: $[O_2] > 91\%$ saturation, pH = 7.9-8.2 and T = 20 +/- 1°C. The test medium was gently aerated during the test and a fluorescent light was used 16 hours a day.

The stock solution of TNPP was prepared using a vehicle solvent, DMF. 5 g. of TNPP were dissolved in made up to 50 mL with DMF. This resulted in a concentration of DMF of 950 mg/L for the highest TNPP concentration tested.

Fish were exposed during 48 hours to six TNPP concentrations (5.8, 10, 18, 32, 58 and 100 mg/L) plus a blank and a control with the vehicle solvent used. Different symptoms were observed at the different test concentrations: moderate effects on swimming behaviour were observed after 24 and 48 hours at the concentration of 5.8 mg/L. Slight effects on the respiratory function has been observed after 48 hours, at 5.8 mg/L (one fish died at this concentration). All fish died at concentrations down to 10 mg/L. A LC_{50} of 7.1 mg/L was calculated.

Study #2 and #3 have to be considered as invalid:

- The tested concentrations were probably very far above the actual water solubility of the substance.
- No analytical follow-up of the test concentrations was performed. As there was no equilibration time to allow dissolution of the substance during the preparation of the test concentration, it is not even clear that the maximum solubility in the test medium was achieved. The report mentions that undissolved substance was observed at all test concentrations.
- All fish died at the lowest test concentration during aeration of the test system at t = 48 h.

3.2.1.1.2 Long-term toxicity

No chronic toxicity test with fish is available.

3.2.1.2 Aquatic invertebrates

3.2.1.2.1 Acute toxicity

Table 3.27 shows a summary of the acute toxicity tests that were performed with aquatic invertebrate species.

Table 3.27: Summary of acute toxicity tests with aquatic invertebrates

Test #	Species	References	Comment	Validity
1	Species: <i>Daphnia magna</i> EC ₅₀ (48 hours) = 0.009 mg/L Method: OECD GL 202	Hydroqual Laboratories Ltd, 2001a	The toxicity value is expressed as nonylphenol measured concentration (nonylphenol is the main hydrolysis product of TNPP).	2
2	Species: <i>Daphnia magna</i> EC ₅₀ (48 hours) = 0.42 mg/L Method: Dir. 84/449/EEC C.2	CIBA-Geigy, 1992b	No analytical monitoring was conducted neither for TNPP nor for its degradation product (nonylphenol). However, test result is comparable with the results of test #1 and other tests conducted with nonylphenol.	3

* 1 = valid; 2 = valid with restrictions; 3 = invalid; 4 = not assignable

Detailed descriptions of the tests are presented hereafter.

Test #1: (Hydroqual Laboratories Ltd, 2001a) The test was initiated with young daphnids less than 24 h old from in-house cultures. Mortality in the stock culture was less than 1% in the week prior to test initiation. Dilution water was dechlorinated City of Calgary tap water (charcoal filtered and aerated). The dilution water had a hardness of 188 mg CaCO₃/L, alkalinity of 100 mg CaCO₃/L, pH of 8.1, and conductivity of 421 ms/cm. The ratios of calcium-to-magnesium and sodium-to-potassium on a weight-to-weight basis were 3.4 and 4.0 respectively. The concentration of dissolved oxygen was 8.2 mg/L (100 % saturation at the test temperature 20 +/- 1°C).

The test solutions were prepared from a stock solution initially containing 100 mg/L of TNPP. The mass of TNPP selected for the test was based on initial attempts to get enough of the hydrolysis products in solution to be acutely toxic to *Daphnia magna*. The method detailed below provided a stock solution that was acutely lethal to *Daphnia magna*.

TNPP (100 mg) was weighed onto a glass Petri dish. The dish and test substance were placed into a two-litre, glass Erlenmeyer flask containing 1 L of dilution water. A magnetic stir bar was added and the mouth of the flask sealed with Parafilm®. The test substance was gently stirred for 78 h at room temperature (20 ± 2 °C). The supernatant containing the hydrolysis products of TNPP was then decanted for preparation of the test solutions. A stock was prepared from the hydrolysed TNPP solution by diluting 100 mL of the supernatant with 900 mL of dilution water (10 mg/L nominal). This solution was then serially diluted with laboratory dilution water to obtain the other eight test concentrations (5.00, 2.50, 1.25, 0.63, 0.31, 0.16, 0.08, and 0.04 mg/L). The concentrations were nominal values based on the total mass of TNPP initially added to the flask and hydrolysed for 78 h (100.0 mg/L).

The organisms were then added to the test vessels in a random fashion (final loading density of one organism per 10 mL of test solution). There were four replicates for each test concentration containing 5 daphnids. The daphnids were not fed during the test. Beakers were placed on a tray and covered with a glass sheet. The test was conducted at conditions similar to the culture conditions. The test vessels were examined at 24 and 48 h, and the number of immobilised organisms recorded along with any observations of unusual behaviour.

The samples of the test solutions were analysed for the major hydrolysis product of TNPP, nonylphenol. Nonylphenol was only detected in the highest treatment at test initiation (0.3 mg/L based on the results of duplicate analyses; detection limit of 0.2 mg/L). Toxicity values were derived based on this measured concentration of nonylphenol. The test concentrations for toxicity values were derived from the single measured value available for nonylphenol (starting value that was serially diluted by a factor of 2 to obtain the numerical values for the test concentrations, all of which were below the detection limit of 0.2 mg/L for nonylphenol).

At test initiation the concentration of dissolved oxygen, temperature, and pH ranged from 8.2 to 8.3 mg/L (99% saturation), 19°C, and 8.1 to 8.3 units, respectively. At test termination, the concentration of dissolved oxygen, temperature, and pH ranged from 7.6 to 7.8 mg/L (96 to 100% saturation), 21°C, and 8.2 to 8.3 units, respectively. Dead organisms were considered immobilised.

Toxicity values were derived based on nominal concentrations for the mixture of TNPP hydrolysis products. These nominal values were likely higher than actual concentrations because of the sparingly soluble nature of the test substance and hydrolysis products. The concentrations

and 95 % confidence limits of the hydrolysis products that immobilised 50 % of the daphnids at 24 and 48 h were 2.2 mg/L (1.7 to 3.0 mg/L) and 0.3 mg/L (0.2 to 0.4 mg/L), respectively. This would correspond to a 24-h LC50 of 66 µg/L and a 48-h LC 50 of 9 µg/L expressed as estimated nonylphenol. The toxic response and presence of detectable levels of the hydrolysis product in solution confirmed that the TNPP had undergone hydrolysis during preparation of the stock solution. TNPP is not soluble in water and the only major hydrolysis product is nonylphenol. Hence, nonylphenol is likely the toxic agent present in the test solutions. The low toxicity result could also be attributed to physical effect although there was no identification of the presence of undissolved material during this test.

No explanation can be found to explain the low toxicity observed during this short-term toxicity testing with daphnids. Indeed, the toxicity observed could not be attributed solely to nonylphenol measured in the test medium if we refer to the EU risk assessment available on this substance.

Industry is asked to perform another test to solve this issue (and to analyze both TNPP – if possible, and NP). This study could also be used in order to test the possible constraints of the realisation of a long-term test that should be performed subsequently. Indeed, TNPP being suspected to be highly hydrophobic, other exposure routes should be study.

Test #2: (CIBA-Geigy, 1992b) Calculated amounts of the test material to produce the desired concentrations were added to the water and were homogeneously distributed. Values are based on the nominal concentrations. Parts of the test substance were visible on the surface of the water at concentrations of 0.1-1.0 mg/L.

One day before the start of exposure, reproductive *Daphnia* are separated from the young (0-24 hours old) by sieving all individuals through an 800 µm sieve. This procedure is repeated immediately prior to exposure and the young are retained for the test. The *Daphnia* (4 replicates of 5 *Daphnia* each) were then transferred into the beakers. Cultures of *Daphnia* were maintained in glass vessels containing approximately 2.5 litres of reconstituted water and maintained at 20 +/- 1°C. The oxygen content ranged from 97 to 103%, the pH ranged from 7.8 to 8.0, and the water temperature was maintained at 21-24°C throughout the experiment.

The EC-50 values were calculated according to the maximum likelihood method, probit model. EC-values were graphically determined on gausso-logarithmic probability paper. The EC₅₀ values at 24 and 48 h were 2.6 and 0.42 mg/L, respectively.

This study has to be considered as invalid:

- The tested concentrations were probably very far above the actual water solubility of the substance.
- No analytical follow-up of the test concentrations was performed. As there was no equilibration time to allow dissolution of the substance during the preparation of the test concentration, it is not even clear that the maximum solubility in the test medium was achieved. The report mentions that undissolved substance was observed at all test concentrations.

3.2.1.2.2 Long-term toxicity

No chronic toxicity test with aquatic invertebrates is available.

3.2.1.3 Algae

Table 3.28 shows a summary of the toxicity tests that were performed with algae species.

Table 3.28: Summary of toxicity tests with algae

Test #	Species	References	Comment	Validity*
1	Species: <i>Selenastrum capricornutum</i> NOEC (72 hours) 100 mg/L (growth rate) Method: OECD GL 201	Hydroqual Laboratories Ltd, 2001b	No significant effects upon algae growth were observed at any test concentration. On the contrary, it seems that the hydrolysis of TNPP during the experiment has increase the phosphorous content of the test medium causing growth stimulation.	2
2	Species: <i>Scenedesmus subspicatus</i> NOEC (72 hours) 100 mg/L (biomass) Method: Dir. 87/302/EEC, part C., p. 89	CIBA-Geigy, 1992c	No significant effects upon biomass were observed at any test concentration.	2

* 1 = valid; 2 = valid with restrictions; 3 = invalid; 4 = not assignable

Detailed descriptions of the tests are presented hereafter.

Test #1: (Hydroqual Laboratories Ltd, 2001b) The test was initiated with exponentially growing cells from in-house cultures maintained at $23 \pm 2^\circ\text{C}$ under continuous light (3,770 lux). The cultures were grown under axenic conditions in 2-L flasks containing 1 L of artificial media, aerated with filtered sterile air. Cell numbers were obtained from optical density measurements at 430 nm calibrated against particle and cell counts at test termination. The dilution water was dechlorinated City of Calgary tap water (charcoal filtered and aerated) spiked with nutrients. The dilution water had a hardness of 198 mg CaCO_3/L , alkalinity of 146 mg CaCO_3/L , pH of 7.6, and conductance of 446 ms/cm.

The test solutions were prepared from a stock solution initially containing 100 mg of TNPP in 1 L of dilution water. The substance was weighed on a glass Petri dish (100 mg) and the dish placed into a 2 L glass, Erlenmeyer flask containing 1 L of dilution water. A magnetic stir bar was added and the mouth of the flask sealed with Parafilm®. The test substance was stirred gently for 78 hours at room temperature ($21 \pm 2^\circ\text{C}$). The test solutions were then prepared from the stock solution of TNPP hydrolysis products as recommended by the OECD for the testing of difficult substances. A 100 mL volume of the hydrolysed stock solution was poured into a 250 mL plastic container for the highest test concentration (100 mg/L nominal test concentration). A second 100 mL volume of the stock solution was poured into another 250-mL container and serially diluted with 100-mL volumes of dilution water to obtain the remaining test concentrations (50.0, 25.0, 12.5, 6.3, 3.1, and 1.6 mg/L nominal test concentrations).

The solutions were spiked with 1 mL of a concentrated nutrient solution and then inoculated (1 mL) to give an initial cell density of $9,664 + 154$ cells/mL. The inoculum was taken from an exponentially growing culture, washed twice with a sodium bicarbonate solution, and the cell number adjusted to give the desired initial cell density in the 100-mL test volume.

The test was conducted in a controlled environment chamber at $23 \pm 2^\circ\text{C}$ under continuous light with intensity at the plate surface of 4,370 lux provided by cool white fluorescent lights.

Two sets of samples were collected for chemical analysis. The first set consisted of samples of the test solutions and control at test initiation. The second set consisted of samples of the test solutions and control incubated under the test conditions for 72 h. The samples were not analysed for TNPP because it is insoluble in water. The samples of the test solutions were analysed for nonylphenol however it was not detected in any of the samples (detection limit of 0.2 mg/L).

The pH at test initiation and termination in the controls and 100.0 mg/L test solution ranged from 7.0 to 8.0. The initial and final control cell densities were 9,664 cells/mL and 404,000 cells/mL, respectively. This was a 42-fold increase in cell density over the 72-h test period. A 16-fold increase was required for a valid test. The test medium contains 0.65 mg/L phosphate. Complete hydrolyses of the test substance (100 mg/L) would yield approximately 12 mg/L of phosphorous acid. The cell density in the highest test concentration at 72 h was 344 % greater than the controls. This represents approximately 1.5 additional doublings of the cell population exposed to the hydrolysed TNPP solution when compared to the controls. The result indicates that hydrolysis of TNPP causes growth stimulation due to the liberation of phosphorous. The LOEC as well as the 24, 48 and 72 h EC₅₀ values were >100 mg/l. The NOEC was the highest concentration tested of 100 mg/l. The level of nonylphenol present in the test solutions under the conditions in which the stock solution was prepared, diluted, and tested was not toxic to unicellular green alga, *Raphidocelis subcapitata*.

Test #2: (CIBA-Geigy, 1992c) Nominal test concentrations of 0, 1.23, 3.7, 11, 33 and 100 mg/L were used (three replicates for the test concentrations and 6 replicates for the blank).

The stock solution was prepared by mixing 200 mg of the test substance with 80 mL water and 1 mL of a 0.8% alkylphenol-polyglycol ether and made up to 100 mL with water. This 100 mL solution was then made up to 1 liter with water. Calculated amounts of the stock solution to produce the desired test concentrations were added to the water. The algae were then transferred into the flasks (100 mL Erlenmeyer flasks, stoppered with aluminium caps, on Lab-Shaker). The cell densities were measured at 24, 48, and 72 hour. The temperature was continuously measured and maintained at $23 \pm 1^\circ\text{C}$. The pH was measured at 0 and 72 hours and ranged from 7.8 to 8.1. The test was conducted under continuous illumination, cold white fluorescent light, $118 \mu\text{E}/\text{m}^2 \text{ sec} \pm 20\%$ (approx. 8000 lux.).

No significant effects upon biomass were observed at any test concentration.

3.2.1.4 Micro-organisms

A respiration inhibition test was conducted using TNPP on activated sludge from the sewage treatment plant of CH-4153 Reinach (CIBA-Geigy, 1988b). The OECD guideline 209 was followed. Sludge concentration was 1.6-1.7 g/L and the temperature was $20 \pm 2^\circ\text{C}$ during the test. The test was performed with dechlorinated drinking water. A reference substance (3,5-dichlorophenol) was also tested and an IC₅₀ of 16 mg/L was determined. A deviation from the guideline is highlighted: instead of a centrifuged sludge, a settled sludge was used. Due to the very low solubility and the expected low toxicity of the substance, only one concentration (100 mg/L) was tested in duplicates during three hours. The test substance was directly added to the test vessel. In one replicate, no inhibition was recorded, in the other, an inhibition of 24% was

observed. This test must be considered invalid as 25% inhibition were found in a replicate. Consequently a NOEC cannot be determined.

As TNPP is not considered as readily biodegradable, the result of the first biodegradability test presented in this report (Hydroqual Laboratories Ltd, 2001c) is not useable for the determination of the $PNEC_{\text{microorganisms}}$. However a supplementary assay was conducted during test #2 of this report (CIBA-Geigy, 1994). Indeed the test substance has also been tested with the reference substance in presence of the inoculum in order to control the toxicity and inhibition of the bacteria's activity by the test substance. The controls of reference and reference together with the test substance meet the specification for ready biodegradability. Therefore, it can be concluded that the test substance has no inhibitory effect on the bacteria at the concentration tested (18.1 mg/L) which is above the solubility limit of TNPP.

3.2.1.5 Calculation of Predicted No Effect Concentration (PNEC)

3.2.1.5.1 Surface water

The PNEC derived in the risk assessment report of nonylphenol will be used for the risk characterisation relating to the NP formed during the use of TNPP

$PNEC_{\text{water}}(\text{NP}) = 0.33 \mu\text{g/L}$ (based on a chronic study with the algae *Scenedesmus subspicatus* and an assessment factor of 10).

Toxicity tests available for TNPP tend to indicate that no toxicity is expected above its water solubility ($< 50 \mu\text{g/L}$).

3.2.1.5.2 Sewage treatment plants

No valid test with micro-organisms is available. However, a supplementary test conducted during a ready biodegradability test tends to demonstrate that TNPP has no inhibitory effect on bacteria at concentrations below its solubility limit. According to the TGD, a PNEC for sewage treatment plants can be derived using an assessment factor of 10 on the non-inhibitory concentration from a ready biodegradation test, i.e. a PNEC of $>1.8 \text{ mg/l}$ can be derived for TNPP.

For nonylphenol, the $PNEC_{\text{STP}}$ of 9.5 mg/L will be retained (E.C., 2002).

3.2.1.5.3 Sediment

In the absence of any ecotoxicological data for sediment-dwelling organisms, the $PNEC_{\text{sed}}$ could be provisionally calculated using the equilibrium partitioning method. However, according to the toxicity tests conducting with aquatic organisms, it seems that TNPP will not have a toxic effect below the upper limit of the estimated water solubility, no $PNEC_{\text{water}}$ could have been derived for TNPP. Consequently, the equilibrium partitioning method cannot be applied to calculate a $PNEC_{\text{sediment}}$ for TNPP.

For nonylphenol, the $PNEC_{\text{sed}}$ of $39 \mu\text{g/kg ww}$ will be retained (E.C., 2002).

3.2.2 Marine compartment

This section will be added when the exposure part for the aquatic compartment (freshwater and freshwater sediment) will be refined.

3.2.3 Terrestrial compartment

3.2.3.1 Terrestrial effect data

No data available.

3.2.3.2 Calculation of PNEC_{soil}

In the absence of any ecotoxicological data for soil organisms, the PNEC_{soil} could be provisionally calculated using the equilibrium partitioning method. However, according to the toxicity tests conducting with aquatic organisms, it seems that TNPP will not have a toxic effect below the upper limit of the estimated water solubility, no PNEC_{water} could have been derived for TNPP. Consequently, the equilibrium partitioning method cannot be applied to calculate a PNEC_{soil} for TNPP.

For nonylphenol, the PNEC_{soil} of 300 µg/kg ww will be retained (E.C., 2002).

3.2.4 Atmosphere

No data available.

3.2.5 Non compartment specific effects relevant to the food chain (secondary poisoning)

A PNEC_{oral} can be calculated based on a NOAEL of 167 mg/kg bw/d determined during a 90-d repeated dose toxicity study on rats.

From this NOAEL, a NOEC of 3340 mg.kg_{food}⁻¹ can be calculated using a conversion factor of 20 (applicable to a test performed on *Rattus norvegicus* - > 6 weeks).

The PNEC_{oral} is then determined using an assessment factor of 90 (duration of the test on mammals is 90 days):

$$PNEC_{oral} = \frac{TOX_{oral}}{AF_{oral}} = \frac{3340}{90} = 37 \text{ mg.kg}_{\text{food}}^{-1}$$

For nonylphenol, a PNEC_{oral} of 10 mg.kg_{food}⁻¹ has been determined (E.C., 2002).

3.3 RISK CHARACTERISATION

Nota: the risk assessment is based on the use of standard TNPP. It should be noticed that two grades of TNPP are put on the marketplace, one with around 5% residual NP, the other one containing less than 0.1% residual NP. The risk assessment of the high pure TNPP would lead the calculation of local concentrations of NP reduced to approximately one third of the one currently available in this report.

3.3.1 Aquatic compartment (including sediment)

Surface water

Results of the RCR calculations for the three European TNPP production sites and for the sites where TNPP is used are presented in Table 3.29 and Table 3.30 for sewage treatment plants and water for NP. Only RCRs for STP for TNPP have been calculated as no effect has been shown for TNPP above its water solubility. However, works are still needed on this aspect and the risk characterisation for TNPP in surface water could be updated based on the results of the conclusion (i) program.

Table 3.29: RCR for STP for the three European TNPP production sites

	RCR _{STP} for TNPP Log Kow 6; sol. 50 µg/L	RCR _{STP} for TNPP QSAR values
Site A	<0.036	<0.047
Site B	<0.058	<0.075
Site C	<0.364	<0.476

Table 3.30: RCR for STP and water for sites using TNPP

Uses	RCR _{STP} for TNPP Log Kow 6; sol. 50 µg/L	RCR _{STP} for TNPP QSAR values	RCR _{STP} for NP	RCR _{water} for NP
PVC films LLDPE films	<0.063	<0.083	3.25x10 ⁻³	11.1
Rubber				
- tyres	<0.032	<0.042	2.88x10 ⁻⁴	2.64
- soles	<0.009	<0.012	1.44x10 ⁻⁴	2.23
HDPE films	<0.021	<0.027	1.05x10 ⁻³	4.83
Other uses	<0.022	<0.028	1.12x10 ⁻³	5.01

Table 3.31: RCR for STP and water for the service life of plastics containing TNPP

Uses	RCR _{STP} for TNPP Log Kow 6; sol. 50 µg/L	RCR _{STP} for TNPP QSAR values	RCR _{STP} for NP	RCR _{water} for NP
PVC films LLDPE films	<1x10 ⁻³	<1x10 ⁻³	5.5x10 ⁻⁵	1.98

Uses	RCR _{STP} for TNPP Log Kow 6; sol. 50 µg/L	RCR _{STP} for TNPP QSAR values	RCR _{STP} for NP	RCR _{water} for NP
HDPE films	<3x10 ⁻⁴	<4x10 ⁻⁴	1.78x10 ⁻⁵	1.87

Furthermore, it should be noted that a risk at the regional level has been calculated at **1.81** for nonylphenol (E.C., 2002).

All local PEC_{water} for NP are above the PNEC_{water} that is derived from the risk assessment on NP (0.33 µg/L). A risk is identified for all stages of the life cycle of standard TNPP except for the production step for which it has been estimated that no NP emission can be assumed⁷.

Sediment compartment

There is a lack of toxicity data that does not enable the determination of a PNEC for this compartment for TNPP. Consequently, no risk characterisation can be done for TNPP.

RCRs calculated from the exposure to NP due to the use of standard TNPP are presented in the following table. For nonylphenol a PNEC of 39 µg/kg ww has been derived (E.C., 2002).

Table 3.32: PEC_{local} for sediment for sites using TNPP

Uses	RCR _{sed} for NP
PVC films	13.7
LLDPE films	
Rubber	
- tyres	2.64
- soles	2.23
HDPE films	7.4
Other uses	7.0

Table 3.33: PEC_{local} for sediment for the service life of plastics containing TNPP

Uses	RCR _{sed} for NP
PVC films	4.56
LLDPE films	
HDPE films	4.45

Furthermore, it should be noted that the contribution of the regional concentrations to the RCR calculated for NP in sediment is **2.64** (E.C., 2002). Consequently, based on NP exposure due to the use of TNPP and effect assessment, a risk is identified for all stages of the life cycle of standard TNPP except for the production step for which it has been estimated that no NP emission can be assumed⁸. No risk characterisation can be done for TNPP as ecotoxicological

⁷ This is in accordance with the RAR for NP where no risk has been identified for the use of NP for TNPP production

⁸ This is in accordance with the RAR for NP where no risk has been identified for the use of NP for TNPP production

test results are lacking.

Conclusions to the risk assessment for the aquatic compartment

Sewage treatment plants (exposure to TNPP and NP)

(ii) There is at present no need for further information and/or testing or for risk reduction measures beyond those that are being applied already.

This conclusion applies to all stages of the life cycle of TNPP.

Freshwater (exposure to TNPP)

(i) There is a need for further information and/or testing.

This conclusion applies to all stages of the life cycle of TNPP.

- There is a need for more information for the effect assessment of TNPP. A new acute toxicity test on *Daphnia* is requested. A long-term testing should be performed subsequently in the case no effect is observed at short term.

Update on the work performed to answer this request: a short-term test with daphnids has been performed by Industry. However, some drawbacks associated with the chemical analysis were identified during the test and the study should be considered invalid (low recovery rates found with the TNPP analysis; too high nominal concentrations of TNPP tested leading to sufficient residual NP concentrations to generate an effect). Based on this experience, a new test is currently being setting-up.

Freshwater (exposure to NP)

(iii) There is a need for limiting the risks; risk reduction measures that are already being applied should be taken into account.

This conclusion applies to all stages of the life cycle of standard TNPP, except for the production step.

The exposure concentrations of NP due to the use of standard TNPP lead to a risk for the aquatic compartment. The PNEC for nonylphenol (E.C., 2002) has been calculated using an assessment factor of 10 on the lowest result of three chronic tests on different trophic levels. Therefore, it is not expected that the PNEC for NP can be realistically refined.

Or

(i) There is a need for further information and/or testing.

The risk characterisation of TNPP (exposure to NP) could be refined based on monitoring studies (NP measurements) at processing sites.

Sediment (exposure to TNPP)

(i) There is a need for further information and/or testing.

This conclusion applies to all stages of the life cycle of standard TNPP.

- Concerning the sediment compartment, no data are available on the toxicity of TNPP toward benthic organisms. Considering the low solubility in water and the suspected high adsorption potential of TNPP, toxicity on sediment dwelling organisms should be studied. Toxicity testings on sediment organisms should be done for the determination of the $PNEC_{sed}$.

Sediment (exposure to NP)

(iii) There is a need for limiting the risks; risk reduction measures that are already being applied should be taken into account.

This conclusion applies to all stages of the life cycle of standard TNPP, except for the production step.

Or

(i) There is a need for further information and/or testing.

As the exposure assessment of NP is based on the equilibrium partitioning method, any refinement of the exposure assessment of NP in freshwater will lead to a refinement of the exposure assessment in sediment (see conclusion (i) for freshwater).

3.3.2 Marine compartment

This section will be added when the exposure part for the aquatic compartment (freshwater and freshwater sediment) will be refined.

3.3.3 Terrestrial compartment

There is a lack of toxicity data that does not enable the determination of a PNEC for this compartment. Consequently, no risk characterisation can be done for TNPP.

RCRs calculated from the exposure to NP due to the use of TNPP are presented in the following table. For nonylphenol a PNEC of 0.3 mg/kg ww has been derived (E.C., 2002).

Table 3.34: PEC_{local} for soil for sites using TNPP

Uses	RCR _{soil} for NP
PVC films	1.5
LLDPE films	
Rubber	
- tyres	0.938
- soles	0.911
HDPE films	1.08
Other uses	1.1

Table 3.35: PEC_{local} for soil for the service life of plastics containing TNPP

Uses	RCR _{soil} for NP
PVC films	0.894
LLDPE films	
HDPE films	0.887

Furthermore, it should be noted that a risk at the regional level has been calculated at **0.88** for nonylphenol (E.C., 2002).

Based on NP exposure and effect assessment, a risk is identified for sites using standard TNPP (formulation and processing combined), except for rubber products.

No risk characterisation can be done for TNPP as ecotoxicological test results are lacking.

Conclusions to the risk assessment for the terrestrial compartment

Soil (exposure to TNPP)

(i) There is a need for further information and/or testing.

This conclusion applies to all stages of the life cycle of TNPP.

- Considering the suspected high adsorption potential of TNPP, toxicity on soil organisms should be studied. Toxicity testings on soil organisms should be done for the determination of the PNEC_{soil}.

Soil (exposure to NP)

(ii) There is at present no need for further information and/or testing or for risk reduction measures beyond those that are being applied already.

This conclusion applies to all stages of the life cycle of TNPP.

Risks have been identified in soil for the use of standard TNPP (formulation and processing combined), except for rubber products. However, this risk is due to the contribution of the regional concentration of NP. Considering the fact that a risk reduction strategy has been developed for NP, its application should also cover the risks identified in this risk assessment.

3.3.4 Atmosphere

No risk characterisation can be carried out for the air compartment since there is no specific effect data.

3.3.5 Non compartment specific effects relevant for the food chain (Secondary poisoning)

Results of the risk characterisation for secondary poisoning are presented in Table 3.36.

Table 3.36: RCRs for secondary poisoning

Uses	RCR _{food} for TNPP	RCR _{terr,food,ch} for TNPP	RCR _{food} for NP	RCR _{terr,food,ch} for NP
PVC films LLDPE films (formulation and processing)	41.3	7.16	0.238	0.076
PVC films LLDPE films (Service life)	1.23	0.282	0.080	0.049
Rubber - tyres - soles (formulation and processing)	21 6.48	3.7 1.2	0.091 0.084	0.051 0.050
HDPE films (formulation and processing)	13.7	2.44	0.129	0.057
HDPE films (Service life)	0.683	0.204	0.078	0.049
Other uses (formulation and processing)	14.4	2.57	0.132	0.058

Based on TNPP exposure and effect assessment, a risk is identified for sites using standard TNPP (formulation and processing combined), except for the service life of HDPE films and PVC/LLDPE films (for the latter, there is no risk only for terrestrial organisms).

For NP, no risk is identified for the releases due to the use of TNPP.

Conclusions to the risk assessment for secondary poisoning

Secondary poisoning (exposure to TNPP)

(i) There is a need for further information and/or testing.

This conclusion applies to all stages of the life cycle of TNPP.

- The bioaccumulation potential of TNPP should be further studied. There are already indications that the bioconcentration factor of TNPP could be low (Cf. Annex 2). Furthermore, the exposure concentrations calculated using this worst case scenario are not plausible in the environment compared to the amount of TNPP released in the environment.

Secondary poisoning (exposure to NP)

(ii) There is at present no need for further information and/or testing or for risk reduction measures beyond those that are being applied already.

This conclusion applies to all stages of the life cycle of TNPP.

3.3.6 PBT assessment

3.3.6.1 PBT assessment for TNPP

- The vP criterion is fulfilled as the substance is non readily biodegradable based on a negative result at a test on ready biodegradability performed according to OECD guidelines 301B and 301D.
- The vB criterion is fulfilled based on the bioaccumulation potential determined with the log Kow of 6.85 taken as a worst case. A log BCF of 4.66 has been calculated for fish.
- Concerning the T criterion, no aquatic toxicity is expected at concentrations above the water solubility of TNPP based on the available set of information. However, a short-term test and long-term test with daphnids are requested.

Conclusions to PBT assessment

(i) There is a need for further information and/or testing.

- Based on the available data, TNPP would be classified as vPvB. However, the vB criterion is fulfilled based on a BCF calculated from an estimated log Kow taken as a worst case. Refinement of this parameter is necessary to conclude the PBT assessment of this chemical.

3.3.6.2 PBT assessment for NP

Properties of NP have been extracted from the EU risk assessment report available for this substance (E.C., 2002).

- Nonylphenol is considered inherently biodegradable. However, a half-life in surface water has been estimated at 150 days. Hence the vP criterion is fulfilled (half-life > 60 days).
- The B criterion is not fulfilled based on the BCF of 1,280 used in the European risk assessment report (BCF < 2000).
- The T criterion is fulfilled since NOECs < 0.01 mg/L have been identified for fish and invertebrates for example.

Based on the properties of nonylphenol, it appears that this substance is neither PBT nor vPvB.

4 HUMAN HEALTH

4.1.1 Human exposed via the environment

The following sections will be updated when agreement will be found in the environmental exposure assessment.

4.1.1.1 Indirect exposure via the environment

4.1.1.2 Human exposed via the environment

4.1.1.2.1 Summary of risk characterisation for exposure via the environment

5 RESULTS

5.1 ENVIRONMENT

Section not updated, see section 3.3.

5.2 HEALTH

6

REFERENCES

- Building Research Establishment Ltd. (2001). <http://www.bre.co.uk/>
- Chemical Information Services (2002). <http://www.chemicalinfo.com/>
- CIBA-Geigy (1988a). Report on the test for acute toxicity test of TK 10417 to Golden Orfe (*Leuciscus idus*), unpublished report, Report No 88 41 26. Basel, Switzerland, June 20, 1999, CIBA-GEIGY: 9.
- CIBA-Geigy (1988b). Report on the test for inhibitory concentration of TK 10417 [TNPP] on aerobic bacteria (OECD-GUIDELINE N° 209), unpublished report, Project No 88 41 73. Basle, Switzerland, June 08, 1988, CIBA: 8.
- CIBA-Geigy (1992a). Report of the acute toxicity test of IRGANOS TNPP to zebra-fish (*Brachydanio rerio*). Basel, Switzerland, August 28, 1992, CIBA: 12.
- CIBA-Geigy (1992b). Report on the acute toxicity test of IRGANOS TNPP on daphnia (*Daphnia magna Straus 1820*). Basel, Switzerland, August 28, 1992, CIBA: 11.
- CIBA-Geigy (1992c). Report on the growth inhibition test of IRGAFOS TNPP to green algae (*Scenedesmus subspicatus*), unpublished report, Report No 92 81 36. Basel, Switzerland, August 28, 1992, CIBA-GEIGY: 12.
- CIBA-Geigy (1994). Report on the test for ready biodegradability of IRGASTAB CH 55 in the carbondioxide evolution test, unpublished report, Test No 94 80 08. Basle, Switzerland, April 18, 1994, CIBA-GEIGY: 11.
- Crompton (2003). WESTON TNPP Phosphite, product information, 08-01-2003.
- Dover (unpublished). Dover Chemical Report TSWR94-179.
- Dover Chemical Corporation (2001). Product technical datasheet for Doverphos HiPure 4 TNPP, Dover: 6 pp.
- E.C. (2002). European Union Risk Assessment Report 4-Nonylphenol (branched) and nonylphenol, Volume 10.
- E.C. (2003). Technical Guidance Document on Risk Assessment in support of Commission Directive 93/67/EEC on Risk Assessment for new notified substances, Commission Regulation (EC) N° 1488/94 on Risk Assessment for existing substances, Directive 98/8/EC of the European Parliament and of the Council concerning the placing of biocidal products on the market. Luxembourg, Office for Official Publications of the European Communities.
- E.C. (2004a). EUSES 2.0, the European Union System for the Evaluation of Substances. National Institute of Public Health and the Environment (RIVM).
- E.C. (2004b). EUSES, the European Union System for the Evaluation of Substances, Version 2.0. National Institute of Public Health and the Environment (RIVM).
- Guterson, D. S. (2001). Acute toxicity of trisnonylphenyl phosphite (TNPP) to fish (Rainbow trout; OECD203), Unpublished report no. 20000865/20002538. Calgary, Alberta, Canada, August 27, 2001, HydroQual Laboratories, Ltd., for General Electric Company, Pittsfield, MA.
- Howe, S. R., P. Surana, M. R. Jakupca and L. Borodinsky (2001). "Potential dietary exposure to p-nonylphenol from food-contact use of tris(nonylphenyl)phosphite (TNPP)." Food Additives and Contaminants 18(11): 1021-1039.
- Hydroqual Laboratories Ltd (2001a). Acute toxicity of trisnonylphenyl phosphite (TNPP) to *Daphnia magna* (OECD 202). Calgary, August 2001, Hydroqual.
- Hydroqual Laboratories Ltd (2001b). Algal growth inhibition test (OECD 201) on trisnonylphenyl phosphite (TNPP). Calgary, August 2001, Hydroqual.
- Hydroqual Laboratories Ltd (2001c). Ready biodegradability of trisnonylphenyl phosphite (TNPP; OECD 301D; CAS 26523-78-4). Calgary, November 2001, Hydroqual.

OECD (2004). Emission scenario document on plastics additives, Number 3, 24 June 2004, Organisation for Economic Co-operation and Development: 125.

OECD (unpublished). SIDS Initial Assessment Report for Tris(2,4-di-tert-butylphenyl)phosphite (CAS 31570-04-4) sponsored by United Kingdom and adopted at SIAM 18 (April 2004).

Opperhuizen, A., E. W. v. d. Volde, F. A. P. C. Gobas, D. A. K. Liem, J. M. D. v. d. Steen and O. Hutzinger (1985). "Relationship between bioconcentration in fish and steric factors of hydrophobic chemicals." *Chemosphere* 14(11-12): 1871-1896.

Phoenix_Chemical_Laboratory (1997). Vapor pressure by isoteniscope (ASTM D2879). Chicago, May 19, 197, General Electric Specialty Chemicals: 1.

Pittsburgh_Testing_Laboratory (1978). Flash Point. Pittsburgh, July 12, 1978: 1.

Reimer&Associates (2001a). Physical/chemical property of tris-nonylphenyl phosphite (TNPP) [26523-78-4] - Boiling temperature (OECD 103). Pittsfield, August 2001, General Electric Company: 10.

Reimer&Associates (2001b). Physical/chemical property of tris-nonylphenyl phosphite (TNPP) [26523-78-4] - Hydrolysis as a function of pH (OECD 111). Pittsfield, August 2001, General Electric Company: 20.

Reimer&Associates (2001c). Physical/chemical property of tris-nonylphenyl phosphite (TNPP) [26523-78-4] - Melting (pour) point (OECD 102). Pittsfield, August 2001, General Electric Company: 10.

Reimer&Associates (2001d). Physical/chemical property of tris-nonylphenyl phosphite (TNPP) [26523-78-4] - n-Octanol/Water partition coefficient. Pittsfield, August 2001, General Electric Company: 4.

Reimer&Associates (2001e). Physical/chemical property of tris-nonylphenyl phosphite (TNPP) [26523-78-4] - Solubility in Water (OECD 105). Pittsfield, August 2001, General Electric Company: 19.

SPIN (2007). Substances in Preparations in Nordic Countries. 2007. <http://www.spin2000.net/spin.html>

Staples, C. A. (2001). Fugacity modeling to estimate transport between environmental compartments for tris-nonylphenyl phosphite (TNPP) (CAS Reg. No.26523-78-4). Fairfax, November 2001, Assessment Technologies, Inc.: 5.

TNO (2004). Preliminary study on the hydrolysis of tris(nonylphenol)phosphite (TNPP). Zeist, 20th September 2004, TNO Nutrition and Food Research: 11.

Ullmann (1985). Ullmann's encyclopedia of industrial chemistry, vol., VCH.

United States Testing Company, I. (1990). Report of Test. Hoboken, New Jersey, February 6, 1990, United States Testing Company, Chemical Services Division: 1.

US EPA and Syracuse Research Corporation (2001). EPI Suite, v.3.10, US EPA.

US-EPA and Syracuse Research Corporation (2004). EPI Suite, v.3.12 (30th November 2004), US EPA.

GLOSSARY

Standard Abbreviation	term	Explanation/Remarks and Alternative Abbreviation(s)
<i>Ann.</i>		Annex
AF		assessment factor
BCF		bioconcentration factor
bw		body weight / <i>Bw, b.w.</i>
°C		degrees Celsius (centigrade)
CAS		Chemical Abstract System
CEC		Commission of the European Communities
CEN		European Committee for Normalisation
CEPE		European Council of the Paint, Printing Ink and Artists' Colours Industry
d		day(s)
d.wt		dry weight / dw
DG		Directorate General
DT ₅₀		period required for 50 percent dissipation (define method of estimation)
DT _{50lab}		period required for 50 percent dissipation under laboratory conditions (define method of estimation)
DT ₉₀		period required for 90 percent dissipation (define method of estimation)
DT _{90field}		period required for 90 percent dissipation under field conditions (define method of estimation)
EC		European Communities
EC		European Commission
EC ₅₀		median effective concentration
EEC		European Economic Community
EINECS		European Inventory of Existing Commercial Chemical Substances
EU		European Union
EUSES		European Union System for the Evaluation of Substances
f _{oc}		Fraction of organic carbon
G		gram(s)

PNEC(s)	Predicted No Effect Concentration(s)
PNEC _{water}	Predicted No Effect Concentration in Water
(Q)SAR	Quantitative Structure Activity Relationship
STP	Sewage Treatment Plant
TGD	Technical Guidance Document ⁹
UV	Ultraviolet Region of Spectrum
UVCB	Unknown or Variable composition, Complex reaction products or Biological material
v/v	volume per volume ratio
w/w	weight per weight ratio
w	gram weight
GLP	Good Laboratory Practice
h	hour(s)
ha	Hectares / h
HPLC	High Pressure Liquid Chromatography
IARC	International Agency for Research on Cancer
C ₅₀	median immobilisation concentration or median inhibitory concentration 1 / <i>explained by a footnote if necessary</i>
ISO	International Standards Organisation
IUPAC	International Union for Pure Applied Chemistry
kg	kilogram(s)
kPa	kilo Pascals
K _{oc}	organic carbon adsorption coefficient
K _{ow}	octanol-water partition coefficient
K _p	Solids water partition coefficient
l	litre(s)
log	logarithm to the basis 10
L(E)C ₅₀	Lethal Concentration, Median
LEV	Local Exhaust Ventilation
m	Meter
µg	microgram(s)

⁹ Commission of the European Communities, 1996. Technical Guidance Documents in Support of the Commission Directive 93/67/EEC on risk assessment for new substances and the Commission Regulation (EC) No 1488/94 on risk assessment for existing substances. Commission of the European Communities, Brussels, Belgium. ISBN 92-827-801[1234]

mg	milligram(s)
MAC	Maximum Accessibility Concentration
MOS	Margins Of Safety
NOAEL	No Observed Adverse Effect Level
NOEC	No Observed Effect Concentration
NOEL	No Observed Effect Level
OEL	Occupational Exposure Limit
OECD	Organisation for Economic Co-operation and Development
OJ	Official Journal
pH	potential hydrogen <i>-logarithm</i> (to the base 10) of the hydrogen ion concentration {H ⁺ }
pKa	<i>-logarithm</i> (to the base 10) of the acid dissociation constant
pKb	<i>-logarithm</i> (to the base 10) of the base dissociation constant
Pa	Pascal unit(s)
PEC	Predicted Environmental Concentration
STP	Sewage Treatment Plant
WWTP	Waste Water Treatment Plant

European Commission

**EUR .:[[click here to insert EUR No.](#)] - European Union Risk Assessment Report
[[click here to insert SUBSTANCE NAME, and volume no.](#)]**

*Editors: B.G. Hansen, S.J. Munn, S. Pakalin, C.J.A. Heidorn, R. Allanou , S. Scheer,
G. Pellegrini, S. Vegro, J.De Bruijn, M.Luotamo, K. Vormann, H. Loonen, F. Berthault,
A. Naughton, V. Anfossi, L. Praderio.([keep this updated](#))*

Luxembourg: Office for Official Publications of the European Communities

2000 – VIII pp. .:[[click here to insert page count, number of pages.](#)]
pp. – 17.0 x 24.0 cm

Environment and quality of life series

ISBN .:[[click here to insert ISBN No.](#)]

Price (excluding VAT) in Luxembourg: EUR .:[[click here to insert price.](#)]

DRAFT

European Union Risk Assessment Report

TRIS (NONYLPHENYL) PHOSPHITE

CAS-No.: 26523-78-4

EINECS-No.: 247-759-6

RISK ASSESSMENT

DRAFT

LEGAL NOTICE

Neither the European Commission nor any person acting on behalf of the Commission is responsible for the use which might be made of the following information

A great deal of additional information on the European Union is available on the Internet.
It can be accessed through the Europa Server
(<http://europa.eu.int>).

DRAFT

Cataloguing data can be found at the end of this publication

Luxembourg: Office for Official Publications of the European Communities, 2002
ISBN ([insert number here](#))

© European Communities, 2002 ([insert correct year here](#))
Reproduction is authorised provided the source is acknowledged.

Printed in Italy

TRIS(NONYLPHENYL) PHOSPHITE

CAS No.: 26523-78-4
EINECS No.: 247-759-6

RISK ASSESSMENT

Draft of February 2006

France

Rapporteur for the risk assessment of tris(nonylphenyl) phosphite is the Ministry of Ecology and Sustainable Development as well as the Ministry of Employment and Social Affairs in co-operation with the Ministry of Public Health. Responsible for the risk evaluation and subsequently for the contents of this report is the rapporteur.
The scientific work on this report has been prepared by :

Environment risk assessment

National Institute for Industrial Environment and Risks (INERIS)
Direction of chronic risks
Parc Technologique Alata
BP n°2
60550 Verneuil-en-Halatte

Human health risk assessment

Effect assessment, exposure and risk characterisation for consumers

National Institute for Industrial Environment and Risks (INERIS)
Direction of chronic risks
Parc Technologique Alata
BP n°2
60550 Verneuil-en-Halatte

Exposure and risk characterisation for workers

Institut National de recherche et Sécurité (INRS)
Département Risques Chimiques et Biologiques
30, rue Olivier Noyer
75680 Paris cedex 14

This Draft Risk Assessment Report is under the responsibility of the Member State rapporteur. In order to avoid possible misinterpretations or misuse of the findings in this

draft, anyone wishing to cite or quote this report is advised to contact the Member State rapporteur beforehand.

DRAFT

Date of Last Literature Search :

[please insert year]

Review of report by MS Technical Experts finalised:

[please insert month and year]

Final report:

[please year]

DRAFT

Foreword

This Draft Risk assessment Report is carried out in accordance with Council Regulation (EEC) 793/93¹ on the evaluation and control of the risks of “existing” substances. “Existing” substances are chemical substances in use within the European Community before September 1981 and listed in the European Inventory of Existing Commercial Chemical Substances. Regulation 793/93 provides a systematic framework for the evaluation of the risks to human health and the environment of these substances if they are produced or imported into the Community in volumes above 10 tonnes per year.

There are four overall stages in the Regulation for reducing the risks: data collection, priority setting, risk assessment and risk reduction. Data provided by Industry are used by Member States and the Commission services to determine the priority of the substances which need to be assessed. For each substance on a priority list, a Member State volunteers to act as “Rapporteur”, undertaking the in-depth Risk Assessment and recommending a strategy to limit the risks of exposure to the substance, if necessary.

The methods for carrying out an in-depth Risk Assessment at Community level are laid down in Commission Regulation (EC) 1488/94², which is supported by a technical guidance document³. Normally, the “Rapporteur” and individual companies producing, importing and/or using the chemicals work closely together to develop a draft Risk Assessment Report, which is then presented at a Meeting of Member State technical experts for endorsement. The Risk Assessment Report is then peer-reviewed by the Scientific Committee on Toxicity, Ecotoxicity and the Environment (CSTEE), now renamed Scientific Committee on Health and Environmental Risks (SCHER) which gives its opinion to the European Commission on the quality of the risk assessment.

This Draft Risk Assessment Report has undergone a discussion in the Competent Group of Member State experts with the aim of reaching consensus by interpreting the underlying scientific information, or including more data, but this work has not yet been totally finalised. The information contained in this Draft Risk Assessment Report does not, therefore, necessarily provide a sufficient basis for decision making regarding the hazards, exposures or the risks associated with the priority substance.

This Draft Risk Assessment Report is under the responsibility of the Member State rapporteur. In order to avoid possible misinterpretations or misuse of the findings in this draft, anyone wishing to cite or quote this report is advised to contact the Member State rapporteur beforehand.

¹ O.J. No L 084, 05/04/199 p.0001 – 0075

² O.J. No L 161, 29/06/1994 p. 0003 – 0011

³ Technical Guidance Document, Part I – V, ISBN 92-827-801 [1234]

0 OVERALL RESULTS OF THE RISK ASSESSMENT

CAS Number: 26523-78-4
EINECS Number: 247-759-6
IUPAC Name: Phenol, nonyl-, phosphite (3:1)

Environment

To be updated

Human health effects assessment

Risk assessment of human exposed via the environment was not discussed and will be updated following the update of environment risk assessment.

() (i) There is a need for further information and/or testing.

(X) (ii) There is at present no need for further information and/or testing and no need for risk reduction measures beyond those, which are being applied already.

This conclusion applies to the assessment of the risk to human health through consumer exposure.

(X) (iii) There is a need for specific measures to limit the risks.

This conclusion applies to the assessment of the risk to human health through worker exposure. It is reached because of concerns for sensitisation as a consequence of dermal exposure arising during manufacture of the substance, manufacture of products or use of preparations containing TNPP.

CONTENTS

1	GENERAL SUBSTANCE INFORMATION	4
1.1	IDENTIFICATION OF THE SUBSTANCE	4
1.2	PURITY/IMPURITIES, ADDITIVES	4
1.2.1	<i>Purity</i>	4
1.2.2	<i>Additives</i>	5
1.3	PHYSICO-CHEMICAL PROPERTIES	5
1.3.1	<i>Physical state (at ntp)</i>	5
1.3.2	<i>Melting point</i>	5
1.3.3	<i>Boiling point</i>	5
1.3.4	<i>Relative density</i>	5
1.3.5	<i>Vapour pressure</i>	5
1.3.6	<i>n-octanol / water partition coefficient</i>	7
1.3.7	<i>Water solubility</i>	8
1.3.8	<i>Flash point</i>	8
1.3.9	<i>Autoflammability</i>	8
1.3.10	<i>Explosivity</i>	9
1.3.11	<i>Oxidising properties</i>	9
1.3.12	<i>Viscosity</i>	9
1.3.13	<i>Henry's Law constant</i>	9
1.3.14	<i>Summary of physico-chemical properties</i>	10
1.4	CLASSIFICATION	10
1.4.1	<i>Current classification</i>	10
1.4.2	<i>Proposed classification</i>	10
2	GENERAL INFORMATION ON EXPOSURE	11
2.1	PRODUCTION	11
2.1.1	<i>Production process</i>	11
2.1.2	<i>Production capacity</i>	12
2.2	USES	13
2.2.1	<i>Introduction</i>	13
2.2.1.1	<i>Industrial use</i>	15
2.2.1.2	<i>Production of Polyvinylchloride (PVC) film</i>	15
2.2.1.3	<i>Production of Polyolefins linear low density polyethylene (LLDPE)</i>	16
2.2.1.4	<i>Production of High density polyethylene (HDPE)</i>	16
2.2.1.5	<i>Production of rubber</i>	16
2.2.2	<i>Other applications</i>	16
2.2.2.1	<i>Use of end-products</i>	17
2.2.2.2	<i>Recovery and disposal</i>	17
2.3	TRENDS	17
3	ENVIRONMENT	19
4	HUMAN HEALTH	20
4.1	HUMAN HEALTH (TOXICITY)	20
4.1.1	<i>Exposure assessment</i>	20
4.1.1.1	<i>General introduction</i>	20
4.1.1.2	<i>Occupational exposure</i>	20
4.1.1.2.1	<i>Scenario 1 : Manufacture of TNPP</i>	22
4.1.1.2.2	<i>Scenario 2 : manufacture of products containing TNPP</i>	22
4.1.1.2.3	<i>Scenario 3 : use of preparations containing TNPP</i>	24
4.1.1.2.4	<i>Occupational exposure summary</i>	24
4.1.1.3	<i>Consumer exposure</i>	25
4.1.1.3.1	<i>Introduction</i>	25
4.1.1.3.2	<i>Potential exposure from migration of TNPP from food contact materials</i>	25
4.1.1.3.3	<i>Consumer exposure summary</i>	29
4.1.1.4	<i>Indirect exposure via the environment</i>	29
4.1.2	<i>Effects assessment : hazard identification and dose (concentration)-response (effect) assessment</i>	30
4.1.2.1	<i>Toxicokinetics, metabolism and distribution</i>	30
4.1.2.2	<i>Acute toxicity</i>	31

4.1.2.2.1	Oral	31
4.1.2.2.2	Inhalation	33
4.1.2.2.3	Dermal	33
4.1.2.2.4	Other	34
4.1.2.2.5	Summary of acute toxicity	34
4.1.2.3	Irritation.....	35
4.1.2.3.1	Skin	35
4.1.2.3.2	Eye	36
4.1.2.3.3	Summary of irritation.....	38
4.1.2.4	Corrosivity.....	38
4.1.2.5	Sensitisation	38
4.1.2.5.1	Skin sensitisation.....	38
4.1.2.5.2	Summary of sensitisation	40
4.1.2.6	Repeated-dose toxicity	40
4.1.2.6.1	Animal data	40
4.1.2.6.2	Summary and discussion of repeated dose toxicity	47
4.1.2.7	Mutagenicity.....	48
4.1.2.7.1	Studies <i>in vitro</i>	48
4.1.2.7.2	Studies <i>in vivo</i>	52
4.1.2.7.3	Summary of mutagenicity	52
4.1.2.8	Carcinogenicity	52
4.1.2.9	Toxicity to reproduction.....	55
4.1.3	Risk characterisation.....	62
4.1.3.1	General aspects.....	62
4.1.3.2	Workers.....	64
4.1.3.2.1	Acute toxicity.....	64
4.1.3.2.2	Irritation.....	65
4.1.3.2.3	Sensitisation	65
4.1.3.2.4	Repeated dose toxicity	65
4.1.3.2.5	Mutagenicity	66
4.1.3.2.6	Carcinogenicity	66
4.1.3.2.7	Toxicity to reproduction.....	66
4.1.3.3	Consumers.....	67
4.1.3.3.1	Introduction.....	67
4.1.3.3.2	Risk characterisation due to migration from food contact materials	67
4.1.3.3.3	Summary of risk characterisation for consumers	68
4.1.3.4	Human exposed via the environment	68
4.1.3.4.1	Summary of risk characterisation for exposure via the environment	68
4.2	HUMAN HEALTH (PHYSICO-CHEMICAL PROPERTIES).....	69
5	RESULTS.....	70
5.1	ENVIRONMENT	70
5.2	HEALTH	70
6	REFERENCES	71
	Annex 1.....	79

TABLES

Table 1-1: Vapour pressure data for TNPP.....	6
Table 1-3: Parameters used for the vapour pressure calculation according to the modified Watson correlation method	7
Table 1-4: Physical and chemical properties of the TNPP.....	10
Table 2-1: Typical quantitative breakdown of TNPP Uses.....	13
Table 2-2: Industrial uses of TNPP in the Nordic Countries (in Tonnes)	14
Table 2-3: Use pattern of TNPP in the Nordic Countries (in Tonnes).....	14
Table 2-4: Global food contact regulations specific to TNPP.....	16
Table 2-6: Industrial Categories (IC), Use Categories (UC) and Main categories (MC) used in EUSES calculations	17
Table 4-1: Summary of reasonable worst case exposures.....	24
Table 4-2: Use of trisnonylphenylphosphite in food-packaging polymers	26
Table 4-3: Potential TNPP exposure from LLDPE.....	28
Table 4-4: Potential TNPP exposure from HIPS.....	29
Table 4-5: Acute oral toxicity of TNPP	32

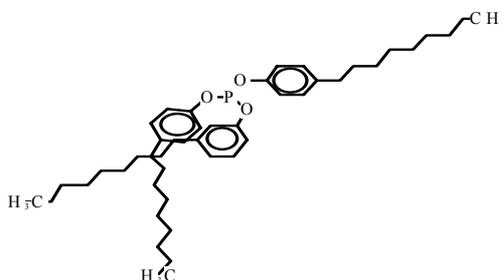
Table 4-6: Skin mean reaction scores (4-hours exposure)	35
Table 4-7: Skin mean reaction scores (24-hours exposure)	36
Table 4-8: Rabbit eye irritation scores (conjunctiva mean reaction scores).....	38
Table 4-9: Summary of Histopathological Findings in F0 generation rats (Negative findings omitted)	53
Table 4-10: Summary of gross and histopathological findings in dogs (negative findings omitted)	54
Table 4-11: Comparison of first two matings in three generations of rats	57
Table 4-12: Summary of F1 offspring toxicity	59
Table 4-13: Calculated internal doses for workers.....	64
Table 4-14: MOSs for systemic effects by repeated exposure	65
Table 4-15: MOSs for reproductive effects by repeated exposure	66
Table 4-16: MOS for systemic effects by repeated exposure.....	67
Table 4-17: MOS for reproductive effects by repeated exposure	67

1 GENERAL SUBSTANCE INFORMATION

Parts of this section will be updated in the next version of the environmental risk assessment.

1.1 IDENTIFICATION OF THE SUBSTANCE

CAS No: 26523-78-4
EINECS No: 247-759-6
IUPAC Name: Phenol, nonyl-, phosphite (3:1)
Molecular formula: C₄₅H₆₉O₃P
Structural Formula:



Molecular weight: 689 g.mol⁻¹
Synonyms and tradenames: Alkanox TNPP, Lowinox TNPP, Irgafos TNPP, Tris(monononylphenyl)phosphite, Tri(nonylphenyl)phosphite, Weston 399, Weston TNPP, Irgastab CH 55, Naugard TNPP, Polygard, Polygard HR, Polygard LC, TNPP, Trisnonylphenylphosphit.

In this assessment, the name Tris(nonylphenyl)phosphite (TNPP) will be used for the substance as this is the most common name.

1.2 PURITY/IMPURITIES, ADDITIVES

1.2.1 Purity

The purity of TNPP is reported as ca. 95 – 100% w/w.

The following impurities may be found in TNPP :

- | | |
|---|-------------|
| - Nonylphenol (CAS 25154-52-3) | < 5% w/w, |
| - Phenol (CAS 108-95-2) | < 1% w/w, |
| - Di(nonylphenyl)phenylphosphite (CAS 25417-08-7) | 0.05% w/w, |
| - Chlorine (CAS 7782-50-5) | 0.005% w/w. |

1.2.2 Additives

1,1',1''-nitritotripropan-2-ol (CAS No: 122-20-3) is an additive that may be found in TNPP in the proportion of 0.5 to 1% w/w.

1.3 PHYSICO-CHEMICAL PROPERTIES

1.3.1 Physical state (at ntp)

TNPP is a viscous liquid at room temperature.

1.3.2 Melting point

Instead of a melting point, a pour point of $6^{\circ}\text{C} \pm 3^{\circ}\text{C}$ was determined (Reimer&Associates, 2001e). A melting point could not be observed using the differential scanning calorimetric (DSC) method because an endothermic event was not observed in the heat flow vs temperature plot. The pour point (the lowest temperature at which the test substance is first observed to flow on warming) is an appropriate measurement for viscous liquid substances. The test was conducted according to ASTM Method D97, as recommended in the OECD 102 guideline.

1.3.3 Boiling point

The boiling point was reported as $>303^{\circ}\text{C}$ (Reimer&Associates, 2001f). The test method was based on OECD 103 guideline. Bubbling was observed for the first 1 to 2 seconds of heating, and then stopped. This was probably due to the boiling of a minor component ($<0.1\%$) present in the test substance. Consequently a new study was undertaken to assess the true boiling point. The TNPP producers have determined that TNPP will begin to degrade before boiling. According to a Thermal Gravimetric Analysis (TGA) of TNPP, the phosphite has an onset of degradation at 322°C under nitrogen.

1.3.4 Relative density

The relative density has been quoted at 0.98 g.cm^{-3} at 20°C (Crompton, 2003).

1.3.5 Vapour pressure

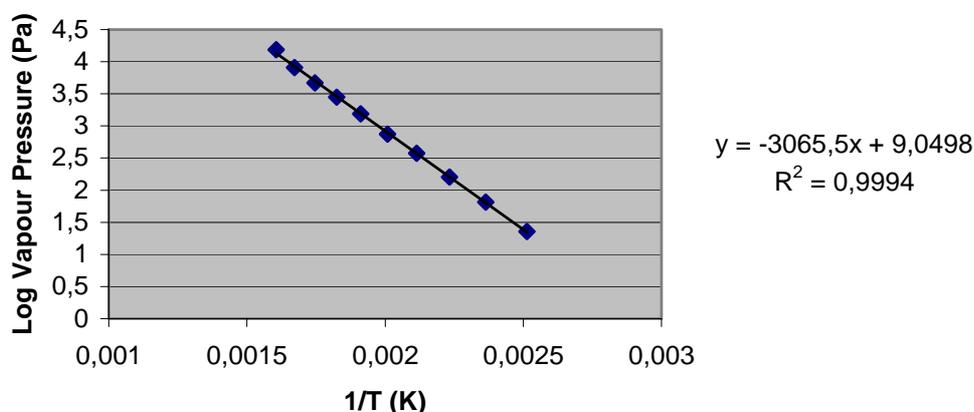
A vapour pressure was estimated using structure activity relationships models developed by the U.S. Environmental Protection Agency and Syracuse Research Corporation (EPIWIN, v. 3.10, US EPA and Syracuse Research Corporation, 2001). The vapour pressure was estimated to 5.10^{-12} Pa (Staples, 2001).

Another value of 0.047 Pa at 20°C was extrapolated from results obtained by isoteniscope (method ASTM D2879) at temperatures ranging from 125 to 375°C (Phoenix_Chemical_Laboratory, 1997). These measured values are displayed in Table 1-1.

Table 1-1: Vapour pressure data for TNPP

Temperature °C	Vapour Pressure (Pa)
125	22.7
150	65.3
175	160
200	373
225	747
250	1533
275	2800
300	4666
325	8133
350	15330
375	65330

A strong relation between the temperature ($1/T$) and the vapour pressure is found. Excluding the last value measured at 375°C, the plot of the above results gives a linear regression with a good reliability (see figure below). Vapour pressures of respectively 0.039 Pa and 0.058 Pa at 20°C and 25°C could be derived from this equation. These results are consistent with the extrapolated value of 0.047 Pa at 20°C found in the study summary in the IUCLID file.

**Figure 1-1: Linear regression between the temperature ($1/T$) and the measured vapour pressures**

The isoteniscope method is recommended for the measurement of vapour pressures between 10^2 and 10^5 Pa. The extrapolated value is three orders of magnitude below this range. Consequently, the value of 0.058 Pa at 25°C would need to be confirmed by another vapour pressure result.

The modified Watson correlation method was also used for the estimation of TNPP vapour pressure. According to this method the vapour pressure can be calculated using the boiling point value in the following equation:

Equation 1-1: Calculation of the vapour pressure according to the modified Watson correlation method

$$\ln P_{vp} = \frac{\Delta H_{VB}}{\Delta Z_b RT_b} \left[1 - \frac{(3 - 2 \frac{T}{T_b})^m}{T/T_b} - 2m(3 - 2 \frac{T}{T_b})^{m-1} \ln \frac{T}{T_b} \right], \text{ with } \frac{\Delta H_{VB}}{T_b} = K_F (8.75 + R \ln T_b)$$

The parameters were chosen as follow:

Table 1-2: Parameters used for the vapour pressure calculation according to the modified Watson correlation method

Parameter	Value	Remark
K_F	1.06	Default
T_b	322°C or 595.15 K	Boiling point, see 1.3.3
ΔZ_b	0.97	Default estimation
T	25°C or 298.15 K	Chosen temperature for subsequent modelling stages
m	0.19	Default value for liquids
R	1.9859 cal.mol ⁻¹ .K ⁻¹	Gas constant (8.314 Pa.m ³ .mol ⁻¹ .K ⁻¹)

The calculation gives a vapour pressure of $3.86 \cdot 10^{-7}$ atm corresponding to 0.039 Pa. The main drawback of this estimation lies in the fact that the method employed only used the boiling point as experimental data entry and the only data available is the temperature of degradation of the substance: 322°C.

On one hand, the extrapolated value of 0.058 Pa has been calculated based on vapour pressures measured by the isoteniscope method which is not recommended for this range of vapour pressure. On the other hand, the vapour pressure has been calculated with the modified Watson correlation method using the temperature of degradation of the substance as a boiling point. Both methods give similar results.

Finally, the value of 0.058 Pa at 25°C, extrapolated from measured vapour pressures at higher temperatures and confirmed by an estimation method, will be used in the risk assessment.

1.3.6 n-octanol / water partition coefficient

The n-octanol-water partition coefficient was estimated using structure activity relationships models developed by the U.S. Environmental Protection Agency and Syracuse Research Corporation (EPIWIN, US EPA and Syracuse Research Corporation, 2001). The log P_{ow} was estimated to 20.05 (US EPA and Syracuse Research Corporation, 2001).

According to Reimer & Associates, 2001c, it was not appropriate to conduct the partition coefficient measurement because the solubility of TNPP in water was too low (see section 1.3.7) and TNPP was also found to be hydrolytically unstable. The n-octanol / water partition coefficient was therefore calculated using the software from Advanced Chemistry Development Inc. (“ACD/LogP DB”). The result of the calculation was found to be 21.6 ± 0.6 (Reimer&Associates, 2001c).

The annex of the OECD guideline 117 presents some K_{ow} calculation methods that can be used to “provide an estimate when experimental methods cannot be applied”. However there are some limitations to the use of such methods. First, the reliability of calculation methods decreases as the complexity of the compound under study increases. Here, TNPP could be classified as a rather complex molecule with a high molecular weight and several functional groups. The domain of application of K_{ow} calculation methods is characterised in terms of chemical structures. For example, some calculation programs cannot be applied to the estimation of K_{ow} for phosphorus compounds including phosphites. Second, the domains of the models is also restricted by the log K_{ow} range of their applicability. In general, clear estimates can be expected

in the region of log K_{ow} 0-5. Some programs have shown good estimates for compounds with log $K_{ow} > 5$ but estimates for log K_{ow} around 10 or above should be considered rather as qualitative than quantitative information (TGD, Part III, Chapter 4, E.C., 2003).

Considering the high hydrophobic potential of TNPP which contains 27 aliphatic and 18 aromatic carbons, a high log Kow value could be expected for this compound. However, in the absence of other data, the highest recommended value of 8 will be used in EUSES model (E.C., 2004a).

1.3.7 Water solubility

A water solubility was estimated using structure activity relationships models developed by the U.S. Environmental Protection Agency and Syracuse Research Corporation (EPIWIN, US EPA and Syracuse Research Corporation, 2001). The water solubility was estimated to $1.3 \cdot 10^{-15}$ mg.L⁻¹ (Staples, 2001).

Experimental water solubility was determined by (Reimer&Associates, 2001a). The flask method based on OECD Guideline 105 was used. TNPP was not detected in the saturated aqueous test solution. Therefore it is concluded that the water solubility of TNPP is below the detection limit of the substance. This detection limit was estimated to be 0.6 mg.L⁻¹, the lowest TNPP concentration that produced a signal that is reliably distinguished from the background signal as determined from chromatograms of TNPP solutions. Therefore, the water solubility of TNPP is < 0.6 mg.L⁻¹ at 24°C.

It was not possible to determine a more accurate result. As a matter of fact, no saturated solution could be obtained during the experiment because of the rapid hydrolysis of TNPP in water. The experimental procedure consisted in mixing TNPP in deionized water for at least 24 hours. Half life of TNPP being estimated to 13 hours, it can be supposed that all the TNPP is degraded during the first day.

In this risk assessment report, when a water solubility result is needed for environmental modelling purposes, the value of 0.6 mg.L⁻¹ will be retained as a worst case estimation. However, it will be also considered that TNPP is almost instantly degraded into nonylphenol and phosphorous acid when released into aquatic compartments.

1.3.8 Flash point

Values of 183°C (internal reference, Great Lakes Chemical, Italia, Milan) and 195°C (Ciba MSDS) were reported using closed cup methods.

Besides, a value of 207°C was reported using the Pensky-Martin apparatus (closed cup) (Pittsburgh_Testing_Laboratory, 1978). This last value will be retained in this risk assessment because the analytical report was available.

1.3.9 Autoflammability

In a MSDS by Uniroyal, a value of 268°C was quoted. Moreover, using the Setchkin method, a result of 440°C was found (United States Testing Company, 1990).

1.3.10 Explosivity

No result could be found in the literature on any explosion limit. However, on the basis of its chemical structure, TNPP is not expected to have explosive properties.

1.3.11 Oxidising properties

No oxidising property was reported for TNPP (internal reference, Great Lakes Chemical, Milan, Italia).

1.3.12 Viscosity

In a product information sheet, a value of 6000 cps at 25°C is quoted (Crompton, 2003). Other values are also presented in this document showing that the viscosity goes from 15000 cps at 15°C to 18 cps at 120°C. The value at 25°C will be retained for the risk assessment.

1.3.13 Henry's Law constant

The Henry's law constant was estimated using structure activity relationships models developed by the U.S. Environmental Protection Agency and Syracuse Research Corporation (EPIWIN, v. 3.10, sub-model HENRYWIN, US EPA and Syracuse Research Corporation, 2001). At 25°C, a value of 66.1 Pa.m³.mol⁻¹ was calculated (US EPA and Syracuse Research Corporation, 2001).

The Henry's law constant can also be estimated from the ratio of the vapour pressure to the water solubility (E.C., 2003):

$$HENRY = \frac{VP \cdot MOLW}{SOL}$$

Using a vapour pressure of 0.058 Pa, a molecular weight of 689 g.mol⁻¹ and a water solubility of 0.6 mg.L⁻¹ the Henry's Law constant is equal to 66.6 Pa.m³.mol⁻¹.

This result is coherent with the QSAR calculation above so the value of 66.6 Pa.m³.mol⁻¹ will be retained in this risk assessment.

1.3.14 Summary of physico-chemical properties

The physico-chemical properties of TNPP used in this risk assessment are summarised in the following table:

Table 1-3: Physical and chemical properties of the TNPP

Property	Value	Comments
Physical state at ntp	Viscous liquid	
Molecular weight	689 g.mol ⁻¹	
Melting Point	6°C ± 3°C	Instead of a melting point, a pour point (more appropriate to viscous liquids) was determined
Boiling Point	322°C	Degradation
Relative density	0.98 g.cm ⁻³	
Vapour pressure	0.058 Pa at 25°C	extrapolated from results obtained by isoteniscope (method ASTM D2879)
Partition coefficient	Log Kow = 21.6 Log Kow = 8 (EUSES)	Calculated with software ACD/LogP DB
Water solubility	<0.6 mg.L ⁻¹	A saturated solution was not obtained and the water solubility result corresponds to the detection limit of the analytical method.
Flash point	207°C	Pensky Martin apparatus (closed cup)
Autoflammability	440°C	Setchkin method
Oxidising properties	No oxidising property	
Henry's law constant	66.6 Pa.m ³ .mol ⁻¹	TGD calculation

1.4 CLASSIFICATION

1.4.1 Current classification

TNPP chemical is not classified under Annex I of Directive 67/547 EEC.

1.4.2 Proposed classification

Human health effects (adopted classification)

Classification was finalised in the Commission working group on the Classification and Labelling of Dangerous Substances in November 2005 (human health):

Symbol : Xi

R-phrase : R43 : May cause sensitization by skin contact.

Environmental effects

To be updated.

2 GENERAL INFORMATION ON EXPOSURE

This section will be updated in the next version of the environmental risk assessment.

2.1 PRODUCTION

TNPP is produced all over the world: Unites States, Europe, India, Korea, Russia, China, etc. (Chemical Information Services, 2002). Three facilities are currently producing TNPP in Europe. On the other hand, the major source of TNPP to Europe is from the United States.

2.1.1 Production process

The manufacturing processes used to produce TNPP are reasonably similar in the various plants in the US and Europe. Figure 2-1 is providing an overview of a typical production process.

TNPP production is carried out in a closed system where nonylphenol (NP) and phosphorus trichloride (PCl_3) are added to the reactor (ca. 3 :1) and held at greater than 110°C to ensure all the PCl_3 is consumed. The HCl by-product is vented to an absorber. The HCL by-product can be filtered and stored for sale or use in other processes. Excess nonylphenol is stripped from the product. The stripped nonylphenol can be recycled. The product TNPP in the reactor after stripping is pumped to a storage tank for packaging and sale. The product may be packaged into drums, isotaners, rail cars, or tank trunks.

Environmental release and exposure

The process is fully automated (computer controlled) in a closed system. The reactor is operated under 3-5 lbs (1.4 – 2.3 kg) of pressure. The vacuum pump vent is the only potential process release to the atmosphere, and it is passed through a carbon filter. The storage tank is kept under nitrogen preventing release to the atmosphere. Nitrogen is also used during transfer and packaging.

Trisnonylphenyl Phosphite (TNPP) Process Overview

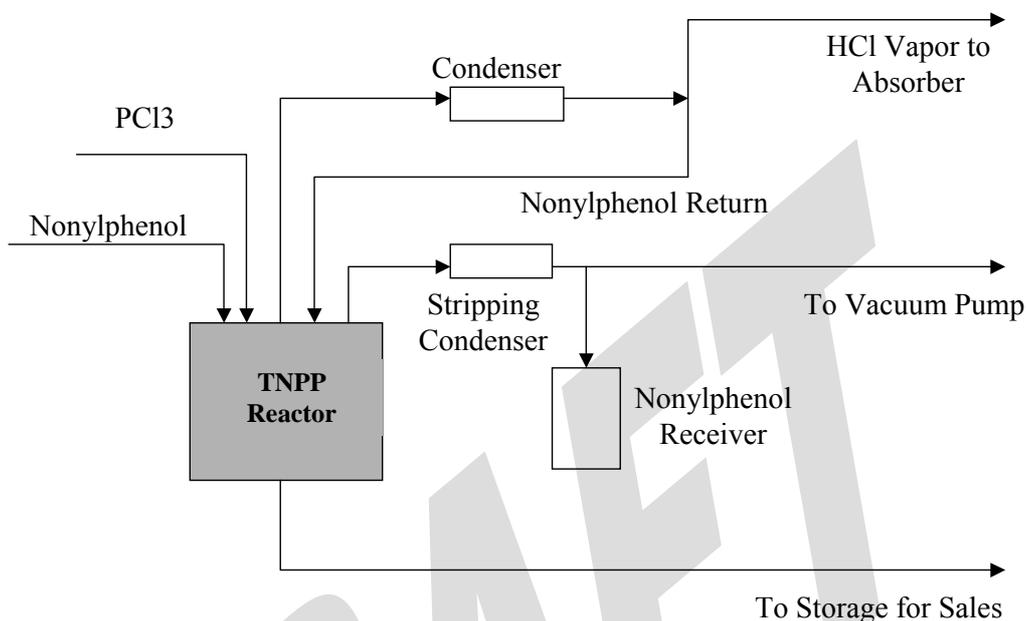


Figure 2-1: Process overview of tris(nonylphenyl)phosphite (TNPP) production

2.1.2 Production capacity

European and North American TNPP producers are organised under the Alkylphenols and Ethoxylates Research Council (APERC), a not-for-profit trade association, whose members have commercial interest in nonylphenol, octylphenol, and derivatives produced from these compounds. Information on production and imports of TNPP in Europe were provided by APERC TNPP Consortium. Hardly any individual volume was provided for each producer/importer.

Three facilities are currently producing TNPP in Europe. A fourth facility ceased TNPP production in 2001. Between 1990 and 1997, the production + import volumes were around 5,000 – 10,000 t/year.

Information is available on the combined estimate of TNPP produced within Europe and imported into Europe over the last three years:

- 1999 – approximately 5,565 tonnes
- 2000 – approximately 5,700 tonnes
- 2001 – approximately 6,800 tonnes

As this information is provided by the APERC TNPP Consortium, it cannot be excluded that these volumes do not take into account shipments of product from producers in other parts of the world than Europe and North America. However, according to the APERC TNPP Consortium, the quantity of TNPP from non-TNPP Consortium companies are not expected to be significant.

European production plants have also reported their production volumes for the year 2001. Imported volume for the same year is also available. Consequently, a total volume in Europe of 8,000 t. calculated with all 2001 data will be used in this report.

2.2 USES

2.2.1 Introduction

TNPP is used as a stabiliser in the processing of various plastic and rubber products. They are used with hindered phenolic antioxidants in plastic food packaging. In the stabilisation process, TNPP is gradually oxidised and nonylphenol is released (Building Research Establishment Ltd., 2001).

TNPP is also used as a secondary antioxidant in polymer formulations (Ullmann, 1985).

About 25 to 35 facilities are processing TNPP in Europe. Their consumption ranges from a few tonnes to over 400 tonnes/year.

An estimate of the breakdown of TNPP uses was developed based on an informal survey of North American and European manufacturers. Quantitative breakdown of TNPP uses are given in Table 2.1. The information pertains to sales of TNPP in 1999. It is expected that the breakdown of uses from the 1999 sales statistics is typical for the current year. Corresponding volumes are calculated using the total tonnage of 8,000 t.

Table 2-1: Typical quantitative breakdown of TNPP Uses

	Percentage of tonnage	Volume (tonnes)	Industrial Category / Use Category
Polyvinylchloride (PVC) film	35%	2,800	IC 11 / UC 49
Polyolefins linear low density polyethylene (LLDPE)	15%	1,200	IC 11 / UC 49
High density polyethylene (HDPE)	10%	800	IC 11 / UC 49
Rubber	37%	2,960	IC 11 / UC 49
Other/Unknown	3%	240	IC 55 / UC 0
TOTAL	100%	8,000	

In the SPIN Database (Substances in Preparations in Nordic Countries), the following industrial uses are described:

Table 2-2: Industrial uses of TNPP in the Nordic Countries (in Tonnes)

	1999 ¹	2000 ²	2001 ³
Manufacture of chemicals and chemical products	156	27	< 0.1
Manufacture of rubber and plastic products	38	105	n. i.
Manufacture of furniture; manufacturing n.e.c.	n. i.	0.4	0.1
Manufacture of fabricated metal products, except machinery and equipment	n. i.	0.2	0.1
Construction	n. i.	0.2	0.1
Manufacture of wood and products of wood and cork, except furniture; manufacture of articles of straw and plaiting materials	< 0.1	< 0.1	0.1
Total	194	132.8	0.4

n. i.: not indicated

²: Information was available for Sweden, Denmark and Norway¹: Information was available for Sweden only³: Information was available for Denmark and Norway.

TNPP is also mentioned in the following industrial categories: publishing, printing and reproduction of recorded media / sale, maintenance and repair of motor vehicles and motorcycles; retail sale of automotive fuel / manufacture of other transport equipment n.e.c. However, the volumes used in such industries could be considered as negligible (> 0.1 t/y in each country).

Besides, the following use pattern is described in the SPIN database:

Table 2-3: Use pattern of TNPP in the Nordic Countries (in Tonnes)

	1999 ¹	2000 ²	2001 ³
Stabilizers	118	120	n.i.
Intermediates	-	1	n. i.
Others	1	1	n. i.
Adhesives, binding agents	n. i.	0.5	< 0.1
Paints, lacquers and varnishes	< 0.1	0.3	< 0.1
Fillers	< 0.1	> 0.1	0.2
Total	119	122.8	0.2

n.i.: not indicated

¹: Information was available for Sweden only²: Information was available for Sweden, Denmark and Norway³: Information was available for Denmark and Norway.

TNPP is also mentioned in the following use categories: lubricants and additives / reprographic agents. However, the volumes used in such applications could be considered as negligible (> 0.1 t/y in each country).

From these tables, it could be stated that TNPP is mainly used as a stabiliser for the manufacture of rubbers and plastic products. The breakdown of TNPP uses described in Table will be used in this risk assessment.

2.2.1.1 Industrial use

Formulation and processing steps are necessary to manufacture plastic and rubber products. Formulation could be defined as the stage where TNPP is combined in a process of blending and mixing into a polymer or into another material while during the processing step, the TNPP containing material is formed. It is not known to what extent formulation and processing may occur at the same site. In the rubber industry, these two steps can often not be viewed separately (E.C., 2003, Emission Scenario Document for IC 15: others: rubber industry).

Therefore, as a worst assumption, formulation and processing stages will be assumed to occur at one site for every uses.

Without any specific information, it could be considered that TNPP is used for polymer processing, in the sub-category “processing of thermoplastics” as a processing aid. This categorisation will be used in the risk assessment for the determination of the default releases factors.

Besides, for plastic and rubber products, stages of private use and recovery may be considered. However, no specific information is available on the possible releases of TNPP during these stages.

All calculations will be performed using EUSES default parameters and, when available, emission factors issued from the emission scenario document on plastics additives (OECD, 2004).

2.2.1.2 Production of Polyvinylchloride (PVC) film

PVC containing TNPP may be used in many products like shower curtains, floorings and wall coverings.

2.2.1.3 Production of Polyolefins linear low density polyethylene (LLDPE)

LLDPE films containing TNPP are used for the manufacture of bags and food packaging. Many national regulations are covering the use of TNPP in food contact materials (Table 2-4

Table 2-4: Global food contact regulations specific to TNPP

Country	Regulation
USA	Food and Drug Administration (FDA) – 21 CFR Part 178.2010
Japan	Self-restrictive Requirements on Food-Contact Articles Japan, Hygienic Olefin and Styrene Plastics Association (JHOSPA) (March 1996), Section A4-2, maximum 1.2%
European Union	Plastics Directive 2002/72/EC, pm/ref. No. 74400, specific migration limit 30 mg/kg
Germany	BfR Recommendation VI, maximum 2.0% total of all stabilisers BGA: maximum 6% in plastics
Netherlands	Food Packaging and Utensils Decree of 01.10.1979 as amended Chapter 1
France	Brochure 1227 (Avril 1990) maximum 1.0%
Italy	Min. Decree of 21.03.1973 maximum 0.3% Min. Decree of 0.04.1985
Spain	Royal Decree 125/1982 of 30.04.1982 Resolution of 4.11.1982
Belgium	Royal Decree of 11.05.1992, specific migration limit 30 mg/kg
United Kingdom	BIBRA/BBF Code of Practice (1991) Rec. No. C.159, maximum 1.0%

2.2.1.4 Production of High density polyethylene (HDPE)

HDPE containing TNPP is used in the manufacture of many products like blow-molded plastic drums or outer wrapping (film) of cigarette boxes or tea boxes.

2.2.1.5 Production of rubber

Rubber containing TNPP are used for example in tires and shoes soles.

2.2.2 Other applications

TNPP is used in other applications than plastic and rubber productions. Using the information provided in the SPIN database, it could be supposed that these other applications include the use of TNPP in publishing, printing and reproduction activities, in the manufacture of products of wood, of fabricated metal products, of furniture and in the construction activities. However, no more specific information is available.

2.2.2.1 Use of end-products

Shower curtains, flooring and wall coverings, bags and food packaging, blow-molded plastic drums, outer wrapping films, tires and shoes soles are examples of plastic and rubber end-products containing TNPP. For all these products, both private and professional end-uses may happen. As a worst case, private use will be considered for all uses in the EUSES program (E.C., 2004b). However, it could be expected that TNPP or NP releases due to the use of end-products are negligible.

2.2.2.2 Recovery and disposal

No information on recovery has been submitted. In view of the end-products containing TNPP that are manufactured, it could be assumed that products containing TNPP may be either recycled into new products, disposed in landfill or incinerated. Therefore, this stage could be considered in the EUSES calculation (E.C., 2004a). However, no default value is actually available for this stage in version 2.0 of the software.

2.3 TRENDS

Releases of TNPP and or NP (nonylphenol) to the environment occur during production, transport, storage, formulation and processing of plastic and rubber products. In addition, releases may also take place through the uses of the end-products. Finally, waste disposal of the end-products may also release TNPP or NP into the environment.

The different industry categories (IC), use categories (UC) and main categories (MC) used in the EUSES calculations are described in Table 2-5

Table 2-5: Industrial Categories (IC), Use Categories (UC) and Main categories (MC) used in EUSES calculations

Life cycle stages		IC	UC	MC	A-Table	B-Table
Production		11	49	I b	A 1.1	B 1.4
PVC films (2,800 t)	Formulation	11	49	III	A 2.1	B 2.3
	Processing	11	49	II	A 3.11	B 3.9
LLDPE films (1,200 t)	Formulation	11	49	III	A 2.1	B 2.3
	Processing	11	49	II	A 3.11	B 3.9
HDPE films (800 t)	Formulation	11	49	III	A 2.1	B 2.3
	Processing	11	49	II	A 3.11	B 3.9
Rubber (2,960 t)	Formulation	11	49	III	A 2.1	B 2.3
	Processing	11	49	II	A 3.11	B 3.9
Others (200 t)	Formulation	15	55	III	A 2.1	B 2.3
	Processing	15	55	II	A 3.16	B 3.14

For tonnage input in the B tables, regional tonnage of TNPP was set to 700 t for the uses for PVC, LLDPE and rubber (maximum reported consumption range for TNPP processing facilities). For the uses in HDPE and other uses, the regional tonnage was respectively set to 800 t and 240 t.

A default fraction of TNPP in formulation is suggested in TGD (E.C., 2003) Emission Scenario Document for rubber Industry: up to 1.5 % (wt) for processing aids used as stabilisers. However, TNPP manufacturers have submitted better approximations of this value, for different formulated products (Personal communication from TNPP consortium, 1st April 2004):

- PVC film 0.8-1.5 %
- Polyolefins 0.1-0.2 %
- Rubber 0.4-1.0 %

As a worst case, the upper limit of these intervals will be used for the exposure assessment. Then, as a worst case too, fractions of the main source and number of days are derived from Tables B using the tonnage as such for each use.

DRAFT

3 ENVIRONMENT

This part was not provided as it will be updated.

DRAFT

4 HUMAN HEALTH

4.1 HUMAN HEALTH (TOXICITY)

4.1.1 Exposure assessment

4.1.1.1 General introduction

TNPP is a viscous liquid with a very low pressure (0.058 Pa at 25°C). It is produced by two companies in Europe. It is used as a stabilizer in plastic (PVC, LLDPE, HDPE) and rubber for the manufacture of many products like :

- shower curtains, floorings, wall coverings (PVC)
- bags for food packaging (LLDPE)
- drums, outer wrapping of cigarette or tea boxes (HDPE)
- tyre and shoe soles (rubber).

According to the TNPP manufacturers, the maximum amount of TNPP in the polymers is 1.5% (0.8-1.5 % in PVC film, 0.1-0.2 % in polyolefins, 0.4-1.0 % in rubber).

Data extracted by INRS in 2004 from the French product register SEPIA showed that 16 preparations out of the 48 000 registered between 1984 and 2004 contained TNPP. These preparations are mainly resins or resin based adhesives. TNPP is always present at a low concentration (< 1 %). Use of TNPP in preparations seems to be a very minor use.

Humans may be exposed to TNPP at workplace, via consumer products and indirectly via the environment. The highest potential exposure is likely to occur during occupational exposure.

Workers are primarily exposed via inhalation and dermal routes. For consumers, the oral route via food contact materials is the most likely.

4.1.1.2 Occupational exposure

Definitions and sources

In this document, unless otherwise stated, the term exposure is used to denote external personal exposure, assessed without taking into account the attenuating effect of any personal protective equipment (PPE) which might have been worn. This definition permits the effects of controls, other than PPE, to be assessed and avoids the considerable uncertainty associated with attempting to precisely quantify the attenuation of exposure brought about by the proper use of PPE. Furthermore, inappropriate use of gloves may even increase dermal uptake.

The estimates generated in this exposure assessment are considered to be worst-case estimates, as they describe high-end or maximum exposures in feasible but not unrealistic situations. They are not intended to account for extreme or unusual use scenarios. The majority of exposures are expected to be well below these estimates.

Since no measured exposure data are available, the assessment of inhalation and dermal exposure is based on model estimates according to the EASE model (Estimation and Assessment of Substance Exposure, version 2). This model is a general purpose predictive model for workplace exposure assessments. It is an electronic, knowledge based, expert system which is used where measured exposure data is limited or not available. It is in widespread use across the European Union for the occupational exposure assessment of new and existing substances.

All models are based upon assumptions. Their outputs are at best approximate and may be wrong. EASE is only intended to give generalised exposure data; it predicts inhalation exposure as ranges for concentrations for continuous exposure at the process under consideration. Dermal exposure is provided by EASE as the quantity of a product adhering to the skin due to a task.

Core exposure information submitted by industry for this assessment was very limited. Therefore most of the parameters chosen to characterise exposure are assumptions based on the knowledge of the assessor about the general circumstances of exposure in the relevant activities. The outcome should be regarded then as a very rough exposure assessment. Given the toxicity profile of TNPP, there is no need to allocate additional effort to generating a detailed assessment of workplace exposure at this initial stage. If there are reasonable grounds for concern for human health at a later stage, the assessment will have to be refined.

Routes of exposure and relevant scenarios

The major occupational routes of exposure to TNPP are inhalation and skin contact. Assuming proper hygiene measures are applied, oral exposure would normally not occur in the workplace.

Exposure may occur during manufacture of TNPP and during handling and further processing in the polymer industry. The following scenarios are regarded as relevant :

- scenario 1 : manufacture of TNPP
- scenario 2 : manufacture of products containing TNPP
- scenario 3 : use of preparations containing TNPP

The stabiliser TNPP is physically bound within the polymer matrix and therefore it could migrate to the surface especially at high temperatures. Release of TNPP from plastic or rubber end products may be a potential way of exposure but due to the very low vapour pressure (0.058 Pa at 25°C) and the small percentage of the stabiliser in the polymers (<1.5 %) exposure to TNPP during subsequent use of products is likely to be negligible.

The number of persons exposed to TNPP is not known.

There are no occupational exposure limits for TNPP.

In the present assessment, inhalation exposure expressed in parts per million (ppm) are converted to mg/m³ using the following approximation : mg TNPP/m³ = ppm x 28.6.

4.1.1.2.1 Scenario 1 : Manufacture of TNPP

TNPP is manufactured in three sites in Europe. According the TNPP consortium (North American and European producers), the manufacture is carried out in a closed system where nonylphenol and phosphorus trichloride are added to the reactor and held at 110°C to ensure all phosphorus trichloride is consumed. After the nonylphenol excess has been removed, TNPP is pumped to a storage tank for packaging. The process is fully automated and normally controlled by 1 or 2 operators.

For the large-scale chemical industry, high standards of control at the workplaces are assumed to be practised even if the containment is breached, e.g. during filling. Exposure may occur during coupling and uncoupling of transfer lines, drumming, cleaning, maintenance, repair works and sampling. It is assumed that such activities are performed with local exhaust ventilation (LEV).

Inhalation exposure

The EASE model estimates for production an inhalation exposure in the range of 0-0.1 ppm (closed system, full containment).

If the system is breached in some activities (like maintenance, sampling, cleaning, filling), concentrations are still in the range of 0-0.1 ppm (non-dispersive use, very low tendency to become airborne).

A full shift exposure level of 0.1 ppm (2.86 mg/m³) is taken as representing a reasonable worst case.

Dermal exposure

Due to the enclosure of the process and control measures taken to minimize skin contact (for example, during transfer to tankers), dermal exposure at the plant is incidental and therefore likely to be low. The main source of potential exposure is during maintenance activities.

The EASE model estimates a dermal exposure in the range of 0-0.1 mg/cm²/day (non dispersive use with direct handling and incidental contact). Assuming exposed skin surface area is 420 cm², maximum external dermal exposure would be 0-42 mg/day.

4.1.1.2.2 Scenario 2 : manufacture of products containing TNPP

According to the TNPP producers, there are 21 facilities in the EU that formulate TNPP (based on 1999 sales information).

Stabilizers are usually added to polymers during the production of powder or pellets (compounding, master batching) which are then transformed into shaped articles (semi-finished or finished products) by different process: extruding, calendaring, injection moulding.

Inhalation exposure

Highest exposure would normally occur during the transfer of the substance to the mixer at ambient temperature and during mixing or transforming at high temperatures. Temperatures

are assumed to be between 100 and 200°C. Exposure to dust during the handling of polymer powders or pellets could also be a source of exposure.

Activities at ambient temperature (transfer of the substance)

The EASE model estimates an inhalation exposure in the range of 0-0.1 ppm (non dispersive, very low tendency to become airborne).

The task is assumed to typically take 2 hours (and is probably not carried out daily), therefore a full shift exposure level of 0.025 ppm (0.72 mg/m³) is taken as a maximum.

Mixing or transforming at high temperatures

It is assumed that these processes are mainly performed using closed systems but the cooling of worked articles, pellets or sheets could be done in open systems. Taking into account the vapour pressure of TNPP at 200°C is about 373 Pa (category “low tendency to become airborne” regarding the volatility), the EASE model estimates an inhalation exposure in the range of 0.5-1 ppm (non dispersive use, with LEV) or 10-20 ppm (non dispersive use, direct handling with dilution ventilation). Lower exposure levels are obtained when it is assumed a use process consisting of inclusion into the polymer matrix : they become in the range of 0.5-1 ppm (non dispersive use, with LEV) or 3-5 ppm (non dispersive use, direct handling with dilution ventilation).

The model overestimates exposure levels, particularly because of non-consideration of the content of TNPP in the polymer. A simple approach based on a reduction of the highest estimated exposure (20 ppm) by a factor equivalent to the TNPP concentration (1.5 %) leads to a maximum exposure of 0.3 ppm (8.58 mg/m³). Duration and frequency of exposure are assumed to be full shift and daily.

Exposure to dust

Measurements during loading of powder are available but it is not appropriate to consider the results as analogous/surrogate. Studies were generally performed with inorganic powder whereas the dust for this scenario comes from plastic or rubber. Therefore the EASE model is used to for estimation of the exposure. EASE estimates dust exposure in the range of 2-5 mg/m³ (dry manipulation with LEV) or 5-50 mg/m³ (dry manipulation, without LEV). Considering a concentration of TNPP in the polymer of 1.5%, the inhalation exposure to dust amounts to 0.03-0.075 mg/m³ with LEV and 0.075-0.75 mg/m³ without LEV. As the task (bag emptying) is assumed to typically take 2 hours, a maximum inhalation exposure of 0.19 mg/m³, full shift, is predicted.

Conclusion

An inhalation exposure level of 8.58 mg/m³ (related to mixing or transforming activities at high temperatures) will be considered as the worst case exposure during manufacture of products containing TNPP.

Dermal exposure

The highest dermal exposure may occur during transfer of the substance. Afterwards TNPP is enclosed in the polymer matrix and exposure is negligible.

The EASE model estimates a dermal exposure in the range of 0.1-1 mg/cm²/day (non dispersive use with direct handling and intermittent contact). Assuming exposed skin surface

area is 420 cm² (palms of hands), maximum external dermal exposure would be 42-420 mg/day.

4.1.1.2.3 Scenario 3 : use of preparations containing TNPP

TNPP may be a component of resin based preparations. It is a minor use of TNPP and when it is present, its concentration in the formulation is very low (<1 %). For complete assessment, the use of adhesives containing TNPP is considered in this scenario.

Inhalation exposure

Activities at ambient temperature

Taking into account the low vapour pressure of TNPP and the low concentration in the preparations, inhalation exposure is likely to be negligible.

Activities at high temperatures (e.g. use of hotmelt adhesives)

Taking into account the vapour pressure of TNPP at 200°C is about 373 Pa (low tendency to become airborne), the EASE model estimates an inhalation exposure in the range of 0.5-1 ppm (non dispersive use, with LEV) or 10-20 ppm (non dispersive use, direct handling with dilution ventilation).

The model overestimates exposure levels, particularly because of non-consideration of the content of TNPP in the preparation. A simple approach based on a reduction of the highest estimated exposure (20 ppm) by a factor equivalent to the TNPP concentration (1 %) leads to a maximum exposure of 0.20 ppm (5.72 mg/m³). Duration and frequency of exposure are assumed to be full shift and daily.

Dermal exposure

The EASE model estimates a dermal exposure in the range of 0.1-1 mg/cm²/day (non dispersive use with direct handling and intermittent contact). Taking into account the concentration of TNPP in the preparation (1%) and assuming exposed skin surface area is 420 cm² (palms of hands), maximum external dermal exposure would be 0.42-4.2 mg/day.

4.1.1.2.4 Occupational exposure summary

Table 4-1: Summary of reasonable worst case exposures

Scenario	8-hour TWA inhalation (mg/m ³)	Dermal (mg/day)
1 - Manufacture	2.86	0-42
2 – Manufacture of products	8.58	42 - 420
3 – Use of preparations	5.72	0.42 - 4.2

4.1.1.3 Consumer exposure

4.1.1.3.1 Introduction

Trisnonylphenylphosphite (TNPP) is used as an antioxidant to stabilise polymers against degradation by ultraviolet light. TNPP is used in plastics such as bathroom curtains, tyres, shoe soles and food packaging. In order to assess the consumer exposure, it is necessary to identify the potential ways of human exposure resulted from inhalation, ingestion or skin contact. First, there is no consumer exposure due to tyres. Besides skin contact may happen with bathroom curtains and shoe soles, but remain short or rather occasional. Moreover TNPP is considered to be a slight irritant to the skin. Thus dermal exposure is not considered to be significant. Furthermore, it seems useful to mention that TNPP might be used in plastic medical devices. Such PVC disposable devices, used for blood transfusion, hemodialysis and peritoneal dialysis for example, are particularly studied as a human potential exposure to phthalates, DEHP (diethylhexylphthalate) in particular (FDA, 2001). On the one hand, the studies conclude that such an exposure has more benefits than adverse health effects as far as public health is concerned (INSPQ, 2004). On the other hand, the results from the alkylphenol work group (AWG, 1998) indicate that no detectable amount of TNPP was shown to migrate from PVC-films in any test conditions. That's why risk assessment for the potential exposure to TNPP from plastic medical devices is not performed. Finally consumer exposure due to food-contact materials is the only source of potential exposure which may be important and that worth being particularly studied. It can also be mentioned as preliminary note that a maximum migration TNPP value into food has been proposed by the European Health & Consumer Protection Directorate-General (DG-SANCO, 2003). Since TNPP doesn't need to be classified on the basis of acute toxicity (see chapter 4.1.2.2), only chronic toxicity due to repeated oral exposure from food-contact materials will be characterised.

TNPP may be partly hydrolysed (acid hydrolysis) and gives nonylphenol. In order to be protective, the TNPP amount in food-contact plastics will be overestimated in so far as the TNPP degradation won't be considered.

4.1.1.3.2 Potential exposure from migration of TNPP from food contact materials

TNPP use and amount in plastics

According Howe *et al.* (Howe, 2001), TNPP is used in three types of food-contact polymers : PVC-films, polyolefins (linear low density polyethylene – LLDPE or ethylene vinyl acetate copolymers – EVA) and rubber. Previously a distinction was often made between “rubber” and “rubber modified polystyrene”. Actually a distinction doesn't seem to be needed and the two categories will be considered as one “rubber” category, i.e. high impact polystyrene – HIPS. Various grades of TNPP are used in these polymers as shown in Table 4-2.

Table 4-2: Use of trisnonylphenylphosphite in food-packaging polymers

Polymer type containing TNPP	TNPP concentration (mass %)
PVC Film	0.8 – 1.5 %
LLDPE	0.1 – 0.2 %
EVA	0.05%
Rubber / HIPS	0.4 – 1.0 %

Calculated exposure

The dietary consumption of TNPP depends on :

- the potential level in food;
- the fraction of an individual's diet likely to contact food materials containing TNPP;
- the total weight of food daily consumed by an individual.

The dietary exposure is calculated using the American Food and Drug Administration (FDA) model which uses consumption and food-type distribution factors. Since no European data are available, numerical values for the different factors are those from FDA, derived from simulated food-contact use (with food-stimulating solvents) (FDA, 2002).

When the accurate different food items in contact with the packaging are well identified, the daily dietary concentration in food is the following:

$$\text{Dietary concentration} = P \times \sum_i (CF_i \times M_i)$$

M_i is the migration value of TNPP in the considered food-type. This migration value M_i is given by the experimental exposure of the food-contact material to a given food simulant (under the time and temperature exposure) representing the food-type. This food-type is classified as aqueous (aq), acidic (ac), alcoholic (al) or fatty (f).

The consumption factor CF_i represents the fraction of the food-type (which is daily consumed) likely to contact the food packaging.

On the opposite, when food items can not be precisely distinguished, the whole diet is divided into aqueous (aq), acidic (ac), alcoholic (al) or fatty (f) food items. The daily dietary concentration in food is thus the following:

$$\text{Dietary concentration} = P \times CF \times (M_{aq} \times f_{aq} + M_{ac} \times f_{ac} + M_{al} \times f_{al} + M_f \times f_f)$$

M_{type} is the migration value of TNPP in the considered *food-type*. This migration value M_{type} is given by the experimental exposure of the food-contact material to a given food simulant (under the time and temperature exposure) representing the food-type, which can be aqueous (aq), acidic (ac), alcoholic (al) or fatty (f). "10% ethanol" or "90% ethanol" indicate thus food simulants.

The distribution factor f_{type} is the fraction of food of each *type* that will contact the material.

The consumption factor CF represents the fraction of the whole daily diet likely to contact the food packaging.

In both equations, P is the percentage of the considered food-packaging containing TNPP.

The estimated daily intake (EDI) for each type of packaging is determined by multiplying the total weight of food consumed by an individual per day (default 3000 g/day, solids and liquids) by the dietary concentration of TNPP in the studied packaging.

Finally the total estimated daily intake (TEDI) represents the sum of each EDI respectively due to PVC films, LLDPE, EVA and HIPS :

$$\text{TEDI} = \text{EDI}_{\text{PVC}} + \text{EDI}_{\text{LLDPE}} + \text{EDI}_{\text{EVA}} + \text{EDI}_{\text{HIPS}}$$

Estimated daily intake due to PVC

All the food-contact PVC-films contain TNPP. However, the results from the alkylphenol work group (AWG, 1998) indicate that no detectable amount of TNPP was shown to migrate from PVC-films in any test conditions. PVC dietary concentration $\equiv 0$ and $\text{EDI}_{\text{PVC}} \equiv 0$

Estimated daily intake due to LLDPE

50% of food-contact LLDPE contains TNPP. Since each food type is precisely identified (frozen food, bag-in-box items, ...) with their accurate CFs, the dietary concentration is :
Dietary concentration = $P \times \sum_i (\text{CF}_i \times M_i)$.

Table 4-3: Potential TNPP exposure from LLDPE

Application		P	CF	M (ppm)	Exposure (ppb)
Film	Produce	0.5	0.04	ND ^a	-
	Frozen	0.5	0.001	ND ^a	-
	Meat/Poultry	0.5	0.002	1.53 ^b	1.53
	Dry	0.5	0.01	ND ^a	-
	Bag-in-box	0.5	0.006	ND ^a	-
	Snack	0.5	0.002	1.53 ^b	1.53
Films/Coatings		0.5	0.0002	1.53 ^b	0.15
Lids/Tubs	Aqueous	0.5	0.0027	ND ^c	-
	Fatty	0.5	0.0013	2.64 ^d	1.72
LLDPE dietary concentration					5.0 ppb
EDI _{LLDPE}					0.015 mg/day

ND ^a: “10% ethanol” data from films have to be used for film uses involving produce, frozen, dry, and bag-in-box. The results shown in Table 12 from (AWG, 1998) indicate that no detectable amount of TNPP was shown to migrate to 10% ethanol under any of the test conditions used.

^b: “95% ethanol” data from films have to be used for film uses involving meat, poultry, and snack, and for films/coatings applications (Table 13, AWG, 1998). The maximum value obtained (migration after 4-day test) is used. 4-day and 10-day migration results are not significantly different.

ND ^c: “10% ethanol” data from plaques (condition of use E = 40°C for 10 days) have to be used for the aqueous lids/tubs applications. No TNPP was shown to migrate (Table 12, AWG, 1998).

^d: “95% ethanol” data from plaques (condition of use E = 40°C for 10 days) have to be used for the fatty lids/tubs applications. The maximum value obtained (migration after 4-day test) is used (Table 13, AWG, 1998).

Estimated daily intake due to EVA

25% of food-contact EVA contains TNPP.

According to the alkylphenol work group, “10% ethanol” data from LLDPE films have to be used for the aqueous food applications and “95% ethanol” data from LLDPE films for the fatty food applications. As far as the aqueous applications are concerned, no TNPP was shown to migrate (Table 12, AWG, 1998). For the fatty applications, the maximum value obtained (migration after 4-day test) is used i.e. 1.53 ppm (4-day and 10-day migration results are not significantly different) (Table 13, AWG, 1998). Since the use level of TNPP in EVA films (500 ppm) is lower than the 1200 ppm TNPP level in the films use for the migration tests and since migration is directly proportional to the concentration, a factor of 42% is used (500/1200). That’s why the TNPP migration level that is used is 0.64 ppm.

$$\text{EVA dietary concentration} = P \times CF \times M_f \times f_f = 0.25 \times 0.04 \times 0.64 \times 0.45 = 2.9 \text{ ppb}$$

$$\text{EDI}_{\text{EVA}} = 0.0087 \text{ mg/day}$$

Estimated daily intake due to HIPS

All the food-contact HIPS-packaging contain TNPP. Since each food type is identified (yoghurts, cheese, ...) with their accurate CFs, the dietary concentration is : Dietary concentration = $P \times \sum_i (CF_i \times M_i)$.

Since the use level of TNPP in HIPS films (10000 ppm max. value) is higher than the 1200 ppm TNPP level in the films use for the migration tests and since migration is directly proportional to the concentration, a factor of 833% is used (10000/1200). M values in Table 4-4 take into account this 833% factor.

Table 4-4: Potential TNPP exposure from HIPS

Application		P	CF	M (ppm)	Exposure (ppb)	
Packaging	Yoghurt cups (aq)	1	0.0036	ND ^a	-	
	Cheese/Cream (aq)	1	0.0036	ND ^a	-	
	Aseptic/Blow moulded (aq)	1	0.0009	ND ^a	-	
Disposable	Fatty	4°C	1	0.0001	0.48 ^b	0.048
		24°C	1	0.0001	0.48 ^b	0.048
		54°C	1	0.0003	11.3 ^c	3.4
	Aqueous	4°C	1	0.0108	ND ^a	-
		24°C	1	0.0188	ND ^a	-
		54°C	1	0.0016	ND ^a	-
	Alcoholic	1	0.0015	ND ^a	-	
HIPS dietary concentrations					3.5 ppb	
EDI_{HIPS}					0.010 mg/day	

ND^a: 10% ethanol data from LLDPE plaques have to be used for the aqueous and alcoholic applications. The results shown in Table 12 from (AWG, 1998) indicate that no detectable amount of TNPP was shown to migrate to 10% ethanol under any of the test conditions used.

^b: 95% ethanol data from LLDPE plaques have to be used for the fatty applications according to the alkylphenol work group (AWG, 1998). Moreover, the disposable uses involve contact with food for 1 or 2 hour max. The closest time frame to this length of time used in the testing is 2 hours. Thus, the 2-hour data have been considered for the HIPS disposables. Finally for the applications at 4°C and 24°C, results in test condition of use E (= 40°C for 10 days) are used.

^c: ^d But for the applications at 54°C, an interpolated M value corresponding to the value between 40°C and 100 °C (condition of use B = 100°C for 2 h, followed by 40°C for 238 h) is used considering that there is a linear relationship between migration and the inverse of temperature (in Kelvin). Thus, the migration value (before correction with the 833% factor) is 1.36 ppm (AWG, 1998).

4.1.1.3.3 Consumer exposure summary

The overall potential dietary exposure, or total estimated daily intake (TEDI), to TNPP from the use in food-contact packaging is the sum of the above EDI values:

$$\text{TEDI} = \text{EDI}_{\text{PVC}} + \text{EDI}_{\text{LLDPE}} + \text{EDI}_{\text{EVA}} + \text{EDI}_{\text{HIPS}}$$

$$\text{TEDI} = 0 + 0.015 + 0.0087 + 0.010$$

$$\text{TEDI} = 0.0337 \text{ mg/day}$$

4.1.1.4 Indirect exposure via the environment

This section was not provided as it will be updated in the next version of the environmental risk assessment.

4.1.2 Effects assessment : hazard identification and dose (concentration)-response (effect) assessment

4.1.2.1 Toxicokinetics, metabolism and distribution

No specific toxicokinetic study was conducted with trisnonylphenyl phosphite (TNPP).

However qualitative information can be derived from the physico-chemical properties of the substance. Considering the relatively high molecular weight of the molecule (MW = 689 g.mol⁻¹), its extremely low water solubility and a very high Log P_{ow}, the absorption of TNPP by the gastro-intestinal tract is expected to be limited.

The vapor pressure of the liquid substance (physical state at 20°C and 101,3 kPa) is very low . Therefore, inhalative exposure can be anticipated only as liquid aerosol.

The molecular weight (> 500) of TNPP, its water solubility (< 1 mg/l) and its Log Pow (> 6) are in favour of a very limited absorption following dermal exposure.

One subject of possible concern is the metabolisation of TNPP into nonylphenol (NP) :

The hydrolysis of phosphite esters is well known, and can be catalysed by either acid or base. The reaction is stepwise and is shown below :



where ArOH = nonylphenol (molecular weight = 220.34 g.mol⁻¹)

(ArO)₃P = TNPP (molecular weight = 689 g.mol⁻¹)

(HO)₂P(O)H = phosphorous acid

With excess water, the hydrolysis goes to completion, yielding 3 molecules of NP and 1 molecule of phosphorous acid for each molecule of TNPP, that is 100 units (mass units) of TNPP will lead to 96 units of NP.

As mentioned above, physico-chemical properties of TNPP suggest that only limited absorption is likely to occur. However, some hydrolysis of TNPP is known to occur under conditions similar to those in the gut.

Hydrolysis of approximately 20% of TNPP to NP in the gastro-intestinal tract is possible based on the following : the hydrolysis t_{1/2} of TNPP to NP at pH 4, 7, and 9 was similar at approximately 13 hours (Reimer, 2001). Assuming an average residency time of TNPP in the upper portion of the gastro-intestinal tract of 4 hours and no significant hydrolysis/absorption in the lower gastro-intestinal tract (consistent with the lack of enterohepatic circulation) (Green, 2001), approximately 20% of the TNPP would be hydrolysed. Thus, a dose of 1000 mg/kg/day TNPP could result in a dose of NP almost up to 200 mg/kg/day.

NP is rapidly absorbed in male and female rats and thereafter is rapidly eliminated, mainly via the bile and feces. A study on rats indicated that saturation of metabolism at a 100 mg/kg dose

occurred in female rats as evidenced by the identification of unmetabolised NP in urine and bile, not seen at a dose of 10 mg/kg. Evaluation of the metabolism of NP in males indicated a number of additional metabolites not observed for females and no indication of metabolic saturation from the 10 to the 100 mg/kg dosage (Green, 2001).

Based on these qualitative toxicokinetic data, default values were chosen for oral, dermal and inhalative absorption :

Oral absorption : as indicated above, the absorption of TNPP by the gastro-intestinal tract is expected to be limited. However no quantitative value is available, then as a worst case assumption for oral route, a default value of 50% is chosen.

Dermal absorption : a default factor of 10% is used as $MW > 500$ and $\text{Log } P_{ow}$ is higher than 4.

Inhalative exposure :. Absorption mechanisms via mucous membranes are expected to be the same by oral and inhalation route, thus a default value of 50% is chosen as a worst case assumption.

4.1.2.2 Acute toxicity

Only data on animals are available.

4.1.2.2.1 Oral

The acute oral toxicity of TNPP has been investigated in three animal studies of different quality.

- The most informative study related to acute oral toxicity was conducted by the Food and Drug Research Laboratories for Naugatuck Chemical Corporation in 1957. The report was scientifically acceptable, although the study was not conducted in compliance with the Good Laboratory Practice (GLP) and the international standardisation of testing methods.

Five groups of ten adult albino rats (5 males and 5 females) were given graded doses of a 50 per cent solution of TNPP (purity not specified) in cottonseed oil. The doses (8.19 - 11.32 - 16.38 - 22.62 and 32.72 gram/kg bw.) were administered by stomach tube.

Following dosage, the rats were observed for appearance, behaviour, bodyweight and mortality for a 14-day period. Rats that died, as well as survivors sacrificed at the end of the experiment, were examined for evidence of gross pathology.

All rats showed evidence of abdominal pain and catharsis after dosage. Highest doses (11.32 gram/kg and above) resulted in urinary incontinence and prostration. Gross pathological findings included hemorrhagic lesions in the gastric mucosa and/or duodenum in a few rats that died, and hemorrhagic lungs. According to the authors, the incidence and severity of the abnormalities at the former site being greater at the higher dose levels. However this assertion couldn't be checked because no table of results related to the gross pathological findings was available.

Mortality resulted at scattered intervals over the first five days Table 4-5 . However, growth of the survivors was essentially normal. The Lethal Dose 50 (LD_{50}) was computed according to the method of Miller and Tainter (1944) and was calculated to be 19.5 ± 3.3 gram/kg bw.

Table 4-5: Acute oral toxicity of TNPP

Dose (g/kg)	8.19	11.32	16.38	22.62	32.72
Number of death	0	3	4	7	7

● Another study was conducted by Hill Top Research in 1965. The purpose of this study was to evaluate and compare the acute oral toxicity of four samples of chemicals in rats (two samples of TNPP and two samples of another chemical). This study was not conducted in compliance with the GLP and the international standardisation of testing methods. The samples were received from Argus Chemical Corporation. For the purposes of this study, the purity of each sample was considered to be 100% and no correction was made for possible impurities. The results were incompletely provided as the page of results related to the second sample of TNPP is missing and all the paper files for this study were since discarded.

Graded doses up to 10 ml/kg of TNPP (equivalent to 9.8 g/kg, based on a relative density of 0.98 g.cm⁻³ at 20°C) were administered orally by stomach tube to six groups of five male albino rats, Holtzman strain. Each TNPP sample was administered as a 10% or 50% volume/volume solution in corn (Mazola) oil at dosage levels of 0.215 - 0.464 - 1.00 - 2.15 - 4.64 and 10.0 ml/kg bw (equivalent to 0.21 – 0.45 – 0.98 – 2.11 – 4.54 and 9.80 g/kg bw). Larger doses could not be administered without exceeding the capacity of the rat stomach.

All animals were observed closely for gross signs of toxicity and mortality at frequent intervals during the day of dosage, and at least once daily thereafter for a total of 14 days. Gross autopsies were performed on the animals that died. At the end of the 14-day observation period, the surviving rats were weighed, sacrificed and gross autopsies were performed.

For sample one, there was no mortality at any dosage level tested. The acute oral LD₅₀ of TNPP for male albino rats was therefore established to be greater than 10.0 ml/kg bw (9.8 g/kg bw).

All the rats exhibited normal appearance and behaviour during the observation period. The average body weight gain for each group of rats was within the normal range of values for rats of the sex, age and strain used in this study. At gross autopsy, the organs of all animals appeared grossly within normal limits.

There was no information related to sample two, except for the summary that mentioned that LD₅₀ was greater than 10.0 ml/kg bw too (9.80 g/kg bw).

● A study conducted by Majlathova (1981) was related to the evaluation of aralkyl phenylphosphite antioxidants by an acute peroral experiment on mice and rats and by epicutaneous and conjunctival test on rabbits. The publication available related to this study was written in Slovakian but it contained an abstract written in English by the authors. The abstract states that the starting LD₅₀ concentration of the TNPP product tested was greater than 10 grams/kg bw but that the storage (time) makes it become more toxic. Yet 5 gram/kg bw do not affect the health condition of the animals.

4.1.2.2.2 Inhalation

No study was found.

4.1.2.2.3 Dermal

- Acute dermal toxicity was studied in a recent and well-conducted study, following OECD guideline 402 (Tay, 2001a).

TNPP (purity not indicated) was evaluated for its potential to produce systemic toxicity or death after a single topical 24-hour application to the skin of albino rabbits at a dose of 2000 mg/kg (limit test). Five male and five female New Zealand White rabbits were used for the test. The test substance was introduced under gauze patches, two single layers thick, and applied directly to the skin of the body surface (approximately 10%) of each of ten animals. At the completion of the exposure period, the skin was gently wiped to remove any test substance still remaining. The animals were observed frequently during the first day, and then a careful clinical examination was made at least once a day. The animals were also observed for signs of erythema and oedema after the exposure period according to the Draize scale for scoring skin reactions. Animals were weighed at day 0 (prior to dose administration), day 7 and day 14. A gross necropsy was performed on all animals whether found dead or sacrificed at the end of the study, on the 14th day.

All animals gained weight during the post treatment-period, except one male rabbit (weight loss = 0.02 kg). In all rabbits, no other sign of systemic toxicity was evident during the course of the study and no animal died. At necropsy, there was no abnormality or lesion noted. No erythema or oedema was observed at any of the test sites.

The LD₅₀ in rabbits was then found to be greater than 2000 mg/kg bw.

- Another recent dermal acute toxicity study, performed in rats by Ciba-Geigy (1992), confirms these results. The study was conducted in compliance GLP and following the OECD Test Guideline 402.

The purity of the test article was > 94%.

Ten young adult albino rats of both sexes (5 males and 5 females) were exposed to the dose of 2000 mg/kg bw. The test article was evenly dispersed on the back of the rat (at least 10% of the body surface was shaved with an electric clipper). After 24h under semi-occlusive conditions the dressing was removed and the skin was cleaned with lukewarm water.

Animals were weighed at day 0 (immediately before application), day 7 and day 14. Mortality and symptoms were observed daily for 14 days and the animals were submitted to a gross necropsy at the end of the observation period.

All animals gained weight during the post treatment-period. No mortality occurred in this study. Piloerection and hunched posture were seen, being common symptoms in acute dermal tests. The animals recovered within 2 days. At necropsy, no deviation from normal morphology was found.

The LD₅₀ in rats of both sexes was then found to be greater than 2000 mg/kg bw.

● The Food and Drug Research Laboratories (1961) indicate that toxicological screening tests, conducted in these laboratories, demonstrated that 24-hour dermal applications in massive doses were not lethal to the rabbits. However, the report for these screening tests was not available.

4.1.2.2.4 Other

An acute intraperitoneal toxicity study was conducted in rat (Ciba-Geigy, 1983).

This study was not conducted in compliance with GLP and the international standardisation of testing methods. The sample provided by the sponsor was Irgafos TNPP (trade name) but no information was provided about its purity.

In this study, 5 male and 5 female albino rats were administered a single dose of 1000 mg/kg bw, by intraperitoneal injection. The vehicle used was distilled water containing 0.5% carboxymethylcellulose and 0.1% polysorbate 80. Animals were weighed at day 1, 7 and 14. Mortality and symptoms were observed daily for 14 days or until all symptoms have disappeared. The animals were submitted to a gross necropsy at the end of the observation period.

Dyspnoea, exophthalmus, ruffled fur and curved body position were seen, being common symptoms in acute tests. Animals recovered within 12 days. No mortality occurred during the study. At autopsy, peritoneal adhesions in the liver and spleen area were found in 8/10 animals.

The LD₅₀ in rats of both sexes was then found to be greater than 1000 mg/kg bw.

4.1.2.2.5 Summary of acute toxicity

No human data is available. In animals, TNPP has a very low acute toxicity by the oral route, with a LD₅₀ value of about 19.5 +/- 3.3 gram/kg bw for the rat. Hemorrhagic lesions in the gastro-intestinal tract and the lungs are seen in some animals, following the administration of a lethal dose. This value was used for the risk assessment. The other studies couldn't be used in the risk assessment due to shortcomings or unavailable study reports. Furthermore a LD₅₀ could not be derived from these studies as no mortality was observed at doses up to the highest doses tested (about 10 g/kg). Nevertheless, these results are in accordance with the value of 19.5 g/kg bw derived from the study from Naugatuck (1957).

The acute toxicity of TNPP by the dermal route seems to be very low too, with a LD₅₀ greater than 2000 mg/kg in rabbits. No data is available on the acute inhalation toxicity, although the non-corrosive and non-irritant nature of TNPP (see section 4.1.2.3.1 on skin irritation) may suggest that toxicity would not be enhanced following exposure by this route.

By intraperitoneal route, the LD₅₀ was found to be > 1000 mg/kg in rats.

Classification and labelling :

According to the criteria of the European Union, this chemical doesn't need to be classified on the basis of its acute toxicity.

4.1.2.3 Irritation

Only animal data are available.

4.1.2.3.1 Skin

- Acute dermal irritation and corrosion were studied in a recent and well-conducted study, following OECD guideline 404 (Tay, 2001b).

The degree of purity of the test substance was 99.3%.

TNPP was evaluated for its potential to produce skin irritation and/or corrosion after a single topical application for four hours to the intact skin of New Zealand White rabbits.

One male and two females were used for the test. A dose of 0.5 ml liquid test substance was applied to a small area (approximately 6 cm²) of skin. The test report does not indicate if the test substance was applied as pure or not, but the hypothesis of a pure substance application is the most probable. Animals were observed for signs of erythema and oedema at 60 minutes and then at 24, 48 and 72 hours after patch removal. Observations were scored according to the Draize scale for scoring skin reactions. Daily clinical observations included all toxicological and pharmacological signs. Animals were weighed at the end of the observation period.

All of the test animals exhibited a gain in body weight during the study. No overt sign of toxicity was evident in any of the animals during the course of the study. Very slight erythema was observed in three out of three rabbits following a 4-hour exposure. By the 24-hour observation point, the irritation was reversed, with no sign of erythema present at the 24-, 48- and 72-hour observations for all three rabbits. No oedema was observed at any of the observation points (see Table 4-6).

Table 4-6: Skin mean reaction scores (4-hours exposure)

Time after exposure	60 minutes		24 Hours		48 Hours		72 Hours	
	Erythema	Edema	Erythema	Edema	Erythema	Edema	Erythema	Edema
	1	0	0	0	0	0	0	0

TNPP was considered to be a very slight irritant to the skin.

- Ciba-Geigy conducted a skin irritation study in rabbits in 1981. The study was not conducted in compliance with GLP and the international standardisation of testing methods, however, the procedure used is based on the Proposed Guidelines of the United States Environmental Agency (US EPA) : "Primary dermal irritation study" (1978).

The sample provided by the sponsor was Irgafos TNPP (trade name) but no information was provided about its purity.

The test was performed on 3 male and 3 female adult New Zealand White rabbits. Before treatment, the entire back and the flank of the rabbits were shaved with electric clipper and immediately before treatment, the shaven skin on one side was slightly scarified. Gauze

patches, laden with 0.5 ml of the test material, were applied to the prepared abraded and intact skin of the rabbits. After 24h under occlusive conditions the dressings were removed and the skin reaction was appraised upon removal and during an observation period of 7 days. The grading system for skin irritation was similar to the Draize scale for scoring skin reactions.

The study report states that in 3/6 animals, the application sites showed necrosis, but it did not give any further information on this effect. In 5/6 animals the erythemas extended beyond the treated areas. Erythema and edema of intact skin were reversed within 7 days, except in abraded skin in 2/6 animals for which erythema was still moderate to severe. The calculated primary irritation index was 2.5 : TNPP was found to cause moderate irritation when applied to intact and abraded rabbit skin (see Table 4-7).

Table 4-7: Skin mean reaction scores (24-hours exposure)

Time after exposure	Erythema		Edema	
	Intact skin	Abraded skin	Intact skin	Abraded skin
24 hours	2.0	2.7	1.0	1.4
72 hours	0.7	1.7	0.0	0.7
7 days	0.2	1.2	0.0	0.3

- In New Zealand rabbits, the acute dermal toxicity study (OECD 402) conducted by Tay (2001a) (see section 4.1.2.2.3 dermal acute toxicity), a single topical 24-hour application of pure substance to the skin exhibited no signs of erythema and oedema at any of the test sites after the exposure period according to the Draize scale for scoring skin reactions.
- The Food and Drug Research Laboratories (1961) indicate that toxicological screening tests conducted in these laboratories demonstrated that TNPP was a primary irritant when applied repeatedly to the skin of rabbits, but that the effects were reversible. The report for these screening tests was not available.

4.1.2.3.2 Eye

- Acute eye irritation and corrosion were studied in a recent and well-conducted study, following OECD guideline 405 (Tay, 2001c).

The degree of purity of the test substance was 99.3%.

TNPP was evaluated for its potential to produce an irritating effect on the ocular tissue of New Zealand White rabbits. Two males and two females were used for the test.

Both eyes of each rabbit were examined for macroscopic findings and were scored before and after treatment. The grading system for ocular irritation was the one presented in OCDE test guideline 405. The eyes of the animals were screened with fluorescein stain before dosing. The left eye of each animal was treated with 0,1 ml of the test substance, the right eye remaining untreated and thus, served as a control. The eyes of the test animals were not washed prior to 24 hours following instillation of the test substance. Following the 24-hour observation, the treated eyes were not rinsed. The initial procedure was performed on one

rabbit, using a 10% dilution of the substance. As no severe effect was observed, the test was performed on three rabbits with the undiluted test substance.

Eyes were examined at 1, 24, 48 and 72 hours after treatment using the scale of grades for ocular lesions. After recording the observations at 24 hours, the eyes of all rabbits were examined with the aid of fluorescein to further characterise corneal opacity. Animals were also observed daily for clinical manifestations and were weighed at the end of the observation period.

All the animals exhibited an increase in body weight during the course of the study. No overt sign of toxicity other than the ocular effects was evident during the course of the study in any of the animals.

Eye Scores :

- No corneal opacity was observed in any of the treated eyes, at any of the observation periods.
- No fluorescein staining was observed in any of the treated eyes, at all observation points.
- The iris response was normal in all treated eyes.
- Three out of three treated eyes exhibited slight conjunctival redness and chemosis at the 1-hour observation point (grading score of 1). They persisted in 2 of 3 animals for 24 hours and were resolved by the 48-hour time point. In the other animal, all signs of irritation were resolved by the 24-hour observation point.

TNPP was considered to be a slight irritant to the ocular tissue of New Zealand White rabbits.

● Ciba-Geigy conducted an eye irritation study in rabbits in 1981. The study was not conducted in compliance with GLP and the international standardisation of testing methods, however, the procedure used is based on the Proposed Guidelines of the US EPA : "Primary eye irritation study" (1978).

The sample provided by the sponsor was Irgafos TNPP (trade name) but there is no information about its purity.

The test was performed on 3 male and 3 female New Zealand White rabbits. 0.1 ml of the test material was inserted into the conjunctival sac of the left eye of the rabbits. In 3 of the 6 rabbits approximately 30 seconds after treatment, the treated eye was flushed with 10 ml of physiological saline. The eye irritation was appraised with a slit-lamp on day 1, 2, 3, 4 and 7 and was scored for each individual rabbit. The grading system for ocular irritation was similar to the one presented in OCDE test guideline 405.

No corneal opacity was observed in any of the treated eyes, and the iris response was normal in all treated eyes at any of the observation periods. Slight redness and chemosis were the only observable effects. They were completely reversible in rabbits with rinsed eye within 7 days. The test material was found to cause minimal irritation when applied to the rabbit eye mucosa, whether the eyes were rinsed or unrinsed (see Table 4-8).

Table 4-8: Rabbit eye irritation scores (conjunctiva mean reaction scores)

Eye treatment	24 Hours		48 Hours		72 Hours		4 days		7 days	
	Not rinsed	Rinsed								
Redness	1	1	0.3	0.7	0.7	0	1	0.3	0.7	0
Chemosis	1	0.7	0.3	0.7	0.3	0.3	0.3	0.3	0	0

4.1.2.3.3 Summary of irritation

No information is available from human studies. Based on the available data on rabbits, it can be assumed that TNPP is a very slight to moderate irritant to the skin, varying according to tests conditions used : TNPP was a very slight irritant when administered to intact skin for a 4-hours exposure, whereas a 24-hour exposure on intact and abraded skin under occlusive conditions elicited more severe irritation properties. The two available studies indicate that TNPP is a slight irritant to the eye. In each case, the effects were generally reversed within a few days.

Classification and labelling :

According to the cutaneous and eye irritation test methods cited in Annex V, similar to OCDE guideline 404 and 405, TNPP should not be classified as an irritant to skin and eye.

4.1.2.4 Corrosivity

The results from the study of Tay (2001b) indicate that after a 4-hour exposure under semi-occlusive conditions TNPP is not corrosive on intact skin (OECD 404 conditions). However, the study conditions of another study (Ciba-Geigy, 1981) elicit corrosive properties of TNPP. These were harsh conditions (24h exposure under occlusive conditions on abraded and non-abraded skin), furthermore the study report indicates no further details on necrosis observed (was necrosis observed on intact or abraded skin? After what time of application the necrosis was observed?). Based on exposure conditions adopted by OECD guideline for classification, the results of the study of Tay were used in the risk assessment.

Classification and labelling :

TNPP should not be classified as corrosive to skin or eye according to the criteria of the European Union.

4.1.2.5 Sensitisation

Only animal data are available.

4.1.2.5.1 Skin sensitisation

- A Buehler Sensitisation Test was performed, following OECD guideline 406 and GLP (Tay, 2001d).

The degree of purity of the test substance was 99.3%.

TNPP was evaluated for its potential to produce allergenic skin reactions following epicutaneous application to albino guinea pigs. 19 males and 19 females were used for the test. Test animals were distributed into the following groups :

Experimental : 10 males/ 10 females

Negative controls : 5 males/ 5 females

Positive controls : 3 males/ 2 females

Preliminary Irritation : 1 male/ 2 females

- Preliminary irritation : four different concentrations were applied to the skin for 6 hours : 100%, 50%, 25% and 10% of the test substance diluted with 0.9% sodium chloride for injection. The test substance was determined to be a non-irritant, therefore, it was used neat for the induction and the challenge phase.

- Induction phase : closed patches were applied directly to the skin and removed after 6 hours of exposure. The test substance was applied once per week for 3 consecutive weeks (days 0, 7 and 14) on one side of the animals. The positive control substance (dinitrochlorobenzene, DNCB) was applied in the same manner. Naive animals, i.e., untreated during the induction phase, served as a negative control group.

- Challenge phase : The challenge test was performed on virgin skin sites of test and naive animals in the same way as the 6-hour test of the induction phase (closed patches).

All animals gained weight during the course of the study. No abnormal clinical observations were evident in any of the animals during the course of the study.

All animals showed no sign of erythema or oedema at the 24 and 48-hour observation points for the challenge phase. No reactions were observed in the negative control group and 100% reactivity was observed in the positive control group at challenge.

● A Maximisation Test was also performed, following OECD guideline 406 and GLP (Ciba-Geigy, 1992).

The degree of purity of the test substance was > 94%.

The test was performed on 10 male and 10 female guinea pigs in the test group and 5 male and 5 female in the control group.

The sensitivity of the strain is checked every six months with a known sensitiser, such as 2,4-dinitrochlorobenzene, paraphenylene-diamine or potassium-dichromate.

Control group : The control group was treated with adjuvant and the vehicle during the induction period. During the challenge period, the group was treated with the vehicle as well as with the test article to check the maximum subirritant concentration of the test article in adjuvant treated animals.

- Induction (weeks 1 and 2) : it was a 2-stage operation. First, intradermal injection into the neck region (adjuvant/saline mixture, 5% of test article in Oleum arachidis (well tolerated dose) and test article in the adjuvant/saline mixture). Second, one week later, closed patch exposure of 10% TNPP in vaseline (concentration leading to erythema reactions) over the injection sites for 48 hours.

- Rest period : during weeks 3 and 4, no treatment was performed.

- Challenge (week 5) : the animals were tested on the flank with 1% TNPP in vaseline (subirritant concentration) and the vehicle alone (occluded administration for 24 hours).

24 and 48h after removing the dressings, the challenge reactions were graded according to the Draize scoring scale. The body weight was recorded at start and end of the test.

All the animals gained weight during the study. No positive reaction was observed in control animals. In the test group, there were 12/20 and 15/20 positive animals respectively 24h and 48h after occlusive epidermal application (showing erythema scores of 1 to 2). Therefore, TNPP is classified as a strong sensitiser in albino guinea pig according to the grading of Magnusson and Kligman.

● In the study of Majlathova (1981), related to the evaluation of aralkyl phenylphosphite antioxidants by an acute peroral experiment on mice and rats and by epicutaneous and conjunctival test on rabbits, the abstract of the publication states that acute toxic contact dermatitis developed in rabbits after epicutaneous administration of Polygard TNPP exposed to time. The changes healed spontaneously ad integrum within three days after drug withdrawal.

4.1.2.5.2 Summary of sensitisation

No human data is available. The results of the Buehler sensitisation test and of the Maximisation test, both conducted on guinea pig and following OECD TG 406, are not in accordance.

Adjuvant-type tests are likely to be more accurate in predicting a probable skin sensitising effect of a substance in humans than those methods not employing Freund's Complete Adjuvant (FCA), and are thus the preferred methods. Then, the results of the Guinea-Pig Maximisation test will be used for the risk assessment, as this test is considered to be more sensitive than the Buehler test.

No information on respiratory tract sensitisation is available.

Classification and labelling :

TNPP needs to be classified as a skin sensitiser according to the criteria of the European Union (Xi, R43).

4.1.2.6 Repeated-dose toxicity

4.1.2.6.1 Animal data

There is no data for the inhalation and the dermal route.

Four studies provide an assessment of the oral repeated dose toxicity of TNPP.

Three of them were conducted between 1957 and 1961 by the Food and Drug Research Laboratories for the Naugatuck Chemical Company and the last one is a recent reproductive/developmental toxicity screening test conducted by Tyl et al. in 2002.

The Food and Drug Research Laboratories conducted a 90-day study in rats in 1957 and two two-year studies, one in rats and the other one in dogs, in 1961.

These studies were not GLP and didn't follow specific EU or international guideline. However all of the documents provided were acceptable, well-documented study reports, which met basic scientific principles.

- In the 90-day range-finding feeding test with rats (strain not specified) (Food and Drug Research Laboratories, 1957), groups of five male and five female rats were exposed to TNPP (purity not specified) via incorporation in the diet, at doses of 0.2, 1.0 and 5.0% of the daily feeding ration, designed to provide each day about 200, 1000 and 5000 mg/kg bw of TNPP.

The rats were inspected daily for appearance and behaviour. Bodyweight and food intake were recorded weekly for 12 weeks and the efficiency of food utilisation (EFU) was calculated. Haematological (haemoglobin, hematocrit and white cell count) and chemical examinations (blood sugar and blood non-protein nitrogen) were made on the blood of two male and two female rats per group at the 12-week period. All rats that died and all survivors (sacrificed at the end of the 90-day period) were examined at autopsy for evidence of gross pathology. The liver weights were determined for all survivors.

Up to the dose of 1000 mg/kg/d, no adverse effect of the ingestion of this chemical was observed.

In the group exposed to a dose of 5000 mg/kg/d, two females died (on the 35th and 49th day respectively). The principal abnormalities seen at autopsy suggest that the deaths were at least in part due to pulmonary pathology (fibrinous exudate in thorax and hemorrhagic lungs).

In this same group, growth and food efficiency were depressed to a highly significant ($p=0.001$) degree in both sexes. These were the only results which were given a statistically significance. The report didn't indicate which statistical test was used, however, it is reasonable to assume that the statistics presented are intergroup comparisons using the Student's t-test.

The haematological observations disclosed no evidence of abnormalities at any dose level. The blood sugar levels were slightly increased at the dose of 200 mg/kg/d and slightly depressed at the 5000 mg/kg/d dose. The blood non-protein nitrogen levels were slightly elevated in rats of both sexes at the 5000 mg/kg/d dose and in the male at the 1000 mg/kg/d dose. However, all values for both of these biochemical components were within normal limits for the rat.

Pathological changes were observed in the lung and the kidney. Pulmonary lesions were lesions commonly seen in laboratory rats. In the kidney, no change was observed in the test groups up to the 1000 mg/kg/d dose level but at the dose of 5000 mg/kg/d, 8 of the 9 animals examined showed evidence of acute and chronic pyelonephritis with foci of calcification. Chronic pyelonephritis typically represents an inflammatory process that begins in the renal pelvis area (either primarily or as an extension from the urinary bladder) and often spreads upwards into the kidney. There can be many contributory factors associated with pyelonephritis, thus it may not be treatment-related.

However, based on pathological changes observed in the kidney at the dose level of 5000 mg/kg/d, this study identifies a NOAEL (No Observed Adverse Effect) of about 1000 mg/kg bw/d (1% TNPP in the diet) and a LOAEL (Low Observed Adverse Effect) of about 5000 mg/kg bw/d (5% TNPP in the diet for a 90-day exposure of rats).

Chronic ingestion studies on TNPP (purity not specified) were conducted in rats and dogs (strains not specified) over a 2-year period. The test material was incorporated in nutritionally adequate rations for the respective species at levels of 1000, 3300 and 10 000 ppm in the diet and comparison was made with control groups receiving the basal diet without the additive.

- The rat experiment (daily intake corresponding to 50, 167 and 500 mg/kg bw), combined a chronic and a reproductive toxicity study. Weanling rats were distributed into four groups of 25 males and 25 females each. Observations were made of behaviour and appearance, growth, food intake and efficiency. At 12 weeks and at approximately half-yearly intervals, clinical examinations were made in ten rats of each sex in the control and highest test level groups and in five of each sex at the lower levels. These include erythrocyte and leukocyte counts, blood haemoglobin, hematocrit, sugar and non-protein nitrogen determinations, and urine examinations for protein, sugar and sediment. Blood cholesterol levels, prothrombin time and various other biochemical determinations were made at several intervals. After the rats were on test for approximately 100 days, reproduction and lactation studies were initiated and mating continued through the lifetime of the females of the F0 generation (leading to a total of six matings). Ten representative rats of each sex from the second litters produced by the females were selected at the time of weaning and placed on the same ration as their respective parents. Observations similar to those described above were carried out through these and two additional descendant generations. Rats that died or were sacrificed when moribund were examined grossly. At the termination of the two-year period of the F0 rats and at various periods after the weaning of the second litter in the descendant generations of rats (72, 36 and 14 weeks respectively for F1, F2 and F3), they were sacrificed and also examined grossly. At the time of autopsy, eight of the major organs were weighed (liver, spleen, heart, testes and ovaries, and the adrenal, thyroid and pituitary glands). Extensive histopathological examinations were carried out in the F0 generation.

At levels up to 167 mg/kg/d, no adverse effect was noted in any generation, with respect to any of the criteria employed.

At the dose of 500 mg/kg/d, there was a slight but statistically significant retardation in growth of the F0 ($p=0,05$), F2 ($p=0,001$) and F3 ($p=0,05$) males and of the F3 females ($p=0,001$). The weight attained at 12 weeks was approximately 20 g lower than in the other corresponding control groups. The efficiency of food utilisation was slightly depressed in these same groups, with a significant difference from controls in F0 and F2 males ($p=0.05$) at the highest dose and in F3 females at the 2 highest doses used ($p=0.001$). In F3 females, the decrease of food utilisation efficiency was dose related.

Haematological examinations were made at five different periods up to the 100th week and they didn't disclose any aberration. In all groups of each generation and at all time intervals, the findings were within normal limits for the rats. Blood sugar levels and blood non-protein nitrogen levels were normal too. At the 10 000 ppm level, serum cholesterol level was elevated in the females of the F2 generation and in the males of the F0 generation. Urinary findings were negative.

The survival data showed few deaths occurring during the first year of the study. Mortality during the second year was significantly higher among the females that were being carried through the reproduction studies than among the males but the effects were not dose-related. As a matter of fact, the survival of the low and middle dosage levels female rats were somewhat better than the control group, which in turn was equal to the 500 mg/kg/d level. Survival of the males at the highest dose level was 68% compared to 76% in the controls and was better than observed with the two lower dose levels.

The table summarising the organ weight of the F0 generation is missing in the study report. However, information could be found in the original study report of the Food and Drug Laboratory Research : 5/25 F0 females were reported to have a very high absolute liver weight (6.08-8.36 g). Furthermore the missing table also indicates an increased absolute kidney weight of F0 males at the dose level of 500 mg/kg/d. No statistical method was applied to the organ weight data.

The authors indicate a possible dose-dependant increase in liver weight of the F0 females receiving 500 mg/kg/d. They suggest that this may be attributable to the stress of frequent pregnancies and lactation with the concomitant increase in food intake and hence, elevation of dosage level.

In the F0 generation, the principal gross abnormalities seen at autopsy were pulmonary (inflammation, congestion, infection, bronchiectasis...), but of a character commonly seen in laboratory-housed rats. They occurred in one-third to one-half of the rats of each group. Other findings were scattered throughout the groups and in various organs, but in no case did there appear to be a dose-relationship. The liver and kidneys were examined in at least 20 rats of each sex per group and 19 additional organs were examined in at least 10 rats of each sex per group. The incidence of positive finding was somewhat greater in the males than in the females ; however, they do not indicate any particular organ to be affected nor any significant differences between the test groups and the controls. Several of the livers that had elevated weights in relation to body weight showed no histopathologic alterations. The tumour incidence at all levels of TNPP was approximately the same as that of the control group and consisted mainly of fibroadenomas and fibromas of mammary origin. The gross findings at autopsy of the rats of the F1, F2 and F3 generations also revealed no dose-related effects, the principal change being in the lungs and the incidence being greater in the older rats. However, in no case was it greater in the test groups than in the controls (Food and Drug Research Laboratories, 1961).

Overall, based on limited observed effects at the highest dose : a slight retardation of growth in males and an elevation of the absolute liver weight in F0 females, a NOAEL of about 167 mg/kg bw/d (3300 ppm of TNPP in the diet) was derived for this study and 500 mg/kg bw/d (10 000 ppm) is considered as a LOAEL.

- In the study with dogs, three males and three females per group were fed with a diet containing 1000, 3300 and 10 000 ppm TNPP. The report didn't indicate the equivalence between the doses expressed in ppm and the doses in mg/kg bw, absorbed by the dogs. No reproduction testing was conducted, but the same observations and clinical tests as in the rats study were carried out in all of the dogs. In addition, particularly in the control and highest dose level groups, analyses were made of blood serum cholesterol levels, alkaline phosphatase, SGO (Serum Glutamooxaloacetate) and SGP (Serum Glutamopyruvate) transaminases, and anticholinesterase activity. A series of neurological reactions were tested in the control and in the highest dose level group at 12 weeks and at frequent periods thereafter (evaluation of behaviour, of patellar, tonic neck and tonic eye reflexes, and of placing, supporting and righting reactions, were made).

When the study had been under way for five month, one dog at the 1000 and another at the 3300 ppm level died with symptoms of encephalitic meningitis. After confirmation of the diagnosis at autopsy, these deaths were considered irrelevant to the study and the animals were replaced.

For all the other dogs, there was no adverse effect on growth, behaviour and appearance, at any dose level, during the course of the study. Neurological reactions were normal.

The haematological data remained in normal ranges throughout the test period for all groups. No abnormal values were found in blood sugar or non-protein nitrogen levels either. Starting at 48 weeks, the blood cholesterol level of one of the 10 000 ppm female dogs was markedly elevated and continued so to the end of the study. At 100 weeks, there were also high cholesterol levels in the females of this level and also in the female controls, but the values were not excessive in the 1000 and 3300 ppm groups. Alkaline phosphates, SGO and SGP transaminases and anticholinesterase values fell within normal limits for dogs. Urinary findings were negative.

Microscopic examination of approximately 23 organs and tissues was realised.

A reticulum cell sarcoma was observed in one male dog at the lowest dose. In this same dog, abnormal germinal epithelial cells were observed in testes. However there was no related finding in any of the dogs at the higher levels.

One female dog exposed to the 3300 ppm level exhibited some granulomas in lungs and a myocarditis and another female dog fed with the highest dose level exhibited reticulated cytoplasm in some hepatic cells. One male dog exposed to the 3300 ppm level exhibited some mucinous degeneration in media of aorta. These findings were considered incidental.

One male dog exposed to the highest dose exhibited a chronic inflammation in renal pelvis. This finding may be linked to the renal impact of TNPP observed in male rats in the 90-day study (described above) and in the OECD TG 421 study (described below).

One other finding of possible significance in the gross or histopathological examinations was a slight to moderate degree of hyperplasia of the thyroid (with focal collections of lymphocytes) in two female dogs at the highest dose level group. A very slight hyperplasia of the thyroid (focal collections of lymphocytes) was also observed in one male control dog (Food and Drug Research Laboratories, 1961).

Based on this thyroid change at the highest dose, a NOAEL of 3300 ppm and a LOAEL of 10 000 ppm of TNPP in the diet were derived from this study with dogs (Food and Drug Research Laboratories, 1961).

- A reproductive/developmental toxicity screening test of TNPP with rats, conducted by Tyl et al., 2002, also provides some information on repeated dose toxicity. This well-conducted study (also described in chapter 4.1.2.9) meets and enhances the OECD testing guideline 421. The purity of the test material was of 99.98%.

TNPP was administered by oral gavage once daily, seven days per week in CD[®] (Sprague-Dawley) rats at dose levels of 50, 200 and 1000 mg/kg/day, at a dose volume of 5 ml/kg/day in Mazola[®] corn oil. Animals were divided in groups of ten per sex per dose. TNPP was administered for two weeks of prebreed exposure (males and females) and two weeks of mating (males and females) for F0 parents. F0 females continued to be dosed for three weeks each of gestation and lactation, as were F1 offspring (ten per sex per treatment group) from weaning through scheduled sacrifice, at approximately 85 days of age. In addition, five F0 males per group from the control and the 1000 mg/kg/day groups were designated as recovery animals and held without dosing for two weeks, after the F0 male dosing period was completed to evaluate recovery from any possible treatment related effects identified in the high dose.

Observations for mortality were made twice daily and the general condition of all animals was checked daily. Clinical examinations, body weights and feed consumption were recorded

regularly. All F0 parental animals in all groups, as well as the recovery males, all retained F1 adults and non selected F1 weanlings in all groups were subjected to a complete gross necropsy, and full histopathology of the organs was performed for five high dose and control F0 and F1 males and females.

Unscheduled deaths occurred in F0 females at 50 and 1000 mg/kg/day (1 and 4 F0 females respectively). The unscheduled deaths of the low dose F0 female during gestation and one of the high dose F0 female during lactation were attributed to dosing errors and were not considered treatment related. Of the three remaining unscheduled F0 females' deaths, all were found on gestation day 22, possibly attributable to dystocia. Dystocia was evident due to the inability of the dams to deliver their pups. Their demise was considered treatment related. However, this effect is linked with reproduction and thus, won't be taken into account for the assessment of repeated dose toxicity.

F0 parental females did not exhibit any other overt adult systemic toxicity at any dose, as evidenced by a lack of statistically significantly different body weights or gross necropsy findings. However, trends towards increased feed consumption in females from the high dose group (except during lactation) were noted. The authors considered this finding as most likely because of the excessive rooting behaviour observed during the dosing period, however, the excess rooting behaviour observed in F0 females at the highest dose tested was observed during gestation and lactation but not during the prebreed and mating period. Gross necropsy and histological findings of F0 parental females exhibited no treatment- or dose-related pattern of incidence or severity at scheduled sacrifice.

There was no unscheduled death for the adult F1 females. There was no significant difference in body weight or weight gain for the F1 females during the post weaning period. Feed consumption values, presumably associated with excessive rooting behaviour, were increased at 1000 mg/kg/day. There were no treatment-related effects for the gross necropsy or histopathological findings.

There was no treatment-related death for the F0 males. Minor systemic toxicity was present at 1000 mg/kg/day, expressed as trend toward decreased body weights and reduced body weight gains. Feed consumption values, presumably associated with excessive rooting behaviour, were increased at 1000 mg/kg/day during mating. Paired kidney weights, both absolute and relative to terminal body and brain weights, were significantly increased at 1000 mg/kg/day. Histological findings included minimal corticomedullary junction mineralisation of the kidney in three males out of five at 1000 mg/kg/day (with no male with this finding at 0 mg/kg/day), which correlated with the increase of kidney weight at this dose. There was no effect on kidney weights in the recovery group.

There was no unscheduled death for the adult F1 males. There was no significant difference in body weight or weight gain during the postweaning period at any dose. Increased feed consumption at 1000 mg/kg/day, considered related to increasing in rooting behaviour was observed. There was no treatment-related effect for gross necropsy findings. However, histologic findings of F1 males included minimal (one male) and moderate (one male) corticomedullary junction mineralisation of the kidney in two males at 1000 mg/kg/day, versus none at 0 mg/kg/day. These findings were considered treatment related.

The renal lesions observed in F0 and F1 males, were characterised by the presence of basophilic deposits of mineral occurring along the corticomedullary junction. These findings were considered treatment related, since this kind of lesion is rarely, if ever, observed in control males (although it is a common finding in control females). The mineralisation

observed within the high-dose F0 and F1 males appeared similar to the mineralisation noted in the female (control and treated) animals. The increased kidney weight data in the males could have been related to the mineralisation.

Corticomedullary mineralisation in the rat is often diet and/or sex related, but in this study, the reason for the presence of corticomedullary mineralisation in the F0 and F1 male rats could not be determined. No evidence of necrosis or other lesions, which could lead to mineralisation within this area, were observed.

A variety of other histopathologic changes were observed in males and females of both generations at both control and dosage levels (e.g., cyst on renal medulla, necrosis of renal tubule epithelium and nephropathy). These changes were typical of the spontaneous microscopic renal pathology that can be observed at this age and in this strain of rat and were not considered treatment related lesions (Tyl *et al.*, 2002).

The abnormal rooting behaviour which is reported in rats at the highest dose level could be associated with a neurotoxic activity of the test compound.

Based on the renal lesions in F0 and F1 males and on the abnormal rooting behaviour (males and females) observed at the dose level of 1000 mg/kg/day, the NOAEL derived from this study for repeated dose toxicity was 200 mg/kg/day.

- A study on delayed neurotoxicity was performed in chicken (Van Velsen *et al.*, 1980). The study report was in Dutch ; data presented in the report come from a summary in English. Chickens were exposed to a mixture of tris(mono- and dinonylphenylphosphite (Polygard^R) and as positive controls two organophosphates, tri-*o*-cresylphosphate (TOCP) and *O*-methyl, *O*-4-bromine-2,5-dichlorophenyl, phenylphosphonothioate (leptofos) known for causing delayed neurotoxicity. Hubbard chickens weighing 1.5 to 2.5 kg were exposed on day 0 to 4 ml olive oil as control, 400 mg leptofos, 500 mg TOCP or 4000 mg Polygard per kg bw by gavage into the gizzard (6 animals per group). Body weights and food intake were determined once a week. Chickens were taken out of the cage every day to observe the gait and appearance. On day 0, 7, 14, 21 and 28, cholinesterase activity in plasma was measured. From day 28 on, autopsy was performed on 6 animals per day by perfusion via heart with 4% formaldehyde and 1% glutaraldehyde in phosphate buffer (3 animals per group) or by abdominal exsanguination (3 animals per group) both after intravenous injection with Nembutal. From all animals, heart, brain, liver, kidney, stomach and gizzard were histopathological investigated. From the perfused animals, the spinal column and peripheral nerves (nervus ischiadicus, nervus tibialis and nervus peronealis with part of the innervated muscles) were dissected.

Body weights were significantly reduced in the leptofos group (to 64%) and TOCP group (to 87%) with no effects in the Polygard group. Food intake was significantly higher in the Polygard groups in the second, third and fourth week, while leptofos induced a lower food intake. Animals in the leptofos and TOCP group showed signs of delayed neurotoxicity. Cholinesterase activity was not changed in any of the groups on all days. TOCP and leptofos showed degenerative changes in the axons and myelin sheath in the spinal cord and peripheral nerves. No degenerated axons were found in the control and Polygard group. Histological investigations of the other organs resulted in no effects caused by the different treatments.

The authors concluded that it is not plausible that Polygard would result in delayed neurotoxicity in humans, based on the lack of clinical and electronmicroscopical detectable symptoms of delayed neurotoxicity after a single exposure to 4000 mg Polygard/kg bw to chickens.

Human data

No human data was available

4.1.2.6.2 Summary and discussion of repeated dose toxicity

For repeated dose toxicity, confidence is gained by the evaluation of several generations in the two-year studies. These studies provide a profile of limited repeated dose toxicity for TNPP.

A 90-day exposure to a dose of 5000 mg/kg/day (5%) of TNPP in rat resulted in the observation of toxic symptoms and of pathological changes in the kidney, but no adverse effect was observed at lower doses. Over a longer period (2-year), ingestion of TNPP at a dose level of 10 000 ppm (corresponding to 500 mg/kg/d in rats) led to a slight retardation of growth in male rats, an increase of the liver weight in F0 female rats and a thyroid change (doubtful relationship to dosage) in dogs. One male dog exposed to 10 000 ppm also exhibited a renal chronic inflammation in pelvis. In these 2-year studies, 3300 ppm of TNPP in the diet (corresponding to 167 mg/kg/d in rats), was derived as a NOAEL, both for rat and dog. In the modified and enhanced OECD TG 421 study with rats, the NOAEL for systemic toxicity was established at 200 mg/kg/day, based on an excessive rooting behaviour in males and females and on a treatment-dependent corticomedullary junction mineralisation of the kidney in males observed at the highest dose level (1000 mg/kg/day). However, microscopic examination was only performed on 5 males and 5 females of the control and the highest dose group, thus, the NOAEL could not be used for the risk assessment.

Based on this lack of information in the study of Tyl *et al.* and on the respective duration of the studies, the NOAEL used for risk assessment for repeated dose toxicity is 3300 ppm (corresponding in rats to 167 mg/kg), derived from the 2-year study in rat (Food and drug research laboratories) and based on the following limited effects : a slight retardation of growth in males and an elevation of the absolute liver weight in F0 females. This NOAEL is rather conservative.

It is relevant to note that the renal mineralisation observed in males (in the OECD 421 study) at the 1000 mg/kg/day dose of TNPP is similar to the lesions routinely observed in NP treated rats (Cunny *et al.*, 1997 ; Chapin *et al.*, 1999).

The effects observed at the 1000 mg/kg/day dose in the modified and enhanced OECD 421 study are likely associated with the known hydrolysis of TNPP to NP.

Factors such as hydration, diet, or intratubular pH may alter the mineral balance within kidneys (Montgomery *et al.*, 1990 ; Kahn *et al.*, 2002). Additionally, compounds with vitamin D activity could promote mineralisation. Compounds such as oestrogen or having estrogenic activity can influence mineralisation as well, however, the high-dose, F0 and F1 females did not show any evidence of increased severity of mineralisation. There are sex-related differences in the renal metabolism and handling of some xenobiotics in the rat kidney which could have also influenced this change. In particular female kidneys present some kind of down regulation to oestrogen-like compounds as they are exposed to a high level of oestrogens in physiological conditions, whereas male kidney which are not exposed to such a high level of oestrogen are more reactive to an oestrogen-like stimulation.

To date, a conclusive mechanism for the kidney mineralisation induced by TNPP or NP couldn't be provided. A recently completed study with NP confirmed the mineralisation in male kidneys with at NOAEL of approximately 15 mg/kg/day. However, efforts to provide greater insight into the mechanism of the effect were unsuccessful.

It could be suggested that abnormal rooting behaviour, reported in rats at 1000 mg/kg/day in the study of Tyl *et al.* (2002) could be linked with a neurotoxic activity of the test compound. However, "rooting in bedding" typically postdosing (but also predosing) in a dose-related incidence was observed in every gavage study performed in rats in the laboratory which conducted the study and in many others too. The consensus is that it is an expression of taste aversion, likely the animal's attempt to get rid of the bad taste in its mouth from the oral gavage dosing. The higher the dose, the more test material, the greater the incidence of rooting; in this study all rooting was observed postdosing. This behavior is therefore considered indicative of a conditioned adaptive behavior. Furthermore, abnormal behaviour was not observed in the other available studies. An unpublished study carried out by the Dutch National Institute of Public Health and Environment, on delayed neurotoxicity in chickens did not show any evidence of delayed neurotoxicity in chickens for TNPP (Van Velsen *et al.*, 1980).

Classification and labelling :

This chemical is not classified according to the criteria of the European Union. R48 should not be applied.

4.1.2.7 Mutagenicity

Only data from *in vitro* test systems are available.

4.1.2.7.1 Studies *in vitro*

Genetic mutations

- A bacterial reverse mutation assay was recently performed by Wagner and Klug, Bioreliance Laboratory (2001). It was conducted according to the GLP and followed the OECD guideline 471. The purity of the test article was of 98 to 99%.

Salmonella typhimurium tester strains TA98, TA100, TA1535 and TA1537 and *Escherichia coli* tester strain WP2 *uvrA* were exposed to TNPP with and without S9 activation. All dose levels of test article, negative controls and positive controls were plated in triplicate.

The substance was dissolved in acetone. Acetone alone was used for the negative controls. Positive control experiments were carried out simultaneously with the following substances : 1) all *Salmonella* strains and WP2 *uvrA* with S9 : 2-aminoanthracene ; 2) TA98 without S9 : 2-nitrofluorene ; 3) TA100 and TA1535 without S9 : sodium azide ; 4) TA1537 without S9 : 9-amino-acridine ; 5) WP2 *uvrA* without S9 : methyl methanesulfonate.

A preliminary toxic assay was used to establish the dose-range over which the test article would be assayed. Vehicle and ten dose levels of the test article were plated, one plate per dose, with overnight cultures of TA98, TA100, TA1535 and TA1537 and WP2 *uvrA* on selective minimal agar in the presence and absence of Aroclor-induced rat liver S9.

The mutagenicity assay was then used to determine the mutagenic potential of the test article. A minimum of five dose levels of test article (0, 75, 200, 600, 1800, 5000 µg/plate) along with appropriate vehicle and positive controls were plated with TA98, TA100, TA1535 and TA1537 and WP2 *urvA* in the presence and absence of Aroclor-induced rat liver S9.

In the preliminary toxicity assay, the maximum dose tested was 5000 µg per plate. Precipitate was observed beginning at 1000 µg per plate. No appreciable toxicity was observed. Based on the findings of the toxicity assay, the maximum dose plated in the mutagenicity assay was 5000 µg per plate. Precipitate was observed beginning at 600 µg per plate. No appreciable toxicity was observed and no positive response was observed with any of the tester strains in the presence and in the absence of metabolic activation.

TNPP was concluded to be negative in the Bacterial Reverse Mutation Assay.

- Another bacterial reverse mutation assay was performed by Ciba Geigy in 1990. This Salmonella/mammalian-microsome mutagenicity test was carried out in accordance with GLP and the OECD guideline 471 (May 26, 1983) with the exception of statistical analysis. The purity of the test substance was > 94%.

Salmonella typhimurium tester strains TA98, TA100, TA1535 and TA1537 were exposed to TNPP with and without S9 activation. The activation mixture contained S9 fraction of liver from rats induced with Aroclor and a solution of co-factors.

The substance was dissolved in acetone. Acetone alone was used for the negative controls. Positive control experiments were carried out simultaneously with the following substances : 1) for strain TA 98 : donorubicin-HCl 2) for strain TA 100 : 4-nitroquinoline-N-oxide 3) for strain TA 1535 : sodium azide 4) for strain TA 1537 : 9(5)-amino-acridine hydrochloride monohydrate.

All dose levels of test article, negative controls and positive controls were plated in triplicate and the experiments were repeated in order to confirm the results.

A preliminary toxic assay (9 concentrations ranging from 20 to 5000 µg/0.1ml) was used to establish the dose-range over which the test article would be assayed. From the results obtained, the highest concentration suitable for the mutagenicity test was found to be 5000 µg/0.1 ml. The following concentrations were used with and without microsomal activation : 313, 1250, 2500 and 5000 µg/0.1 ml.

The test substance is considered to be positive in this test system if a reproducible increase of the mean number of revertants per plate above that of the negative control, at any concentration level, by at least a factor of 1.5 and 2 respectively for strain TA 100 and for strains TA 98, TA 1535 and TA 1537, is observed.

In the experiments performed without and with microsomal activation, comparison of the number of histidine-prototrophic mutants in the controls and after treatment with TNPP revealed no marked differences. At the concentrations of 2500 and 5000 µg/0.1 ml, the test substance precipitated in soft agar.

No evidence of the induction of point mutations by TNPP or by its metabolites formed as a result of microsomal activation was detectable in the strains of *Salmonella typhimurium* used in these experiments.

- Other Ames tests were conducted in a large Japanese study (Hachiya, 1987), which aim was the evaluation of chemical genotoxicity by a series of short-term tests (82 substances

were subjected to a battery of short-term assays). However, documentation is insufficient for an assessment of the study, as the report is in Japanese version, except for the summary and the table of results.

The results with TNPP were all negative with and without metabolic activation for the *Salmonella typhimurium* strains TA97, TA98, TA100 and TA102 and for *Escherichia coli* strain WP2/pKM102, at concentrations up to 5000 µg/plate.

- An *in vitro* mammalian cell gene mutation test was conducted too, following OECD guideline 476 and in compliance with GLP (San and Clarke, 2001). The purity of the test article was of 98 to 99%.

TNPP was tested in the L5178Y/TK^{+/-} Mouse Lymphoma Mutagenesis Assay. Exposures were for 4 hours in the absence and presence of metabolic activation (Aroclor-induced rat liver S9). The vehicle used was acetone. Methyl methanesulfonate was used as the positive control for the non-activated test system and 7,12-Dimethylbenz(a)anthracene was used as positive control for the S9- activated test system.

Based on the results of the preliminary toxicity assay, the doses chosen for treatment of the mutagenesis assay ranged from 1.0 to 200 µg/ml for both the non-activated and S9-activated cultures. Visible precipitate was present at concentrations of ≥ 100 µg/ml in treatment medium. No visible precipitate was present at concentrations of ≤ 50 µg/ml in treatment medium. The doses chosen for cloning ranged from 5.0 to 100 µg/ml with and without S9 activation. No cloned cultures exhibited mutant frequencies, that were at least 55 mutants per 10⁶ clonable cells over that of the solvent control. Toxicity in the cloned cultures was not observed at any dose levels.

TNPP was concluded to be negative in the L5178Y/TK^{+/-} Mouse Lymphoma Mutagenesis Assay.

- An *in vitro* gene mutation test with Chinese hamster cells V79 was also conducted by Ciba-Geigy (1990). The test was carried out in accordance with GLP and the OECD guideline 476. The purity of the test substance was > 94%.

TNPP was dissolved in ethanol. Two negative controls (ethanol) and one positive control (N-nitroso-dimethylamine) were also tested. The study was conducted with and without activation (Aroclor-induced rat liver S9). The cells were treated in the experiments with microsomal activation for 5 hours and in the experiments without microsomal activation for 21 hours. The results of each original experiment were confirmed in a second and independent experiment (confirmatory experiment). Based on the results of a preliminary toxicity assay, the original experiments were performed at the following concentrations with microsomal activation : 0.6, 1.2, 2.4, 4.8, 7.2, 9.6 and 12.0 µg/ml and without microsomal activation : 0.3, 0.6, 1.2, 2.4, 3.6, 4.8 and 6.0 µg/ml. Because the intended toxicity was not obtained in the original experiments, in the confirmatory experiments, the concentrations applied were increased to 0.8, 1.6, 3.2, 6.4, 9.6, 12.8 and 16.0 µg/ml with microsomal activation and to 0.4, 0.8, 1.6, 3.2, 4.8, 6.4 and 8.0 µg/ml without microsomal activation. In all the experiments, comparison of the number of mutant colonies in the controls and in the cultures treated with the various concentrations of the test substance revealed no significant deviation of the mutant frequencies.

In both investigations with and without microsomal activation, criteria for a negative response were reached : a difference in the treated and untreated dishes of at least 20 clones per 10⁶

cells plated was not detected and there was no indication of a concentration mutant-frequency relation in any experiment.

TNPP was concluded to be negative in the gene mutation test with Chinese hamster cells V79.

Chromosomal effects

- One mammalian chromosome aberration assay, following OECD guideline 473 and in compliance with the GLP was carried out on Chinese hamster ovary (CHO) cells both in the absence and presence of metabolic activation (Aroclor-induced S9 activation system) (Gudi and Brown, 2001). The purity of the test article was of 98 to 99%.

Acetone was determined to be the solvent of choice, based on the solubility of the test article and compatibility with the target cells. Mitomycin C was used as the positive control in the non-activated test system and cyclophosphamide was used as the positive control for the S9 activated test system.

The preliminary toxicity assay was performed for the purpose of selecting dose levels for the chromosome aberration assay and consisted of an evaluation of test article effect on cell growth. The chromosome aberration assay was performed using standard procedures (Evans, 1976), by exposing duplicate cultures of CHO cells to the test article as well as positive and solvent controls.

Dose levels for the chromosome aberration assay were selected based upon the lowest precipitating dose : the doses chosen ranged from 18.75 to 200 µg/ml for both the non-activated and the S9 activated 4-hour exposure groups and from 6.25 to 150 µg/ml for the non-activated 20-hour continuous exposure group. No toxicity and no statistically significant structural or numerical chromosomal aberrations were observed under any treatment condition in the assay.

TNPP was concluded to be negative for the induction of structural and numerical chromosome aberration in Chinese hamster ovary cells.

- Another mammalian chromosome aberration assay, following OECD guideline 473 and in compliance with GLP was carried out on Chinese hamster ovary cells (cell line CCL 61) both in the absence and presence of metabolic activation (Aroclor-induced S9 activation system) (Ciba-Geigy, 1990). The purity of the test article was > 94%.

TNPP was dissolved in acetone. Acetone was used as negative control. Mitomycin C 0.2µg/ml, a mutagen not requiring S9 activation and cyclophosphamide 40.0 µg/ml, which requires activation, were used as positive controls.

Based on the results of a preliminary toxicity assay, the 2 experiments of the original study were performed at the following concentrations : 62.5, 125.0 and 250.0 µg/ml (with activation and without activation). Since at the upper concentration of 250.0 µg/ml, only metaphase of inferior quality, insufficient for scoring were present, the concentrations of 31.25, 62.5 and 125.0 µg/ml were selected for the four experiments of the confirmatory study (with and without activation).

The number of cells with specific chromosomal aberrations in the treatment groups showed no marked difference in comparison with the negative control. The incidence of changes observed is within the range of spontaneous aberrations inherent to this particular cell line used.

No evidence of clastogenic effects was obtained in Chinese hamster ovary cell *in vitro* treated with TNPP.

4.1.2.7.2 Studies *in vivo*

No *in vivo* study is available.

4.1.2.7.3 Summary of mutagenicity

In vitro mutagenetic tests did not reveal any genotoxic effect in six well-conducted tests, two Bacterial Reverse Mutation Assays, two *in vitro* Mammalian Cell Gene Mutation Tests, and two *in vitro* Mammalian Chromosome Aberration Tests.

Although neither human data nor *in vivo* tests are available, the available data from *in vitro* tests support the view that TNPP is a non-genotoxic substance.

Classification and labelling :

This chemical is not classifiable as mutagenic according to the criteria of the European Union.

4.1.2.8 Carcinogenicity

There is no carcinogenicity study available, however, two studies that were described in the repeated dose toxicity section (4.1.2.6) were conducted on rats and dogs for 2 years. These studies are not in compliance with international guidelines for the assessment of carcinogenicity because the number of animals used was not sufficient, compared to what is required by the international guidelines. In the rat experiment, there were only 25 rats per sex for each group instead of 50 in the guideline, and not all of them were examined histopathologically for all organs : the liver and kidneys were examined in at least 20 rats of each sex per group and 19 additional organs were examined in at least 10 rats of each sex per group. In the experiment with dogs, 3 dogs per sex per group were used.

In these chronic ingestion studies, TNPP was incorporated in nutritionally adequate rations for the respective species at levels of 1000, 3300 and 10 000 ppm in the diet and comparison was made with control groups receiving the basal diet without the additive.

In the rat experiment, histopathological examinations of the liver and kidney of all except an occasional (autolysed) animal and of the spleen, adrenal, thyroid and pituitary gland, heart, stomach, small and large intestine, pancreas, bladder, gonads, salivary glands, lymph nodes, lungs, bone marrow, muscle, brain and spinal cord in at least half the rats at all levels in F0 generation were conducted. All tumours were examined microscopically in F0 generation.

The tumour incidence (Table 4-9) at all levels of TNPP was approximately the same as that of the control group and consisted mainly of non-malignant tumours (fibroadenomas and fibromas of mammary origin).

In the dog experiment (Table 4-10), a reticulum cell sarcoma was observed in one male dog at the lowest dose. In this same dog, abnormal germinal epithelial cells were observed in testes. However there was no related finding in any of the dogs at the higher levels. (Food and Drug Research Laboratories, 1961).

Table 4-9: Summary of Histopathological Findings in F0 generation rats (Negative findings omitted)

Organ and finding	Dose (mg/kg)							
	0		50		167		500	
	M	F	M	F	M	F	M	F
Tumours :								
Necrotic tumour	1							
Reticulum cell sarcoma	1		2					
Fibroadenoma		6	1	1	1	3		8
Fibroma		1	2	1		1	1	1
Adenoma		1		1		1		1
Linoma			1					
Malignant papillary mesothelioma				1				
Squamous carcinoma				1				
Adenocarcinoma					1			
Spindle cell sarcoma					1			
Liposarcoma					1			
Teratoma						1		
Angiomyoma								1
Plasma cell tumour involving spleen, liver, BM, LN and soft tissues								1
Mammary carcinoma								1

¹ : Liver and kidneys examined in 20 or more rats per sex per group ; the remaining 19 tissues were examined in at least 10 rats per sex per group.

Table 4-10: Summary of gross and histopathological findings in dogs (negative findings omitted)

Level	Dog No & sex ⁴	Fate ⁵	Gross Abnormalities	Histopathological Findings ⁶
<u>ppm</u> :				
None	476M 482M 488M 492M 474F 483F	S-2yr " " " " "	Colon : few petechial hemorrhages Spleen mottled	Thyroid : very slight hyperplasia and focal collections of lymphocytes
1000	479M 489M 493M 472F 495F	S-2yr " D 655 days S-2yr "	Possible tracheal insufflation of food	Mucinous degeneration in media of aorta (1 of 2 sections) Reticulum cell sarcoma in lung, liver, spleen and lymph node. Abnormal germinal epithelial cells in testes. Kidneys : small foci chronic inflammation in medulla and pelvis
3300	477M 490M 494M 473F 485F	S-2yr " " " "	Focal hemorrhages in small intestine	Granulomas in lung; myocarditis
10 000	478M 481M 491M 475F 486F 487F	S-2yr " " " " "	Hemorrhagic mucosa in intestines	Kidneys : chronic inflammation in pelvis Thyroid : slight focal hyperplasia collections of lymphocytes. Moderate diffuse hyperplasia; Liver : reticulated cytoplasm in some hepatic cells

⁴ Dogs No. 464 F (3300 ppm) and 571F (1000 ppm) had not completed the 2-year period at the time of this data collection.

⁵ S = sacrificed, D = died

⁶ Organs examined were liver, kidneys, spleen, aorta, heart, lungs, stomach, small and large intestines, pancreas, gall bladder, urinary bladder, salivary glands, thymus, gonads, adrenal and thyroid glands, lymph nodes, bone marrow, muscle, brain, spinal cord and pituitary gland.

Summary

There is no reliable study available on carcinogenicity, however, on the basis of the information currently available on mutagenicity, TNPP is considered as a non-genotoxic substance, so concerns for cancer caused by a genotoxic mechanism are low.

Considering the potential for carcinogenicity by a non-genotoxic mechanism, no evidence of a significant increase of tumour incidence was found in the 2-year chronic studies carried out on a small sample of rats and dogs.

Although only limited data are available, these data tend to indicate that TNPP is not of concern for a carcinogenic potential.

Classification and labelling :

This chemical is not classifiable as a carcinogen according to the criteria of the European Union.

4.1.2.9 Toxicity to reproduction

Reproduction and developmental toxicity were evaluated in two oral multi-generation studies in rats and in one non-standard study on chick embryos.

Fertility and reproductive toxicity

- The 2-year study on rats (described in the repeated dose toxicity section 4.1.2.6), combines a chronic and a three-generation reproductive toxicity study. This study is not GLP and it doesn't follow specific international guideline, however, this multi-generation study is well-documented and meets basic scientific principles.

Rats were distributed into four groups of 25 males and 25 females each and fed with a diet containing TNPP at a level designed to be equivalent to approximately 1000, 3300 and 10 000 ppm (daily intake corresponding to 50, 167 and 500 mg/kg bw).

After the rats were on test for approximately 100 days, reproduction and lactation studies were initiated and mating continued through the lifetime of the females of the F0 generation. From the second litters born of these dams, young were raised and carried through similar feeding and reproduction studies. Their young, and in turn the descendant of that generation, were carried through similar experiments, making a total of three descendant generations studied. Reproduction and lactation experiments on the F0 generation were carried through six matings. The F1 and F2 generations were carried through only two matings and the F3 generation was not mated.

Records were kept of the date of birth, the number of pups born, their weight and survival during lactation. The criteria employed for evaluating the performance of the rats were a series of indexes for fertility, gestation, viability and lactation.

Reproductive organs (testes and ovaries) were autopsied and weighed.

Growth was normal at all dosage levels in F0, F1 and F2 females. At the dose level of 500 mg/kg/d, there was a slight but statistically significant retardation in growth of the F2 ($p=0,001$) and F3 ($p=0,05$) males and of the F3 females ($p=0,001$), along with a decrease in the efficiency of food utilisation for F2 males ($p=0.05$) at the highest dose and F3 females at

the 2 highest doses used ($p=0.001$). In F3 females, the decrease of food utilisation efficiency was dose related.

The authors state that the findings related to reproductive parameters were comparable with observations in rats of the stock colony and were considered normal. No statistical analysis was performed but this seems true for most of the parameters. There was no indication of adverse effect in the F0 generation at any dose level. However, diminution in the number of pups born per litter in the F1 and F2 high dose groups, and a small decrease in the fertility and viability indexes in F2 at this same high dose level exposure were observed (see Table 4-11). (Food and Drug Research Laboratories, 1961).

Based on those results, indicating a possible effect on reproduction at the dose of 500 mg/kg/d, a NOAEL for reproduction of 167 mg/kg bw/day can be derived.

DRAFT

Table 4-11: Comparison of first two matings in three generations of rats

Dose	Generation	Total No. of mating	No. litters born alive	Pups born alive	Pups per litter born	No. litters weaned	Average weight of pups at weaning ⁷	F.I. ⁸	G.I. ⁹	V.I. ¹⁰	L.I. ¹¹
Mg/kg							Gm				
None	F0	49	41	328	8.0	34	40.0	98.0	82.9	87.2	96.2
	F1	20	19	216	11.3	19	36.3	95.0	100.0	87.0	89.5
	F2	20	17	151	8.9	16	42.7	90.0	94.5	93.2	87.5
50	F0	49	40	354	8.8	36	36.5	91.8	90.0	91.8	88.0
	F1	20	20	213	10.7	20	41.6	100.0	100.0	96.0	90.0
	F2	20	19	159	8.4	16	40.0	95.0	94.5	87.6	81.1
167	F0	50	45	415	9.2	41	37.9	94.0	95.7	95.7	87.7
	F1	20	20	212	10.6	20	40.1	100.0	100.0	95.5	94.5
	F2	20	19	151	8.0	12	42.6	95.0	100.0	94.5	71.0
500	F0	48	40	337	8.4	37	36.0	100.0	83.3	93.8	87.3
	F1	17	16	113	7.0	13	36.0	100.0	100.0	93.5	96.0
	F2	20	17	122	7.3	13	43.8	85.0	100.0	79.7	89.7

⁷ At 21 days⁸ Fertility index = (No. pregnancies / No. matings) X 100⁹ Gestation index = (No. litters born alive / pregnancies) X 100¹⁰ Viability index = (No. pups at 1d. / No. pups born alive) X 100¹¹ Lactation index = (No. pups at 21d. / No. pups at 1d.) X 100

● A study combining a reproductive and a developmental toxicity screening of TNPP was conducted on CD[®] (Sprague-Dawley) rats (Tyl *et al.*, 2002). This recent, well-conducted study, was performed in compliance with OECD Guideline 421 and GLP. It exceeded the OECD TG 421 study design as follows : enhanced evaluation of toxicity in the F0 generation, including the evaluation of a recovery group of males ; evaluation of developmental landmarks in the F1 generation (time of vaginal opening or preputial separation, normality and length of oestrous cycle) ; and following the F1 offspring to adulthood, with continued exposure and assessment of reproductive structures and functions including potential effect on sperm.

The purity of the test material was of 99,98%.

TNPP was administered by oral gavage once daily, seven days per week in rats at dose levels of 50, 200 and 1000 mg/kg/day, at a dose volume of 5 ml/kg/day in Mazola[®] corn oil, ten animals/sex/dose. TNPP was administered for two weeks of prebreed exposure (males and females) and two weeks of mating (males and females) for F0 parents. F0 females continued to be dosed for three weeks each of gestation, lactation, as were F1 from weaning through scheduled sacrifice. On the day of birth (postnatal day or pnd 0), anogenital distance was measured and bodyweight recorded for all live F1 pups in all litters. They were also examined to determine the number of viable and stillborn pups from each litter. Thereafter, litters were evaluated for survival on pnd 4, 7, 14 and at weaning (pnd 21). F1 litters were culled on pnd 4 to yield, as nearly as possible, five males and five females per litter. The culled F1 pups were weighed, euthanized and necropsied with complete external and visceral examinations. For the remaining F1 pups, survival indices were calculated at least weakly through weaning (pnd 21). At weaning, at least one female and one male (whenever possible) from each F1 litter were randomly selected to continue treatment for approximately seven more weeks, with dosing for F1 selected pups begun on pnd 22 until all pups were at least 85 days of age. F1 postweaning observations and procedures for each retained female included examination for patency of vaginal opening (from pnd 22 until acquisition of vaginal opening). Oestrous cyclicity and normality were evaluated by vaginal smears from F1 females taken daily the last three weeks of the postwean exposure period prior to scheduled sacrifice. For each retained F1 male offspring, observations for the cleavage of the balanopreputial gland (preputial separation) began at 35 days of age and continued until acquisition of preputial separation. Andrologic assessment was also performed on the F1 retained males at necropsy. All F0 parental animals and retained F1 adults were subjected to a complete gross necropsy with the following organs : ovaries, uterus with cervix and vagina, prostate, epididymides, testes and seminal vesicles with coagulating glands and their fluids.

In this part of the hazard identification, the results of this study will focus on the effects related to reproduction and development.

Three of ten pregnant F0 females at 1000 mg/kg/day died in late pregnancy (gestation day 22). These deaths may have been related to dystocia, since the dams appeared to be unable to deliver their normal appearing pups. Examination of the F1 pups and the necropsy of 3 dams indicated that the pups were full term and normal in external appearance. Two F0 females respectively exposed to 50 mg/kg/day (during gestation) and 1000 mg/kg/day (during lactation) were also found dead. But these deaths were attributed to dosing errors and were not considered treatment related.

Ovary weights (absolute and relative to terminal body and brain weights) were significantly decreased at 1000 mg/kg/day in F0 but not F1 adult females.

There was no effect of exposure to TNPP on any F0 reproductive indices during the production of F1 offspring. Mating, fertility, pregnancy and gestational indices were equivalent across groups ; gestational length was equivalent across all groups. According to the authors, there was also no difference across groups for the number of total implantation sites per litter, percent post-implantation loss per litter, or number of total, live or dead pups per litter at birth.

There was evidence of F1 offspring toxicity (see Table 4-12) observed postnatally at 1000 mg/kg/day, expressed, according to the authors as reduced litter size on postnatal day (pnd) 4, but not on postnatal day 0. This finding could be linked with maternal toxicity, expressed as a behaviour change towards its litter. However, the assumption of the study report (reduced litter size on pnd 4 but not on pnd 0) is not in accordance with the tables of values provided in the report. The tables indicate that there is a reduction of litter size at the highest dose, compared to the control group and that this reduction is the same at pnd 0 and at pnd 4 tending to indicate a direct offspring toxicity and not an indirect one, linked with a maternal behaviour change.

There was no treatment-related death for the adult F1 males and females. No effect on reproductive parameters, developmental landmarks, F1 oestrous, or F1 andrology was observed. In F1 males, paired epididymides weight, relative to terminal body weights, were significantly decreased at 1000 mg/kg/day.. There was no treatment-related effect for gross necropsy findings in F1 males and females.

Table 4-12: Summary of F1 offspring toxicity

	Trisnonylphenyl Phosphite (mg/kg/day)			
	0	50	200	1000
N° of live litters				
Postnatal Day 0	10	8	10	7
Postnatal Day 4	10	7 ^a	10	7
Postnatal Day 7	10	7	10	7
Postnatal Day 14	10	7	10	7
Postnatal Day 21	10	7	10	6 ^b
Average number of live pups per litter (pnd 0)	14.9** ± 0.5	12.8 ± 1.6	15.9 ± 0.6	12.0 ± 1.4
Average number of live pups per litter (pnd 4, precull)	14.8** ± 0.5	14.3 ± 0.6	15.6 ± 0.5	12.0* ± 1.4
Average number of live pups per litter (pnd 7, postcull)	9.8 ± 0.1	10.0 ± 0.0	10.0 ± 0.0	9.1 ± 0.9
Average number of live pups per litter (pnd 14, postcull)	9.8 ± 0.1	10.0 ± 0.0	10.0 ± 0.0	9.1 ± 0.9
Average number of live pups per litter (pnd 21, postcull)	9.8 ± 0.1	10.0 ± 0.0	10.0 ± 0.0	10.0 ± 0.0

^a The entire litter for female 30 was missing and presumed dead on postnatal day 4.

^b Female 24 was found dead (possible dosing error) on postnatal day 15, therefore, her entire litter had to be euthanized and was not included in any parameters after postnatal day 14.

* p<0.05 Dunnett's test

** p<0.05 ANOVA test

The decrease in ovary weight (absolute and relative) in F0 females and the decrease in relative paired epididymides weight in F1 males, at 1000 mg/kg/d may be related to an hormonal,

oestrogen-like effect of the substance. Actually, regarding the decreased paired epididymides weight, the difference in the dosing period could be a reasonable explanation for, why this organ weight is decreased in the F1 males and not in the F0 males. The F1 males are dosed during the critical period of reproductive system development thereby enhancing sensitivity to endocrine disrupters compared to the parent generation, which are only dosed during adulthood. This may be associated with the known hydrolysis of TNPP to NP. The oestrogen-like activity of NP was evidenced in many studies. However, the role played respectively by NP and TNPP for this effect can't be established here. Furthermore, andrology parameters measured at 1000 mg/kg/day didn't reveal any change compared to controls.

Based on a slight reduction of the litter size, on a slight decrease in relative paired epididymides weight in F1 males and on signs of maternal toxicity (death on gestation day 22, decrease in ovary weight) at 1000 mg/kg/day, NOAELs for maternal and offspring toxicity of 200 mg/kg/day were derived from this study.

Developmental toxicity

- In this reproductive and a developmental toxicity screening study (Tyl *et al.*, 2002), no effect was observed in developmental landmarks in the F1 generation (time of vaginal opening or preputial separation, normality and length of oestrous cycle) at any dose level.

All F1 pups culled on pnd 4 (see Table 4-12) were subjected to a complete external and visceral examination, including examination of all thoracic and abdominal organs, bisection of kidneys and heart dissection. These examinations didn't reveal any developmental effect up to the dose level of 1000 mg/kg/day. The NOAEL for teratogenicity is ≥ 1000 mg/kg/day.

However, it must be underlined that while the OECD 421 study design specifies termination of the study on pnd 4, with external and internal examination of the F1 pups at this time, the modified study design used in this study provides, for continuation of the F1 offspring, with continuing exposure until sexual maturity. Thus, to provide data on the pnd 4 pups, the pups culled to standardise litters on pnd 4 were euthanised and subjected to complete gross necropsy, but this number is very limited since F1 litters were culled on pnd 4 to yield, as nearly as possible, five males and five females per litter. This leads to nearly 2 animals in the highest dose group and 4 in the other groups. The other pups were subjected to a complete gross necropsy at weaning (pnd 21), except for at least one male and one female per litter that were selected to continue treatment for seven more weeks. No malformation was observed at any stage.

- The effects of TNPP on the survival rate of chick embryos were studied. This non-standard study is very shortly reported and is not considered as an adequate developmental toxicity study. The materials employed (purity not indicated) were submitted to the Food and Drug Research Laboratories by Weston Chemical, Inc.

A dose of 5 mg of TNPP in corn oil was injected directly into the yolk sac of fertile hen's eggs. Two replicate runs were conducted, in which groups of 16 fertile White Leghorn eggs were used per group. With each series, there were four comparison groups : (1) untreated eggs, as a control on hatchability, (2) eggs which were drilled and the needle inserted into the yolk sac, without the injection of anything, (3) eggs injected with distilled sterile water and (4) eggs injected with corn oil from the same lot used to prepare the sample solution.

The result was given in terms of percent of survival of the embryos at 5, 10 and 18 days of incubation and in terms of the final number of live chicks which hatched.

It was evident that any manipulation that disrupted the integrity of the egg membrane caused some mortality of the embryos. Mortality was further slightly increased by the injection of either water or plain corn oil. No statistical test was performed but conclusion was made difficult as results appeared to be different between the two runs, one of them showing a very slight effect of the injection of TNPP and the other one showing a more important effect on embryos' survival. Examination of embryos that died before hatching and of the newly hatched chicks revealed no gross abnormality or malformation peculiar to the test groups (Food and Drug Research Laboratories, 1971).

Summary and discussion

TNPP exposure over four generations didn't reveal any significant effect on reproduction up to 500 mg/kg/d, the highest dose tested, except for a possible reduction of litter size, born from F1 and F2 generations at the highest dose. This slight tendency seems to be confirmed by the OECD 421 study in which a slight but significant litter size reduction was observed at the highest dose (1000 mg/kg/day). In this same study, maternal toxicity was observed at the dose of 1000 mg/kg/day. At the dose of 1000 mg/kg/day a decrease of the ovary weight of F0 females and the decrease of epididymides weight in F1 males suggest an oestrogen-like activity of the test substance (directly or indirectly via the hydrolysis of TNPP into NP). No other significant effects on reproductive toxicity were observed in this study.

Phenomenon of dystocia observed in dams at the highest dose in the study of Tyl (2002) is viewed as maternal toxicity, due from the adjustments of dosing volume on gd 14 and especially on gd 20, resulting in over dosing the dams in late gestation. Actually, the dosing volume of the test chemical was adjusted for each dam based on each new body weight. This means that the dosing volumes for the F0 dams during gestation were adjusted on gd 0, 7, 14, and 20. The pregnant rat CD (SD) females gain approximately 150 g or more during gestation but with the body weight gain from gd 14 to parturition (the "last trimester") of at least 100 g, due almost entirely to the rapid growth of the uterine contents. For gavage studies, test chemical intake (in mg/day) during this period is increased by as much as 30% because of the adjustment for maternal body weight, especially from gd 20 to parturition (gd 22 ± 1). Thus, the dose in mg/kg/day, based on the actual maternal body weight minus the uterine contents, is similarly increased by ~30%. This can result in overdosing the dam (and conceptuses) and is likely the cause of the excessive periparturitional maternal toxicity observed.

The risk of increased maternal toxicity in late pregnancy from bolus gavage dosing is due to: (a) the maternal liver (although it is enlarged in late pregnancy in response to the pregnancy and the increased test chemical load) is not enlarged commensurate with the increased test chemical dose; (b) test chemical is likely not equally distributed between maternal and fetal compartments, so the relative maternal burden may be even greater; and (c) gastrointestinal tract motility is reduced in late pregnancy, so there is likely increased absorption of the test chemical from the gut due to longer transit times.

Based on these observations the NOAELs for reproductive toxicity and for maternal toxicity, were 200 mg/kg/day, derived from the OECD 421 study (considered as a key study for risk characterisation as a recent study, following OECD guideline).

No indication of any developmental effect was observed in both of the studies. $NOAEL_{\text{terato}} \geq 1000$ mg/kg/day, although these parameters were observed on a very reduced number of animals.

It has been assumed above that the effects observed at the 1000 mg/kg/day dose in the modified and enhanced OECD TG 421 study were likely associated with the known hydrolysis of TNPP to

NP. For example, the ovary weight changes observed at the high dose level of TNPP are consistent with changes observed for high doses of NP in multi-generation studies (Nagao, 2001). The dose of NP required to elicit this decrease in ovary weight was 50 mg/kg/day by gavage and approximately 150 mg/kg/day in dosed feed. A decrease in epididymis weight is also reported in two rat studies with gestational exposure to nonylphenol (Hossaini et al., 2001; Han et al., 2004).

Further, since saturation of metabolism of NP has not been shown for male rats, the lack of oestrogen-like effects even at the high dose of 1000 mg/kg/day TNPP is predictable since metabolites of NP have been shown to have no estrogenic activity. The only indication of that kind of activity is the observation of the decrease of paired epididymides weight, relative to terminal body weight in F1 males. However, for F1 females, the lack of effects at the 1000 mg/kg/day dose of TNPP on developmental landmarks, specifically advanced vaginal opening seen with higher doses of NP due to oestrogen-like activity, is of interest. Odum *et al.* (2002) have shown that the estrogenic stimulus must be present at or near the time the developmental landmark occurs for the effect to manifest. Therefore, it is possible that rate of hydrolysis of TNPP to NP or the absorption of NP is lower in juvenile rats. The observation of dystocia in the parental animals on gestation day 22 and death of the dams is not present in other studies with TNPP or NP. However, the large gavage dose of TNPP used in these studies could provide for different kinetics to provide a NP dose even higher than that attained in feeding studies.

In conclusion, TNPP is not a reproductive toxicant at doses up to 200 mg/kg/day. Effects above this dose may result from the hydrolysis of TNPP to NP providing NP equivalent doses up to 200 mg/kg/day with a 1000 mg/kg/day dose of TNPP.

Classification and labelling :

This chemical is not classified as toxic to reproduction (fertility and development) according to the criteria of the European Union.

4.1.3 Risk characterisation

4.1.3.1 General aspects

No human data are available, so this assessment of the hazardous properties of TNPP is based only on animal data.

No studies on Toxicokinetics were conducted so the only information we have are those that can be derived from the physico-chemical properties of the molecule : a relatively high molecular weight ($MW = 689\text{g}\cdot\text{mol}^{-1}$), an extremely low water solubility and a high $\text{Log } P_{ow}$. Thus absorption of TNPP is expected to be limited. Results from a hydrolysis test let think that hydrolysis of approximately 20% of TNPP to NP in the gastro-intestinal tract is possible.

From many studies by oral, dermal or intraperitoneal route, it can be assumed that the acute toxicity of TNPP is very low. The derived LD_{50} for oral route is 19.5 ± 3.3 gram/kg. Acute dermal and intraperitoneal limit tests (at 2000 mg/kg and 1000 mg/kg respectively) did not cause any mortality. No data is available on the acute inhalation toxicity, although the non-corrosive

and very slight irritant nature of TNPP may suggest that toxicity would not be enhanced following exposure by this route. It is to note that tests conditions proposed by OECD guidelines don't elicit irritant or corrosive properties of the test substance although stronger conditions may reveal a moderate irritation or a possible corrosive action of TNPP to the skin. TNPP is a slight eye irritant.

According to the maximisation test, TNPP showed a strong grade of skin-sensitising potential in albino guinea pigs. Although a Buehler sensitisation test gave a negative result, the maximisation test is preferred as it is the most sensitive one. No information on respiratory tract sensitisation is available.

The main toxic effect is a renal impact observed in F0 and F1 male rats in a reproductive/developmental toxicity screening test, at the highest dose of 1000 mg/kg/day by oral route (microscopic examination was only performed in control and high dose group animals in this study). Paired kidney weights, both absolute and relative to terminal body and brain weights, were significantly increased at this dose and histological findings included corticomedullary junction mineralisation of the kidney. The renal lesions observed in F0 and F1 males were characterised by the presence of basophilic deposits of mineral occurring along the corticomedullary junction. In a 2-year study on rat, an oral dose of 10 000 ppm (corresponding to 500 mg/kg/day) led to a few effects on growth and liver weight. A NOAEL of 3300 ppm, corresponding to 167 mg/kg/day was derived from this long-term repeated dose toxicity study and can be used in the risk characterisation.

No repeated dose studies with inhalation and dermal application route were available.

Concerning mutagenicity, TNPP was negative in all of the six well-conducted studies available *in vitro* (two Bacterial Reverse Mutation Assays, two *in vitro* Mammalian Cell Gene Mutation Tests, and two *in vitro* Mammalian Chromosome Aberration Tests). No *in vivo* study was available, overall, the evidence indicates that TNPP is not mutagenic.

There are no carcinogenic studies conducted according to international guidelines. However, some information on the carcinogenic potential of TNPP can be derived from other data. On the basis of the information currently available, it is unlikely that TNPP is mutagenic, so concerns for cancer caused by a genotoxic mechanism are low. Considering the potential for carcinogenicity by a non-genotoxic mechanism, no evidence of an increase of tumour incidence was seen in 2-year repeated dose toxicity studies in rat and dog. There is low concern for carcinogenicity by a non-genotoxic mechanism too.

TNPP exposure over several generations didn't reveal any significant effect on reproduction up to the highest dose tested (500 mg/kg/day) in F0 but a slight reduction of litter size born from F1 and F2 generations, which tended to be confirmed by the OECD 421 study in which a slight but significant litter size reduction was observed at the highest dose (1000 mg/kg/day). In this same study, maternal toxicity was observed at the dose of 1000 mg/kg/day. At the dose of 1000 mg/kg/day a decrease of the ovary weight of F0 females and the decrease of epididymides weight in F1 males suggest an oestrogen-like activity of the test substance (directly or indirectly via the hydrolysis of TNPP into NP).

The NOAELs for reproductive toxicity and for maternal toxicity, were derived from the OECD 421 study and were considered to be 200 mg/kg/day.

No indication of any developmental effect was observed in both of the studies. $NOAEL_{terato}$ is \geq 1000 mg/kg/day, although these parameters were observed on a very reduced number of animals

on pnd 4 due to the modified procedure of the study. However, internal and external examination on a larger number of pups, on pnd 21 also gave negative results.

Overall for the risk characterisation :

$$\text{NOAEL}_{\text{maternal}} = 200 \text{ mg/kg/day}$$

$$\text{NOAEL}_{\text{repro}} = 200 \text{ mg/kg/day}$$

$$\text{NOAEL}_{\text{terato}} \geq 1000 \text{ mg/kg/day}$$

4.1.3.2 Workers

Occupational exposure may occur by inhalation and dermal route during manufacture of TNPP, manufacture of products and use of preparations containing TNPP.

Route-to-route extrapolation and calculation of internal doses

Inhalation and dermal route are the relevant occupational routes whereas all NOAELs are available by oral route only. Therefore route-to-route extrapolation has to be done and corrections should be made for differences in bioavailability as determined by percentages of absorption.

There are no data on the absorption of TNPP for the different routes of exposure. For oral absorption (starting route), a default value of 50 % may be chosen. For inhalation route, a default absorption of 50 % is proposed. For dermal absorption, a default value of 10 % can be used based on a $MW > 500$ and $\log P_{ow}$ higher than 4.

Internal doses are presented in table 4.4 via the different routes for each scenario. They are calculated using a human body weight of 70 kg and a ventilation rate of $10 \text{ m}^3/8 \text{ hours}$.

Table4-13: Calculated internal doses for workers

Scenario	Route of penetration				Combined routes
	Inhalation		Dermal		
	External exposure mg/m ³	Internal dose mg/kg/day	External exposure mg/day	Maximum internal dose mg/kg/day	Internal dose mg/kg/day
1 - TNPP manufacture	2.86	0.20	0 - 42	0.06	0.26
2 - Manufacture of products containing TNPP	8.58	0.61	42 - 420	0.60	1.21
3 - Use of preparations containing TNPP	5.72	0.41	0.42 - 4.2	0.006	0.42

For risk characterisation at the workplace, MOSs should normally be determined for route-specific as well as combined inhalation and dermal exposure. For simplification, only MOSs derived from combined exposure are presented.

4.1.3.2.1 Acute toxicity

Acute dermal toxicity was found to be very low ($\text{LD}_{50} > 2000 \text{ mg/kg}$). No data is available for acute inhalation toxicity but taking into account the very low acute toxicity by dermal and oral routes and that TNPP is a very slight to moderate irritant, inhalation acute toxicity is likely to be low as well. Acute toxicity is not considered of concern.

Conclusion (ii) for all scenarios

4.1.3.2.2 Irritation

TNPP is considered as a slight skin and eyes irritant and it may be presumed that it does not induce significant respiratory irritation. Therefore irritative effects are not considered of concern.

Conclusion (ii) for all scenarios

4.1.3.2.3 Sensitisation

One study conducted according to Buehler gave a negative response while a positive result was observed in a maximisation test. Thus TNPP is classified as a skin sensitizer.

No human data are available, however, according to the TNPP consortium, no case of sensitisation was observed at existing production sites. There are no data on respiratory sensitisation.

Exposure to TNPP during manufacture of the substance, manufacture of products and use of preparations may lead to concern. Risk reduction measures which should be applied as a result of its classification as the proper use of personal protective equipment can effectively reduce sensitisation at the work place. However, if protective equipment is not used properly and conscientiously and appropriate work procedures are not followed, it is likely that sensitisation might be induced in the worker. Although proper personal protection use and work procedure might be in use in most of the plants handling TNPP, there is no certainty that this is the situation of all plants in the EU. Conclusion iii is drawn in all worker scenarios. This conclusion is mitigated given the non dispersive use of the substance and the lack of reported case of sensitisation.

Conclusion (iii) for all scenarios

4.1.3.2.4 Repeated dose toxicity

Comparing the estimated combined internal exposure with the NOAELs of 167 mg/kg/day derived from a 2-year study in rats, the following MOSs can be calculated:

Table 4-14: MOSs for systemic effects by repeated exposure

Scenario	Internal Exposure mg/kg/day	Internal NOAEL mg/kg/day	MOS	Conclusion
1 - Manufacture	0.26	83.5	321	ii
2 - Manufacture of products	1.21	83.5	69	ii
3 - Use of preparations	0.42	83.5	199	ii

The effects observed at the LOAEL in the 2-year study in rats (500 mg/kg/day) are changes on growth and liver weight. The main toxic effect is a renal impact observed in a reproductive/developmental toxicity screening test in rats at 1000 mg/kg/day.

A minimal MOS of 50 can be derived from the following assessment factors:

- 10 for interspecies differences (default value)
- 5 for intraspecies differences (homogeneous population)
- 1 for type of the effect
- 1 for the confidence in the data base.

Compared to the minimal MOS, the MOSs are considered acceptable.

Conclusion (ii) for all scenarios

4.1.3.2.5 Mutagenicity

Available *in vitro* data do not reveal a genotoxic potential. Effects are not anticipated to occur.

Conclusion (ii) for all scenarios

4.1.3.2.6 Carcinogenicity

Data concerning carcinogenicity are not available. Based on results of mutagenicity testing, TNPP is not anticipated to be a genotoxic carcinogen. There is a low concern for carcinogenicity by a non-genotoxic mechanism too.

Conclusion (ii) for all scenarios

4.1.3.2.7 Toxicity to reproduction

Fertility and reproductive toxicity

Comparing the estimated combined internal exposure with the NOAEL of 200 mg/kg/day derived from a reproductive/developmental study in rats, the following MOSs can be calculated :

Table 4-15: MOSs for reproductive effects by repeated exposure

Scenario	Internal Exposure mg/kg/day	Internal NOAEL mg/kg/day	MOS	Conclusion
1 - Manufacture	0.26	100	385	ii
2 - Manufacture of products	1.21	100	87	ii
3 - Use of preparations	0.42	100	238	ii

The adverse effects observed at 1000 mg/kg/day in the reproductive/developmental study in rats are decrease of ovary weight in F0 females, a decrease of epididymes weight in F1 males and a slight litter size reduction. No other significant reproductive effects were observed.

A minimal MOS of 50 can be derived from the following assessment factors:

- 10 for interspecies differences (default value)
- 5 for intraspecies differences (homogeneous population)
- 1 for the type of the effect
- 1 for the confidence in the data base

Compared to the minimal MOS, the MOSs are considered acceptable.

Conclusion (ii) for all scenarios

Developmental effects

No indication of any developmental effect was observed up to the highest dose of 1000 mg/kg/day. Effects are not anticipated to occur.

Conclusion (ii) for all scenarios

4.1.3.3 Consumers

4.1.3.3.1 Introduction

Risk may occur by ingestion of food in contact with plastic containing TNPP. It is the only route of significant exposure for the consumer.

4.1.3.3.2 Risk characterisation due to migration from food contact materials

The total daily intake due to food-contact materials has been estimated to 0,0337 mg/day. For an adult with a bodyweight of 70 kg, the systemic dose resulting from this unique route of ingestion is 0,48 µg/kg/day. Systemic and reproductive effects are observed in animals with repeated dose. With the available NOAELs, the following MOSs can be calculated:

Table 4-16: MOS for systemic effects by repeated exposure

Scenario	Exposure µg/kg/day	NOAEL mg/kg/day	MOS	Concern for risks to human health	Conclusion
Food contact materials	0.48	167	350000	low	ii

Table 4-17: MOS for reproductive effects by repeated exposure

Scenario	Exposure µg/kg/day	NOAEL mg/kg/day	MOS	Concern for risks to human health	Conclusion
Food contact materials	0.48	200	420000	low	ii

4.1.3.3 Summary of risk characterisation for consumers

Repeated dose toxicity and reproductive effects are of low concern (**conclusion ii**).

4.1.3.4 Human exposed via the environment

This section was not provided as it will be updated in the next version of the environmental risk assessment.

4.1.3.4.1 Summary of risk characterisation for exposure via the environment

This section was not provided as it will be updated in the next version of the environmental risk assessment

DRAFT

4.2 HUMAN HEALTH (PHYSICO-CHEMICAL PROPERTIES)

Exposure assessment

The exposure assessment, to the extent it is related to physico-chemical properties, has already been discussed. No specific exposure information is available.

Effects assessment : Hazard identification and dose (concentration) - response (effect) assessment

Explosivity

TNPP has no explosive properties.

Flammability

TNPP has a very low degree of flammability (flash point : 207°C).

Oxidising potential

TNPP has no oxidising potential.

Risk characterisation

TNPP has neither explosive nor oxidising properties. The likelihood of an adverse effect deriving from flammability is very low.

Conclusion (ii) for all scenarios

DRAFT

5 RESULTS

5.1 ENVIRONMENT

To be updated

5.2 HEALTH

Risk assessment of human exposed via the environment was not discussed and will be updated following the update of environment risk assessment.

() (i) There is a need for further information and/or testing.

(X) (ii) There is at present no need for further information and/or testing and no need for risk reduction measures beyond those, which are being applied already.

This conclusion applies to the assessment of the risk to human health through consumer exposure.

(X) (iii) There is a need for specific measures to limit the risks.

This conclusion applies to the assessment of the risk to human health through worker exposure. It is reached because of concerns for sensitisation as a consequence of dermal exposure arising during manufacture of the substance, manufacture of products or use of preparations containing TNPP.

6 REFERENCES

Health part

- AWG (1998) Alkylphenol Work Group, Potential dietary exposure to p-nonylphenol from food contact use of tris(nonylphenyl)phosphite (TNPP)
- Chapin RE, Delaney J, Wang Y, Lanning L, Davis B, Collins B, Mintz N, Wolfe G, 1999. The effects of 4-nonylphenol in rats: a multigeneration reproduction study. *Toxicol Sci Nov* 52:1 80-91
- Ciba-Geigy, 1992. Acute dermal toxicity in the rat. Test n° 924059.
- Ciba-Geigy, 1983. Acute intraperitoneal LD₅₀ in rat. Project n° 831156.
- Ciba-Geigy, 1981. Skin irritation in the rabbit after single application of TK 10 417. Project n° 811807.
- Ciba-Geigy, 1981. Eye irritation in the rabbit after single application of TK 10 417. Project n° 811806
- Ciba-Geigy, 1992. Skin sensitisation test in the Guinea pig, Maximisation test. Test number 924058.
- Ciba-Geigy, 1990. Gene mutation test with Chinese hamster cells V79. test number 894549.
- Ciba-Geigy, 1990. Salmonella/Mammalian-microsome mutagenicity test. Test number 894547.
- Ciba-Geigy, 1990. Chromosome studies on Chinese hamster ovary cell line CCL 61 in vitro. Test number 894548.
- Cunny HC, Mayes BA, Rosica KA, Trutter JA, Van Miller JP, 1997. Subchronic toxicity (90-day) study with para-nonylphenol in rats. *Regul Toxicol Pharmacol Oct* 26:2 172-8
- DG Sanco - European Health & Consumer Protection Directorate-General. Provisional list of monomers and additives notified to European Commission as substances which may be used in the manufacture of plastics intended to come into contact with foodstuffs (updated to 25 July 2003). <http://europa.eu.int/comm/food/food/chemicalsafety/foodcontact/synoptic>
- European Union Risk Assessment Report on Nonylphenol ; Priority Setting, 2nd Priority List. R017_0104_env_hh
- Evans HJ, 1976. Cytological methods for detecting chemical mutagens. A. Hollaender (Ed.), *Chemical Mutagens, Principles and Methods for their Detection*, vol. 4. Plenum Press, New York, NY.
- FDA (2001) Safety Assessment of Di(2-ethylhexyl)phthalate (DEHP) Released from PVC Medical Devices. <http://www.fda.gov/cdrh/ost/dehp-pvc.pdf>
- FDA (2002) Preparation of Food Contact Notifications and Food Additive Petitions for Food Contact Substances: Chemistry Recommendations, FINAL GUIDANCE, April 2002
<http://www.cfsan.fda.gov/~lrd/foodadd.html>
- Food and Drug Research Laboratories, 1957. Toxicological studies with TNPP. Acute oral DL₅₀. Subacute Feeding Tests. Unpublished report n°72652
- Food and Drug Research Laboratories, 1961. Two-year feeding studies of TNPP in rats and dogs. Unpublished report n°79060
- Food and Drug Research Laboratories, 1971. The effects of Three Samples on the Survival Rate of Chick Embryos. Unpublished report n° 0582
- Green T, 2001. Nonylphenol : Bioavailability in the rat. CTL Report CTL/024139 ; Submitted to APERC October 24, 2001
- Gudi R and Brown C, 2001. *In Vitro* Mammalian Chromosome Aberration test. Unpublished report n° AA37MU.331.BTL, BioReliance, Rockville, MD 20850
- Hachiya N, 1987. Evaluation of Chemical Genotoxicity by a series of Short-term Tests. *Akita J. Med*, 14, 269-292

- Han XD, Tu ZG, Gong Y, Shen SN, Wang XY, Kang LN, Hou YY, Chen JX. The toxic effects of nonylphenol on the reproductive system of male rats. *Reprod. Toxicol.* 2004, 19, 215-221
- Hill Top Research, 1965. Acute oral administration of nonylated phenyl phosphite (samples #1 and #2), Mark 488, and Mark 492 to rats. Unpublished report
- Hossaini A, Dalgaard M, Vinggaard AM, Frandsen M, Larsen J-J. *In utero* reproductive study in rats exposed to nonylphenol. *Reprod. Toxicol.* 2001, 15, 537-543
- Howe *et al.* (2001) Potential dietary exposure to p-nonylphenol from food contact use of tris(nonylphenyl)phosphite (TNPP), *Food Additives and Contaminants*, 2001, Vol.18(11), 1021 – 1039
- INSPQ - Institut National de Santé Publique du Québec, Les phtalates : état des connaissances sur la toxicité et l'exposition de la population générale. (2004) <http://www.inspq.qc.ca/pdf/publications/252-Phtalates.pdf>
- Kahn KNM and Alden CL, 2002. Kidney. In. *Handbook of Toxicologic Pathology*, 2nd Ed. Academic Press, San Diego, CA, pp.255-336
- Majlathova L, Lekarskej F, Komenskeho M, 1981. Evaluation of aralkyl phenylphosphite antioxidants by an acute peroral experiment on mice and rats and by epicutaneous and conjonctival test on rabbits. *Bratisl. Lek. Listy* ; vol 76, ISS 3, 315-26
- Miller LC, Tainter ML, *Proc. Soc. Exptl. Biol. And Med.*, 57, 261 (1944)
- Montgomery CA and Seely JC, 1990. Kidney. In. *Pathology of the Fischer rat*. Academic Press, San Diego, CA, pp. 127-153
- Nagao T, Wada K, Marumo H, Yoshimura S, Ono H, 2001. Reproductive effects of nonylphenol in rats after gavage administration : a two-generation study. *Reprod Toxicol* May-Jun 15:3 293-315
- Odum J, Lefevre PA, Tinwell H, Van Miller JP, Joiner RL, Chapin RE, Wallis NT, Ashby J, 2002. Comparison of the developmental and reproductive toxicity of diethylstilbestrol administered to rats in utero, lactationally, preweaning, or postweaning. *Toxicol Sci* Jul 68:1 147-63
- Reimer GJ, 2001. Physical/Chemical Property of Tris-nonylphenol Phospite (TNPP). Hydrolysis as a function of pH (OECD 111). Unpublished report n° 1711234, Reimer Analytical and Associates Inc., Vancouver, BC, V6S 1B6
- San RHC and Clarke JJ, 2001. *In Vitro* Mammalian Cell Gene Mutation Test (L5178Y/TK[±] Mouse Lymphoma Assay). Unpublished report n° AA37MU.702.BTL, BioReliance, Rockville, MD 20850
- Tyl RW, Hamby BH, Myers CB, Marr MC, 2002. Reproductive/Developmental Toxicity Screening Test of Trisnonylphenyl Phosphite (TNPP) administred via Oral Gavage to CD[®] (Sprague-Dawley) Rats (Modified OECD 421). Unpublisbed report n° 65C-07895.300, Center for Life Sciences and Toxicology, Research triangle park, NC 27709-2194
- Tay CH, 2001a. Tris-nonylphenol (TNPP) : Acute dermal toxicity study - (OECD 402). Unpublished Report n° 01-4176-G1, Toxicon Corporation
- Tay CH, 2001b. Tris-nonylphenol (TNPP) : Acute dermal irritation/corrosion - (OECD 404). Unpublished Report n° 01-4176-G2, Toxicon Corporation
- Tay CH, 2001c. Tris-nonylphenol (TNPP) : Acute eye irritation/corrosion - (OECD 405). Unpublished Report n° 01-4176-G3, Toxicon Corporation
- Tay CH, 2001d. Tris-nonylphenol (TNPP) : Buehler sensitisation test - (OECD 406). Unpublished Report n° 01-4176-G4, Toxicon Corporation
- F.L van Velsen, J.A.M.A. Dormans, G.J.A. Speijers, and M.J. van Logten, 1980. Research on delayed neurotoxicity in chickens after treatment with organophosphites. RIVM report 78/80
- Wagner VO and Klug ML, 2001. Bacterial Reverse Mutation Assay. Unpublished report n° AA37MU.502.BTL, BioReliance, Rockville, MD 20850

Environment part

Building Research Establishment Ltd. (2001). <http://www.bre.co.uk/>

Chemical Information Services (2002). <http://www.chemicalinfo.com/>

Crompton (2003). WESTON TNPP Phosphite, product information, 08-01-2003.

E.C. (2003). Technical Guidance Document on Risk Assessment in support of Commission Directive 93/67/EEC on Risk Assessment for new notified substances, Commission Regulation (EC) N° 1488/94 on Risk Assessment for existing substances, Directive 98/8/EC of the European Parliament and of the Council concerning the placing of biocidal products on the market. Luxembourg, Office for Official Publications of the European Communities.

E.C. (2004a). EUSES 2.0, the European Union System for the Evaluation of Substances. National Institute of Public Health and the Environment (RIVM).

E.C. (2004b). EUSES, the European Union System for the Evaluation of Substances, Version 2.0. National Institute of Public Health and the Environment (RIVM).

OECD (2004). Emission scenario document on plastics additives, Organisation for Economic Co-operation and Development: 125.

Phoenix_Chemical_Laboratory (1997). Vapor pressure by isoteniscope (ASTM D2879). Chicago, May 19, 197, General Electric Specialty Chemicals: 1.

Pittsburgh_Testing_Laboratory (1978). Flash Point. Pittsburgh, July 12, 1978: 1.

Reimer&Associates (2001a). Physical/chemical property of tris-nonylphenyl phosphite (TNPP) [26523-78-4] - Solubility in Water (OECD 105). Pittsfield, August 2001, General Electric Company: 19.

Reimer&Associates (2001c). Physical/chemical property of tris-nonylphenyl phosphite (TNPP) [26523-78-4] - n-Octanol/Water partition coefficient. Pittsfield, August 2001, General Electric Company: 4.

Reimer&Associates (2001e). Physical/chemical property of tris-nonylphenyl phosphite (TNPP) [26523-78-4] - Melting (pour) point (OECD 102). Pittsfield, August 2001, General Electric Company: 10.

Reimer&Associates (2001f). Physical/chemical property of tris-nonylphenyl phosphite (TNPP) [26523-78-4] - Boiling temperature (OECD 103). Pittsfield, August 2001, General Electric Company: 10.

Staples, C. A. (2001). Fugacity modeling to estimate transport between environmental compartments for tris-nonylphenyl phosphite (TNPP) (CAS Reg. No.26523-78-4). Fairfax, November 2001, Assessment Technologies, Inc.: 5.

Ullmann (1985). Ullmann's encyclopedia of industrial chemistry, vol., VCH.

United States Testing Company, I. (1990). Report of Test. Hoboken, New Jersey, February 6, 1990, United States Testing Company, Chemical Services Division: 1.

US EPA and Syracuse Research Corporation (2001). EPI Suite, v.3.10, US EPA.

GLOSSARY

Standard Abbreviation	term	Explanation/Remarks and Alternative Abbreviation(s)
<i>Ann.</i>		Annex
AF		assessment factor
BCF		bioconcentration factor
bw		body weight / <i>Bw</i> , <i>b.w.</i>
°C		degrees Celsius (centigrade)
CAS		Chemical Abstract System
CEC		Commission of the European Communities
CEN		European Committee for Normalisation
CEPE		European Council of the Paint, Printing Ink and Artists' Colours Industry
d		day(s)
d.wt		dry weight / dw
DG		Directorate General
DT ₅₀		period required for 50 percent dissipation (define method of estimation)
DT _{50lab}		period required for 50 percent dissipation under laboratory conditions (define method of estimation)
DT ₉₀		period required for 90 percent dissipation (define method of estimation)
DT _{90field}		period required for 90 percent dissipation under field conditions (define method of estimation)
EC		European Communities
EC		European Commission
EC ₅₀		median effective concentration
EEC		European Economic Community
EINECS		European Inventory of Existing Commercial Chemical Substances
EU		European Union
EUSES		European Union System for the Evaluation of Substances
f _{oc}		Fraction of organic carbon
G		gram(s)

PNEC(s)	Predicted No Effect Concentration(s)
PNEC _{water}	Predicted No Effect Concentration in Water
(Q)SAR	Quantitative Structure Activity Relationship
STP	Sewage Treatment Plant
TGD	Technical Guidance Document ¹²
UV	Ultraviolet Region of Spectrum
UVCB	Unknown or Variable composition, Complex reaction products or Biological material
v/v	volume per volume ratio
w/w	weight per weight ratio
w	gram weight
GLP	Good Laboratory Practice
h	hour(s)
ha	Hectares / <i>h</i>
HPLC	High Pressure Liquid Chromatography
IARC	International Agency for Research on Cancer
C ₅₀	median immobilisation concentration or median inhibitory concentration 1 / <i>explained by a footnote if necessary</i>
ISO	International Standards Organisation
IUPAC	International Union for Pure Applied Chemistry
kg	kilogram(s)
kPa	kilo Pascals
K _{oc}	organic carbon adsorption coefficient
K _{ow}	octanol-water partition coefficient
K _p	Solids water partition coefficient
l	litre(s)
log	logarithm to the basis 10
L(E)C ₅₀	Lethal Concentration, Median
LEV	Local Exhaust Ventilation
m	Meter
µg	microgram(s)

¹² Commission of the European Communities, 1996. Technical Guidance Documents in Support of the Commission Directive 93/67/EEC on risk assessment for new substances and the Commission Regulation (EC) No 1488/94 on risk assessment for existing substances. Commission of the European Communities, Brussels, Belgium. ISBN 92-827-801[1234]

mg	milligram(s)
MAC	Maximum Accessibility Concentration
MOS	Margins Of Safety
NOAEL	No Observed Adverse Effect Level
NOEC	No Observed Effect Concentration
NOEL	No Observed Effect Level
OEL	Occupational Exposure Limit
OECD	Organisation for Economic Co-operation and Development
OJ	Official Journal
pH	potential hydrogen <i>-logarithm</i> (to the base 10) of the hydrogen ion concentration {H ⁺ }
pKa	<i>-logarithm</i> (to the base 10) of the acid dissociation constant
pKb	<i>-logarithm</i> (to the base 10) of the base dissociation constant
Pa	Pascal unit(s)
PEC	Predicted Environmental Concentration
STP	Sewage Treatment Plant
WWTP	Waste Water Treatment Plant

ANNEX 1

European Commission

**EUR .:[[click here to insert EUR No.](#)] - European Union Risk Assessment Report
[[click here to insert SUBSTANCE NAME, and volume no.](#)]**

Editors: B.G. Hansen, S.J. Munn, S. Pakalin, C.J.A. Heidorn, R. Allanou, S. Scheer, G. Pellegrini, S. Vegro, J.De Bruijn, M.Luotamo, K. Vormann, H. Loonen, F. Berthault, A. Naughton, V. Anfossi, L. Praderio. ([keep this updated](#))

Luxembourg: Office for Official Publications of the European Communities

2000 – VIII pp. .:[[click here to insert page count, number of pages.](#)]
pp. – 17.0 x 24.0 cm

Environment and quality of life series

ISBN .:[[click here to insert ISBN No.](#)]

Price (excluding VAT) in Luxembourg: EUR .:[[click here to insert price.](#)]

The report provides the comprehensive risk assessment of the substance Tris(nonylphenyl) phosphate (TNPP). It has been prepared by France in the frame of Council Regulation (EEC) No. 793/93 on the evaluation and control of the risks of existing substances, following the principles for assessment of the risks to humans and the environment, laid down in Commission Regulation (EC) No. 1488/94.

Part II – Human Health

The HH part of the evaluation considers the emissions and the resulting exposure to human populations in all life cycle steps. The scenarios for occupational exposure and consumer exposure have been examined and the possible risks have been identified, while the human exposure via the environment has not been assessed

There is only concern for workers because of sensitisation problems, but not for consumers.

The conclusions of this report will lead to risk reduction measures proposed by the Commission's committee on risk reduction strategies set up in support of Council Regulation (EEC) N. 793/93.